

# In Lesson 1

- **Functional genomics is a field of molecular biology based on genome-wide sequencing data.**
- **Genome-wide sequencing data describe genomic regulatory regions that control gene expression**
- **Gene expression dysregulation may be linked to the disease**
- **Understanding molecular mechanisms of disease outcome opens the way to discovery drug and identify biomarkers**



# How SNPs play a **FUNCTIONAL** role in disease:

## Impact on transcription

- **Changing consensus sequences for transcription factors binding sites**
- **Changing interaction between for transcription factors**
- **Changing epigenetic profiling of specific genomic regions**
- **Changing long range interaction between two genomic regions**



# In this Lesson

- **Enhancer Overview**
- **Genomic regulatory network to define cell identity**
- **Genetic variations meaning in cell identity**

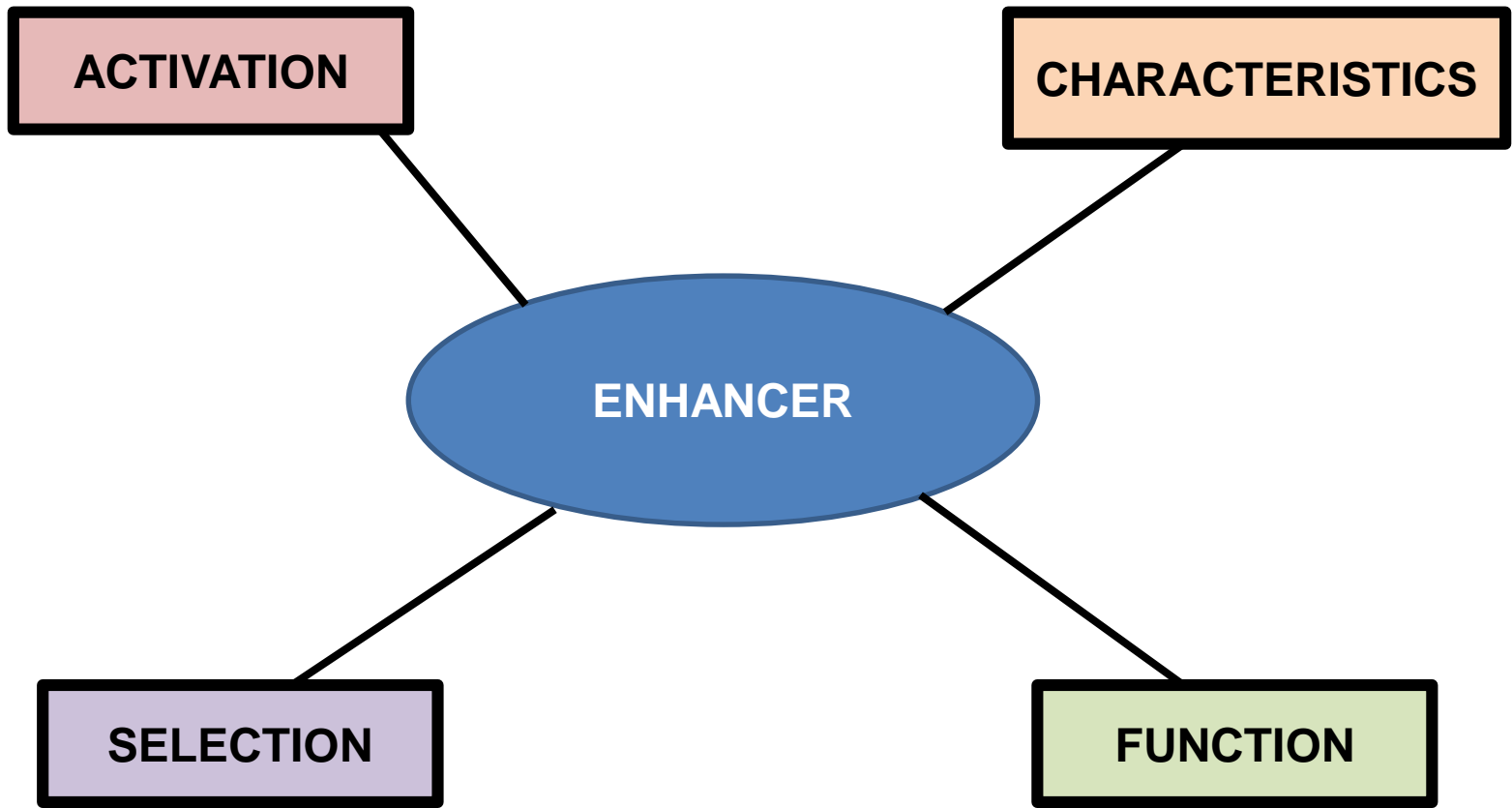


# The selection and function of cell type-specific enhancers

*Sven Heinz<sup>1</sup>, Casey E. Romanoski<sup>2</sup>, Christopher Benner<sup>1</sup> and Christopher K. Glass<sup>2,3</sup>*

**Abstract** | The human body contains several hundred cell types, all of which share the same genome. In metazoans, much of the regulatory code that drives cell type-specific gene expression is located in distal elements called enhancers. Although mammalian genomes contain millions of potential enhancers, only a small subset of them is active in a given cell type. Cell type-specific enhancer selection involves the binding of lineage-determining transcription factors that prime enhancers. Signal-dependent transcription factors bind to primed enhancers, which enables these broadly expressed factors to regulate gene expression in a cell type-specific manner. The expression of genes that specify cell type identity and function is associated with densely spaced clusters of active enhancers known as super-enhancers. The functions of enhancers and super-enhancers are influenced by, and affect, higher-order genomic organization.



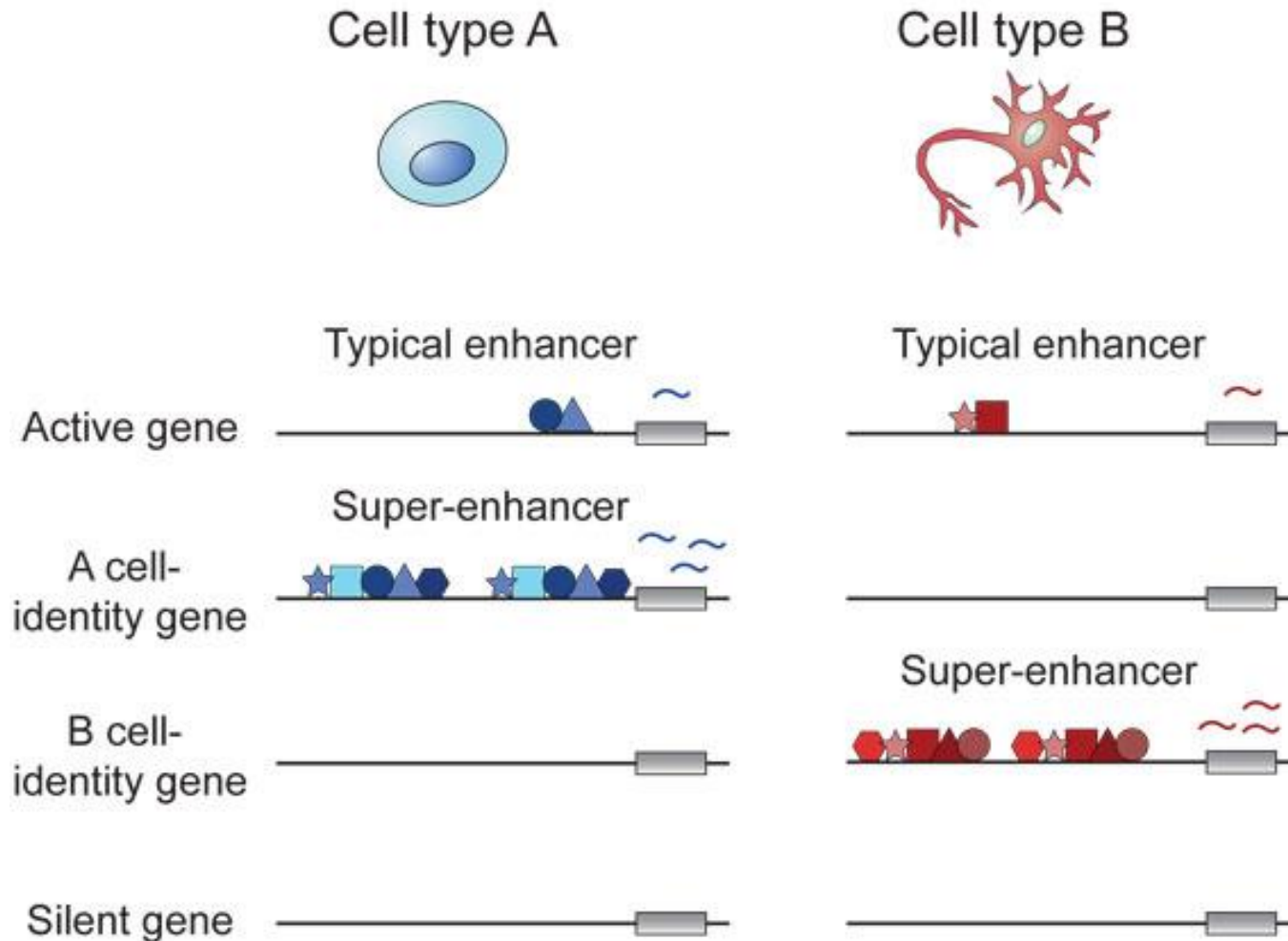


# Enhancer Characteristics

- **Enhancers are regulatory elements in proximity of genes**
- **Each cell has a set of enhancers**
- **Enhancers have motifs for sequence-specific transcription factors**
- **Enhancers are marked with epigenetic modifications**
- **Enhancers are in different states of activation**

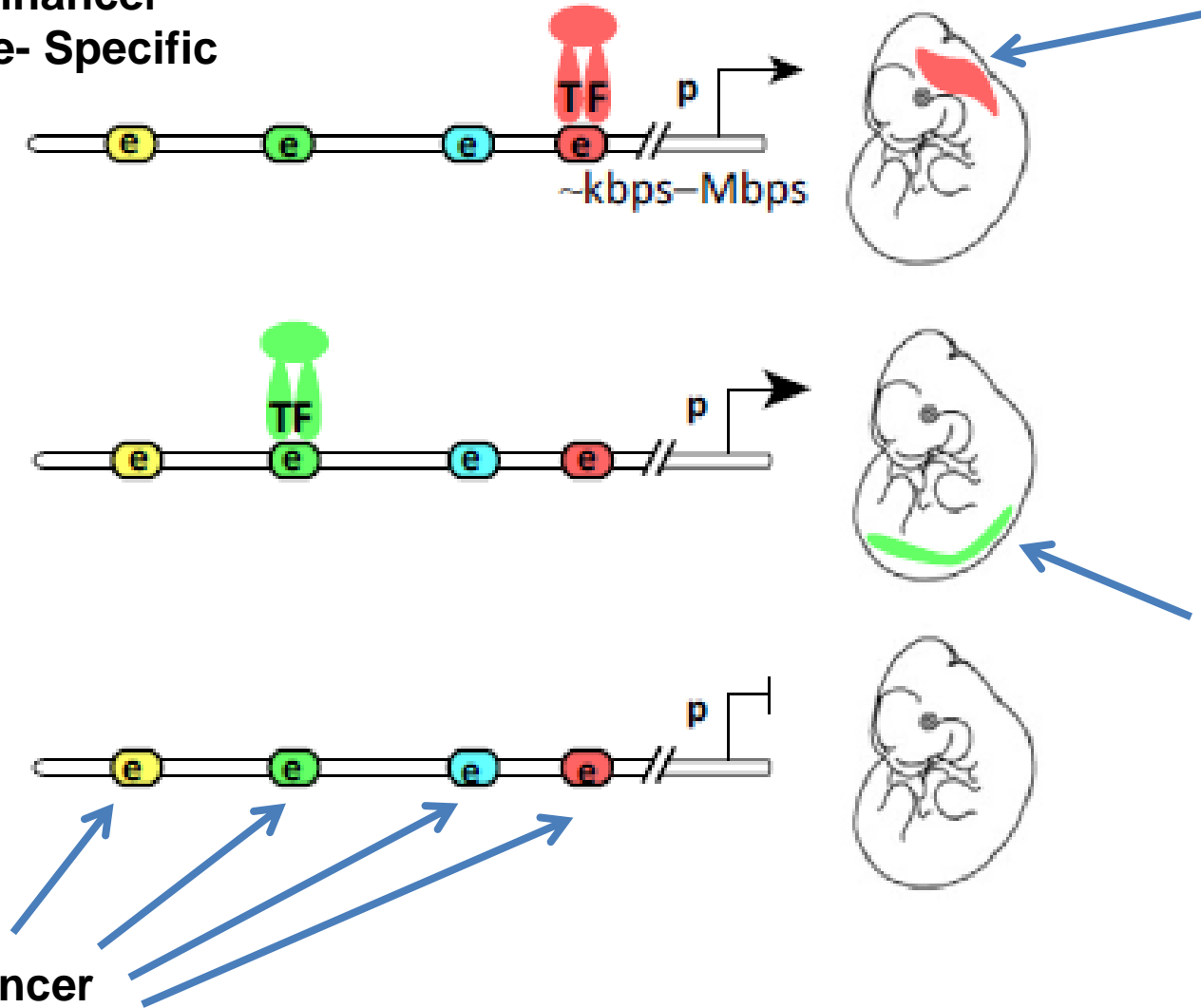


# MUCH OF THE **REGULATORY CODE** THAT DRIVES CELL-TYPE-SPECIFIC GENE EXPRESSION IS LOCATED IN DISTAL ELEMENTS CALLED **ENHANCERS**



# CELL TYPE USE A SMALL SUBSET OF MILLIONS OF POTENTIAL ENHANCERS

Active Enhancer  
in Cell-Type- Specific



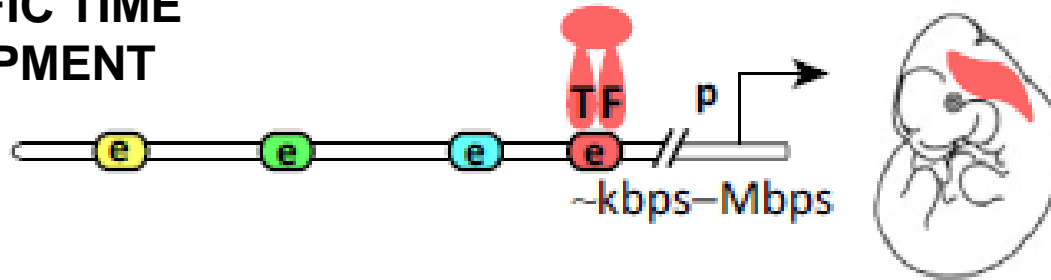
Enhancers in tissue/cell-specific gene expression



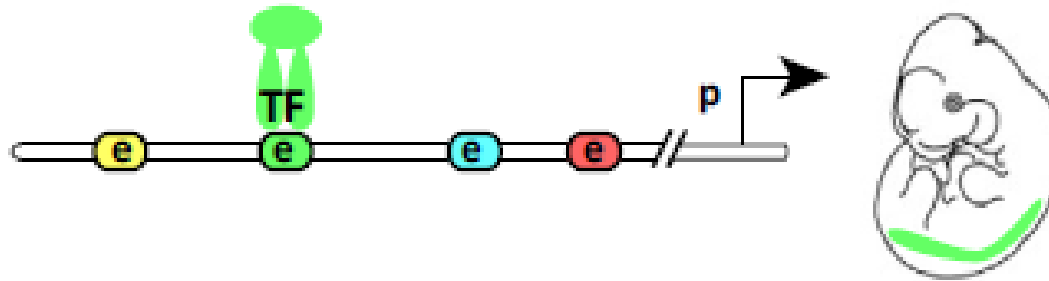


# CELL TYPE USE A SMALL SUBSET OF MILLIONS OF POTENTIAL ENHANCERS

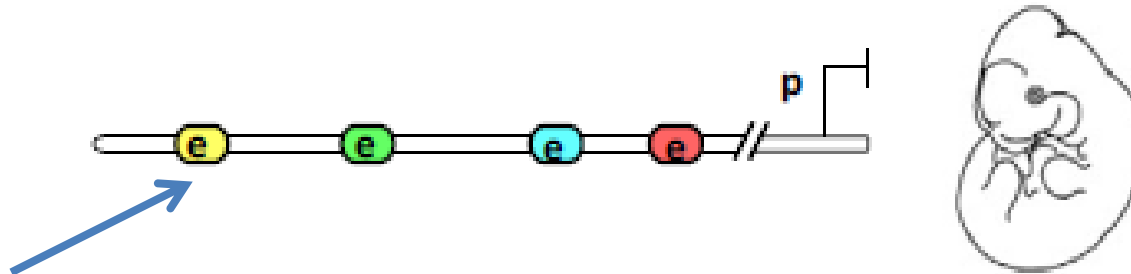
Active Enhancer  
during SPECIFIC TIME  
OF DEVELOPMENT



Stage 1



Stage 2

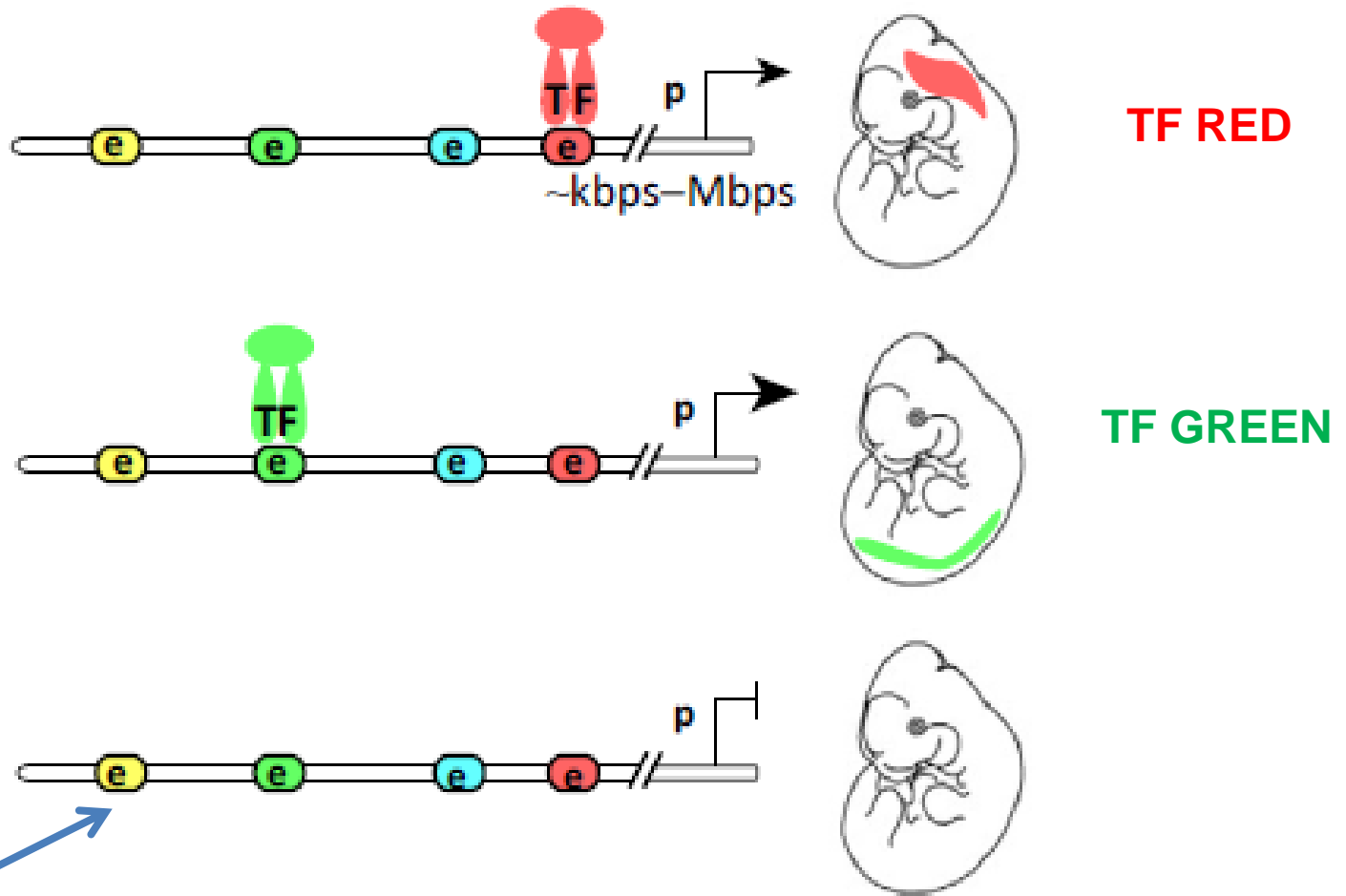


Potential enhancers

Enhancers in tissue/cell-specific gene expression



# LINEAGE-DETERMINING TRANSCRIPTION FACTORS BIND AT CELL-TYPE SPECIFIC ENHANCERS

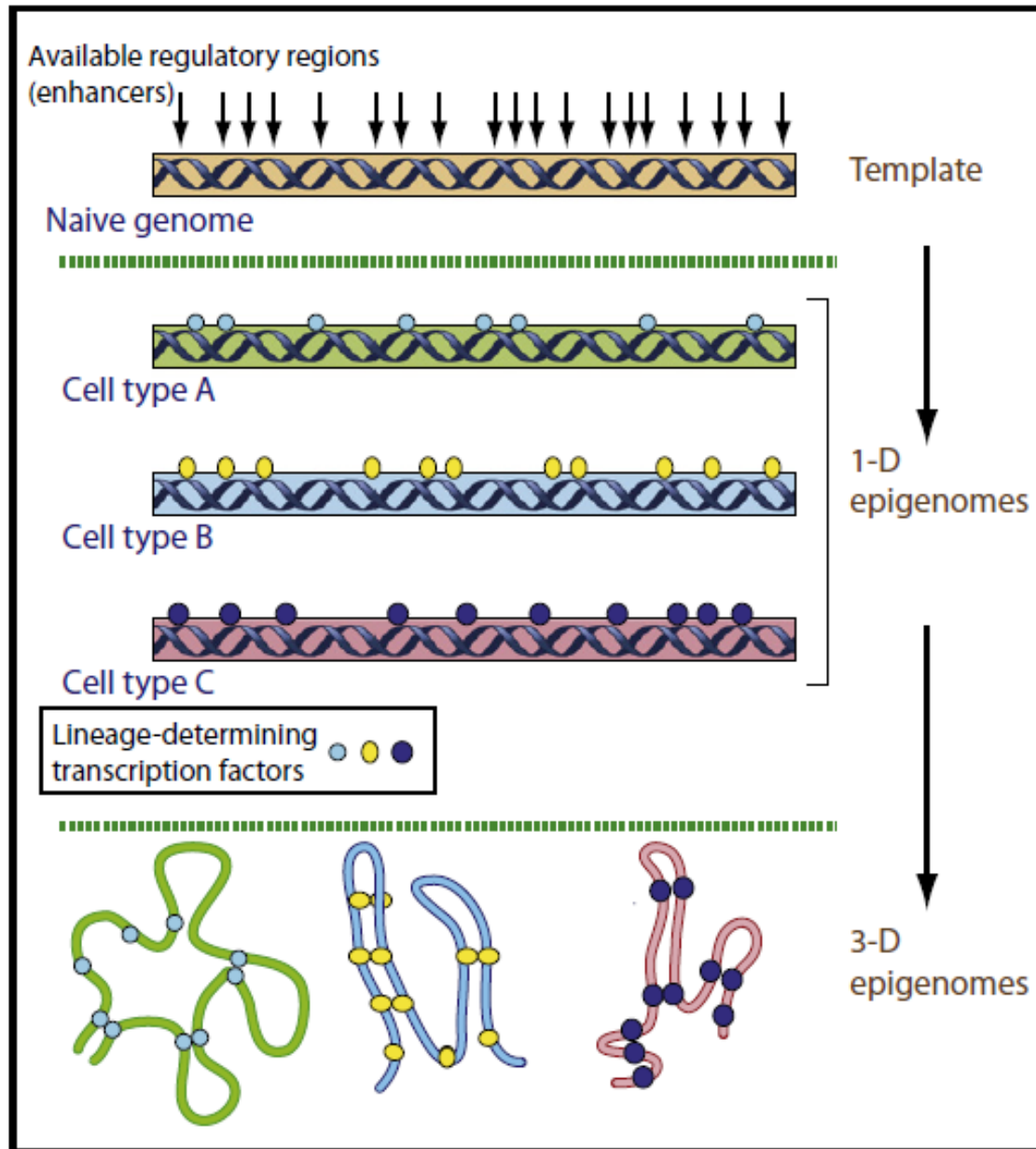


Potential enhancers

Enhancers in tissue/cell-specific gene expression

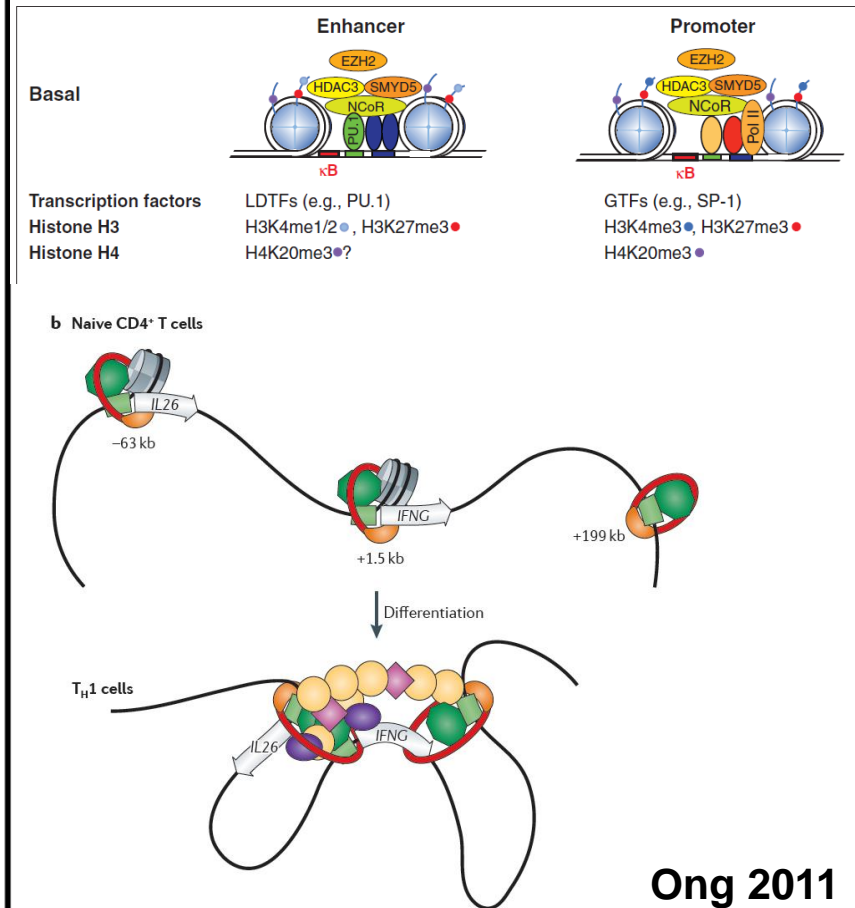
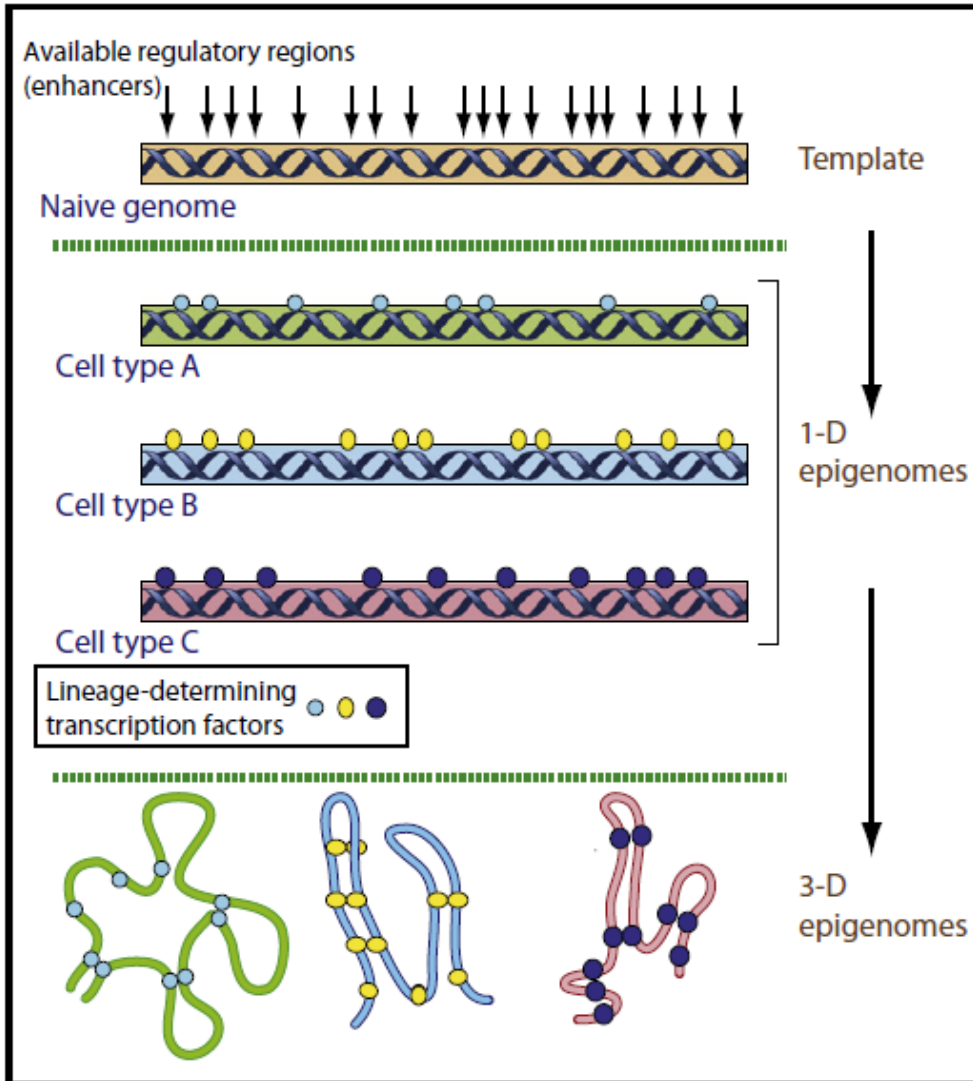


# EACH CELL HAS ACTIVE ENHANCERS



# Maintaining Cell Identity through Global Control of Genomic Organization

Gioacchino Natoli<sup>1,\*</sup>



Ong 2011

# TRANSCRIPTION FACTOR BINDS SPECIFIC CONSENSUS SEQUENCE IN ACTIVE ENHANCER

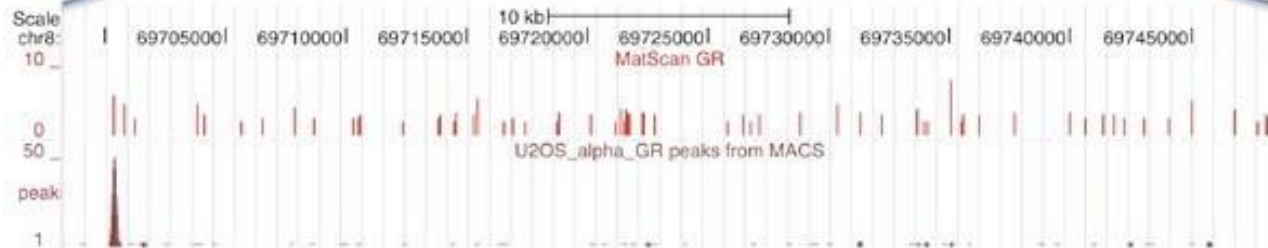


GR binding site motif found in approx. every 1000bp in genome



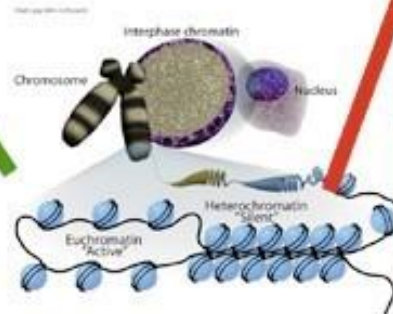
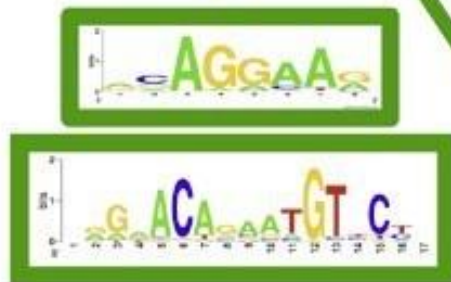
Red line marks  
GR binding motif:

Peak reflects actual  
GR Binding event:



Genetic and epigenetic  
inputs directing binding

Genetic and epigenetic inputs  
that restrict binding



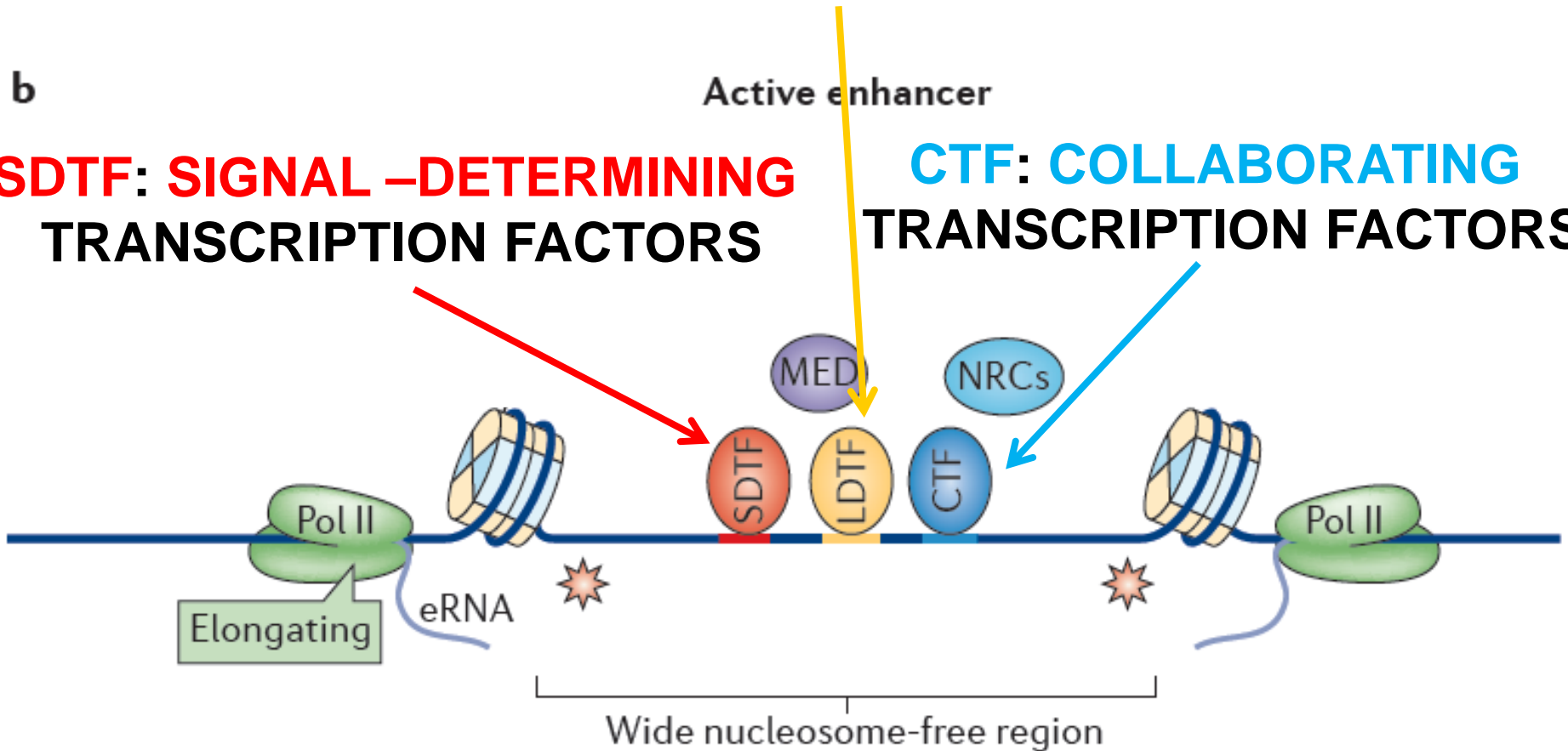
# TRANSCRIPTION FACTORS THAT BIND ENHANCERS

## LDTF: LINEAGE – DETERMINING TRANSCRIPTION FACTORS

b

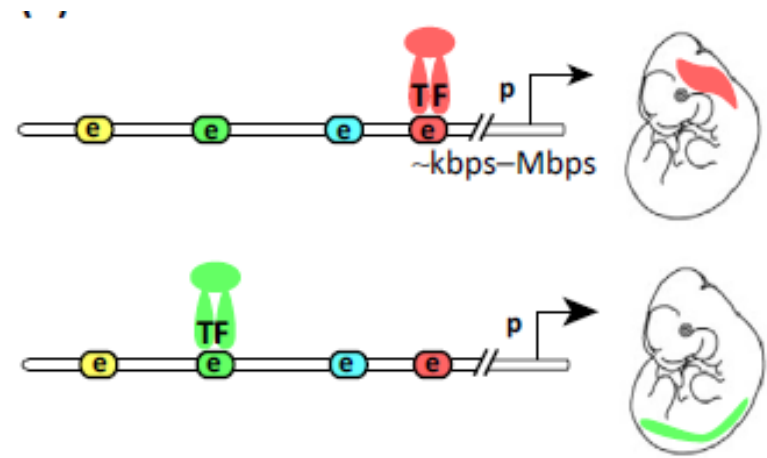
**SDTF: SIGNAL – DETERMINING  
TRANSCRIPTION FACTORS**

**CTF: COLLABORATING  
TRANSCRIPTION FACTORS**

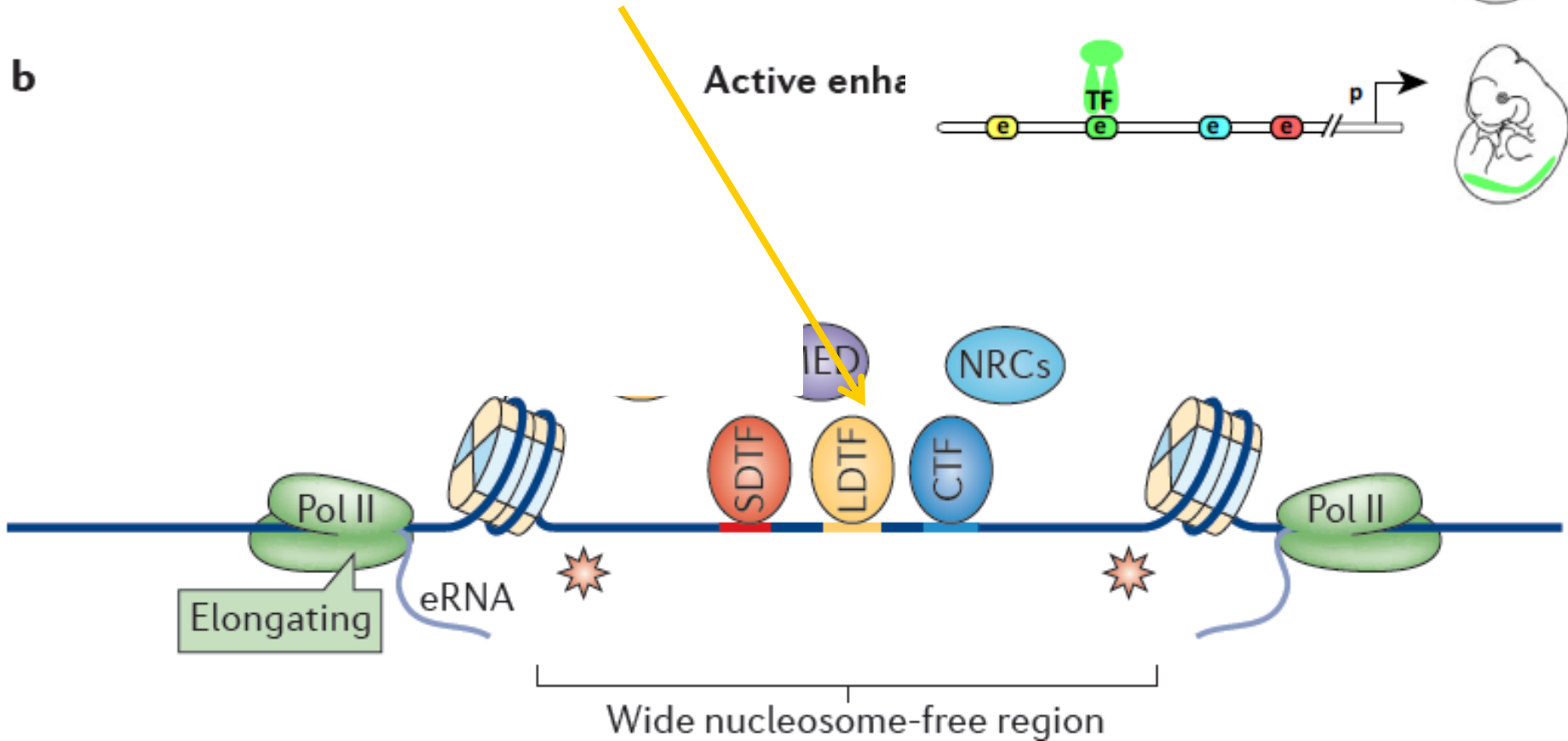


# TRANSCRIPTION FACTORS THAT BIND ENHANCERS

## LDTF: LINEAGE –DETERMINING TRANSCRIPTION FACTORS



b



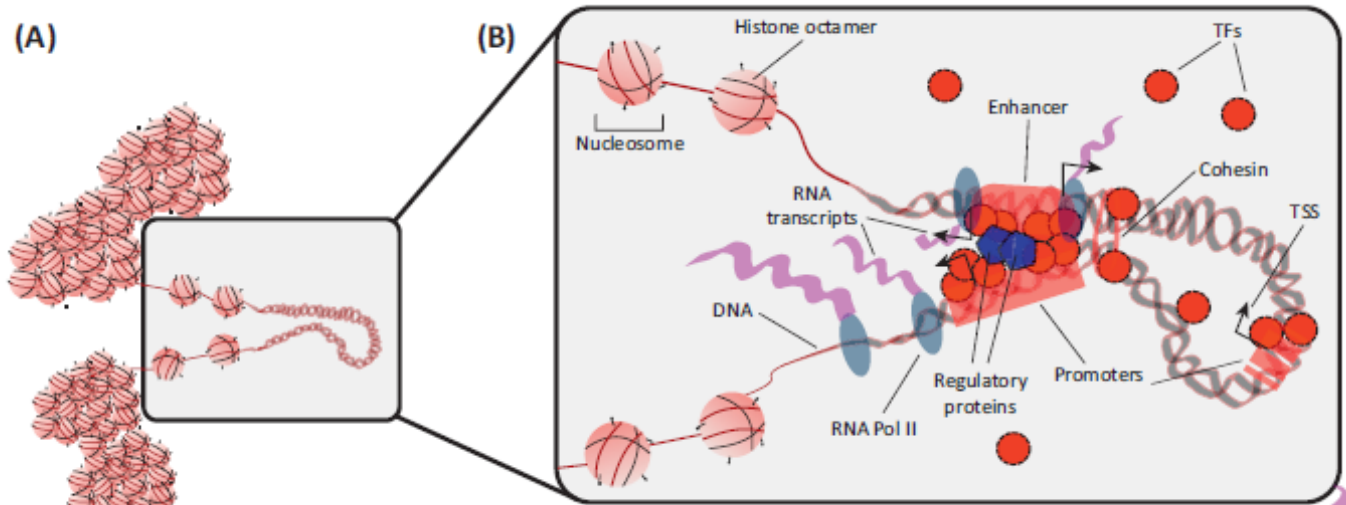
# Enhancer Activation

- **Transcription Factors bind specific genomic regions and allow access to other proteins remodelling chromatin**
- **Differentiation states and external stimuli induce enhancers activation**



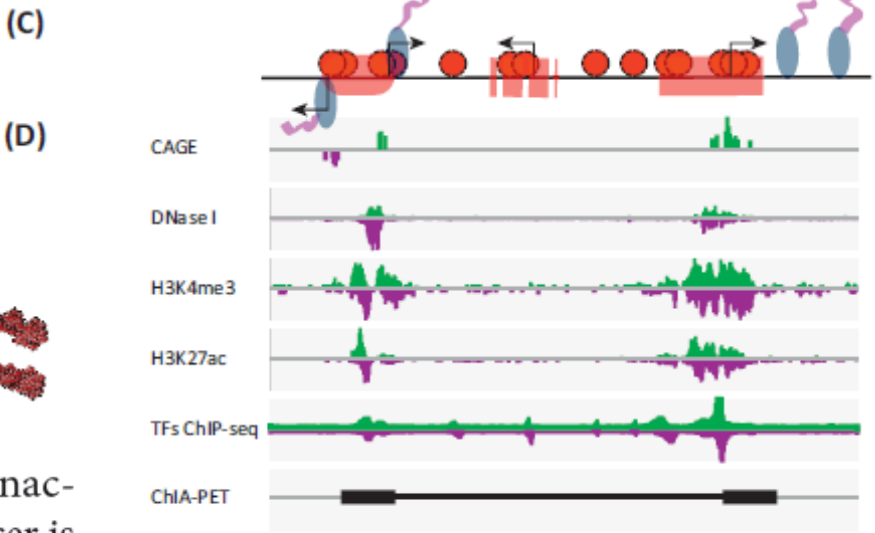


# ACTIVE ENHANCER



# INACTIVE ENHANCER

Enhancer states can broadly be classified as inactive, primed, poised or active<sup>22</sup>. An inactive enhancer is essentially buried in compact chromatin and is devoid of transcription factor binding and histone modifications.



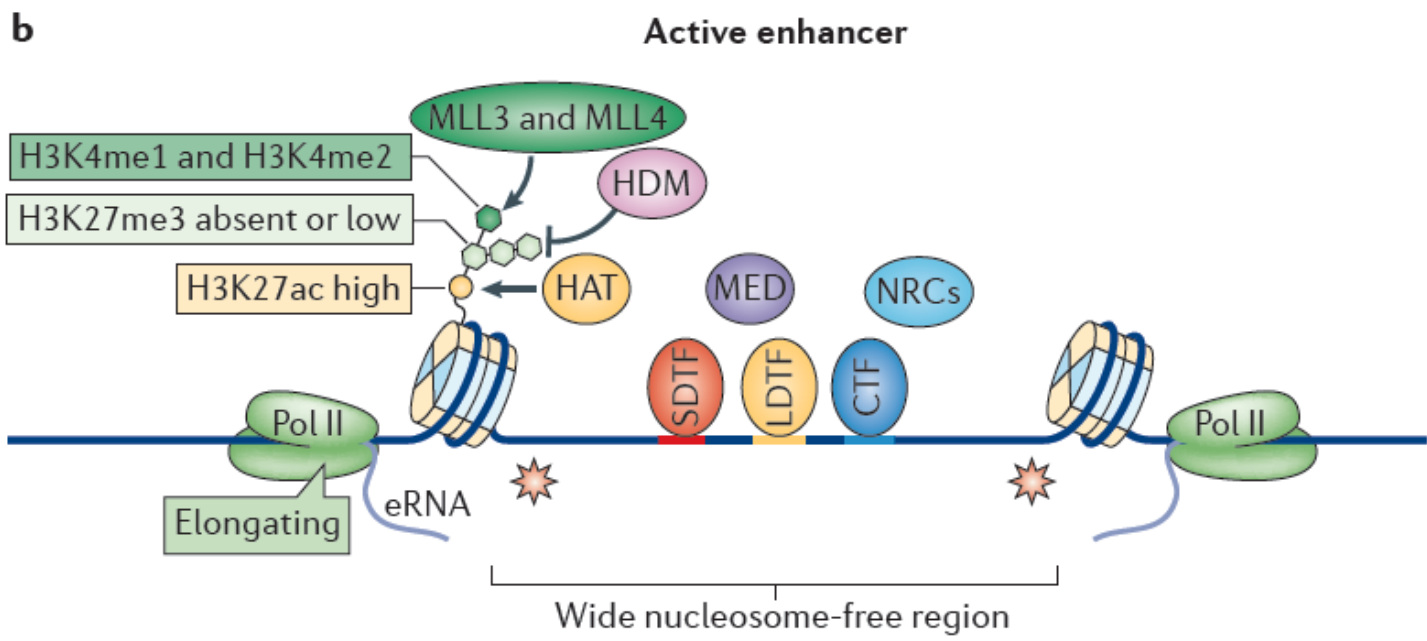
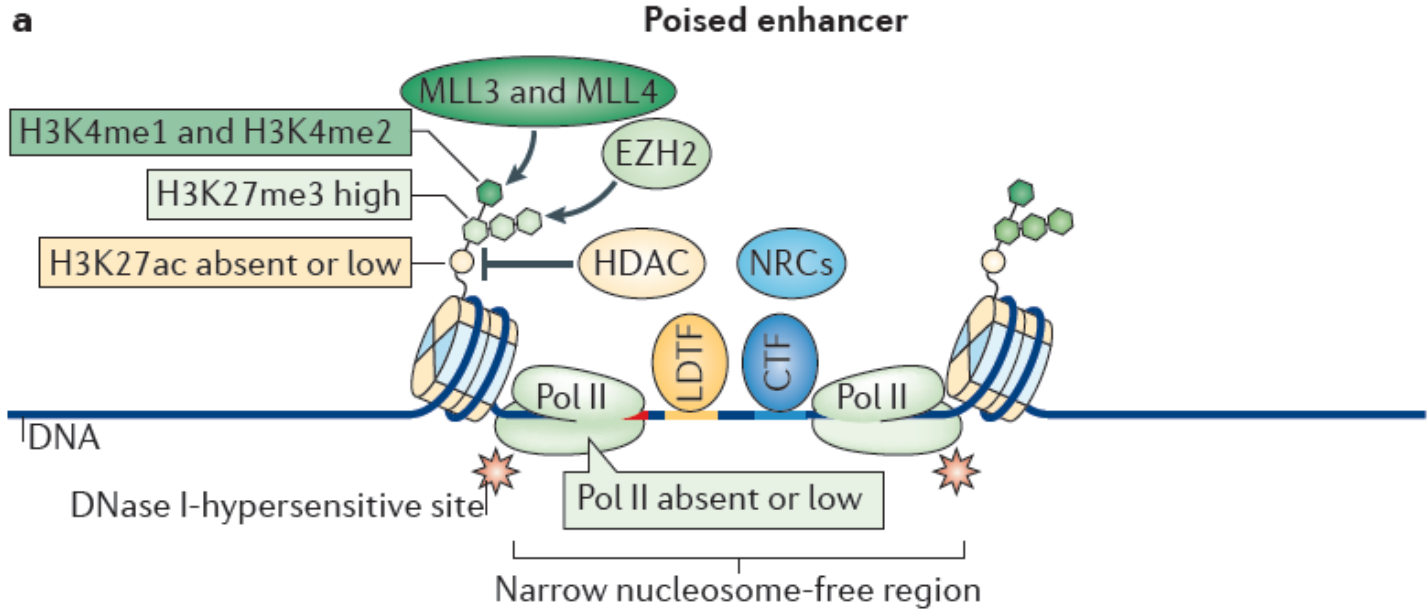
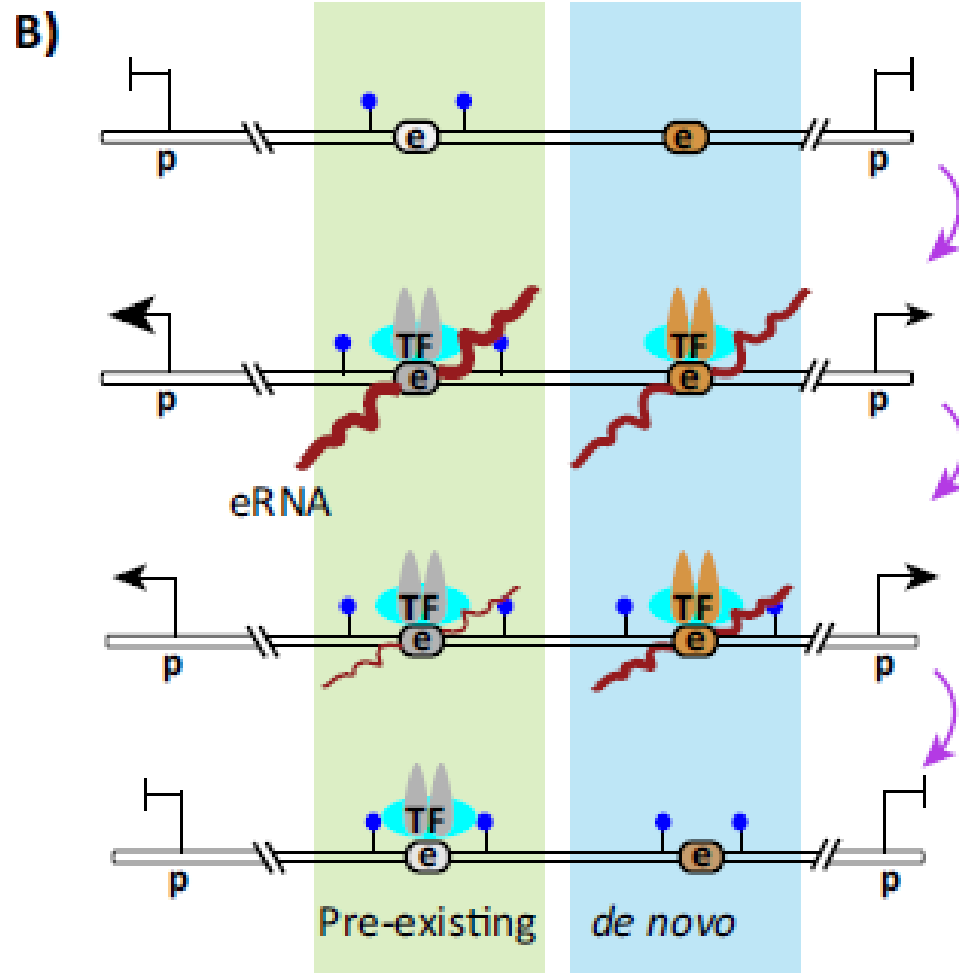


Figure 1 | **The anatomies of poised and active enhancers.** The characteristic features

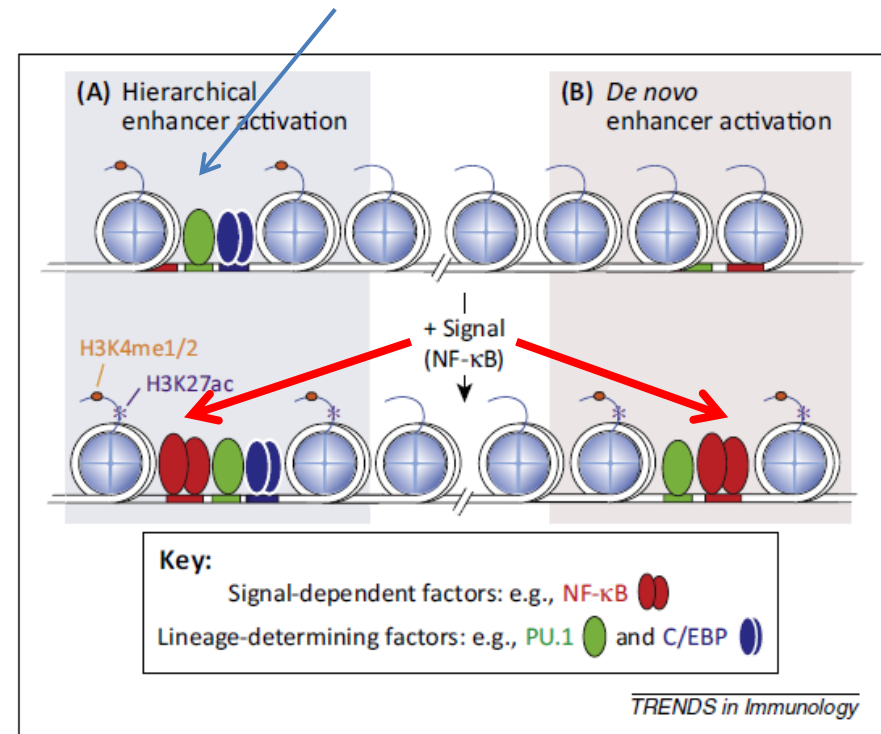


# LDTF: LINEAGE – DETERMINING TRANSCRIPTION FACTORS



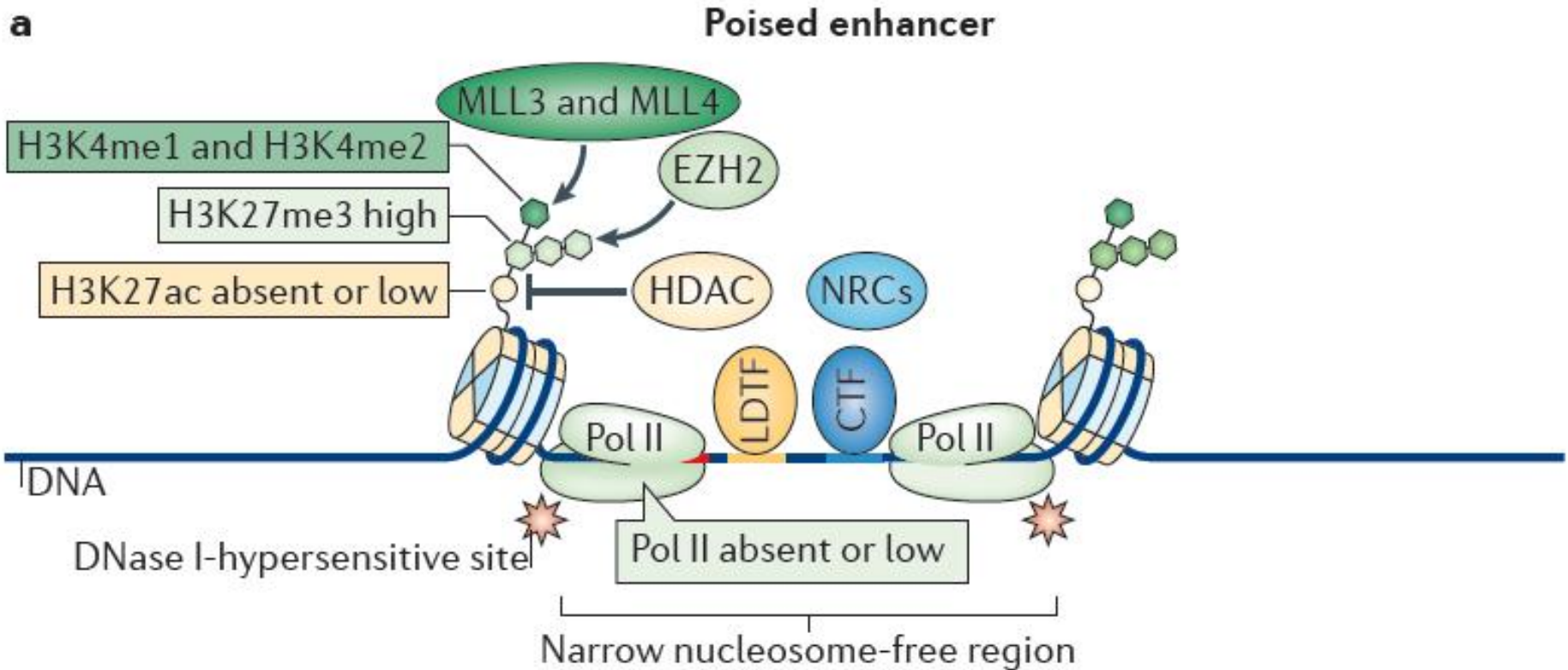
† Histone methylation ( $H3K4me^{1/2}$ )

Enhancers in stimulus-induced gene activation



**Figure 3.** Chromatin transitions to active enhancers involve interactions between cell lineage-determining transcription factors and signal-dependent factors. **(A)** Enhancers primed by lineage-determining factors frequently require signal-dependent transcription factor binding to gain H3K27ac and become active. **(B)** Active enhancers can also be selected by interactions between signal-dependent factors and lineage-determining factors. Abbreviations: C/EBP, CCAAT/enhancer binding protein; NF-κB, nuclear factor-κB; PU.1, transcription factor originally named spleen focus forming virus (SFFV) proviral integration oncogene.

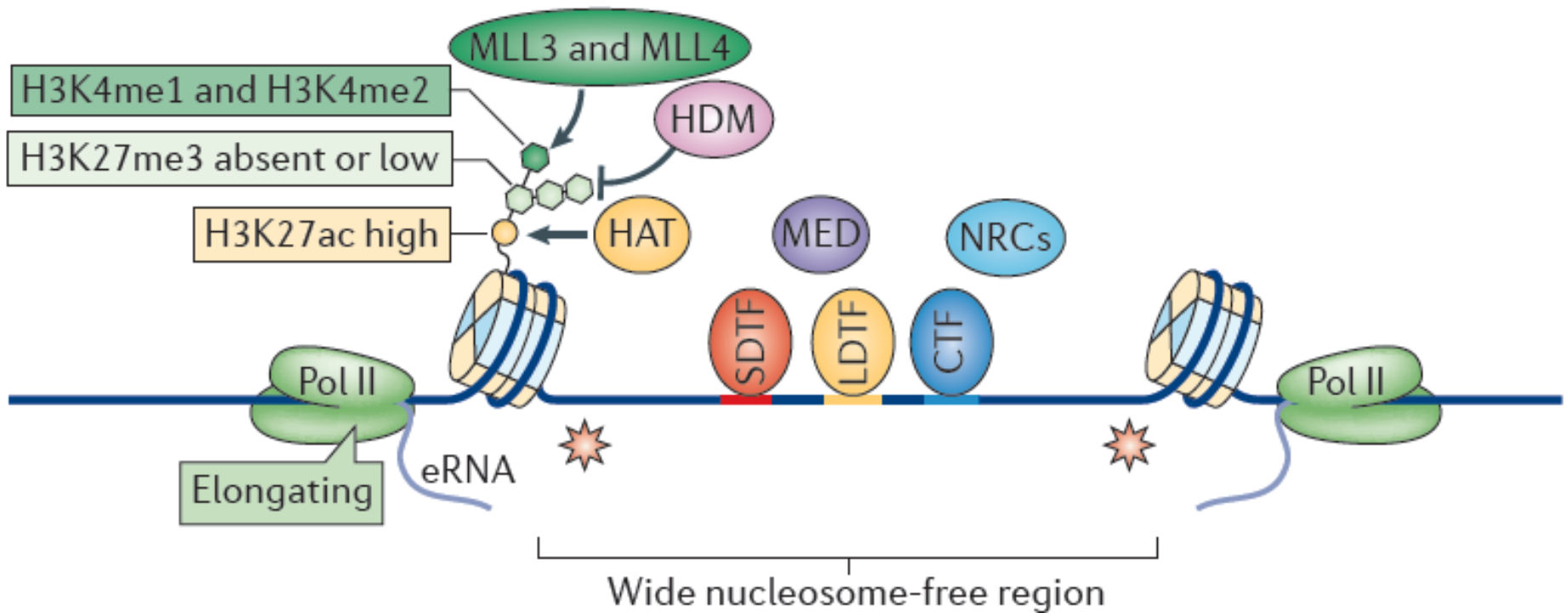
H3K4me1, H3K4me2, **lack histone acetylation and Pol II,**  
**high H3K27me3**  
mark **POISED ENHANCERS**



H3K4me1, H3K4me2, **high H3K27Ac**, lack H3K27me3,  
**presence of Pol II and RNA transcript**  
mark **ACTIVE ENHANCERS**

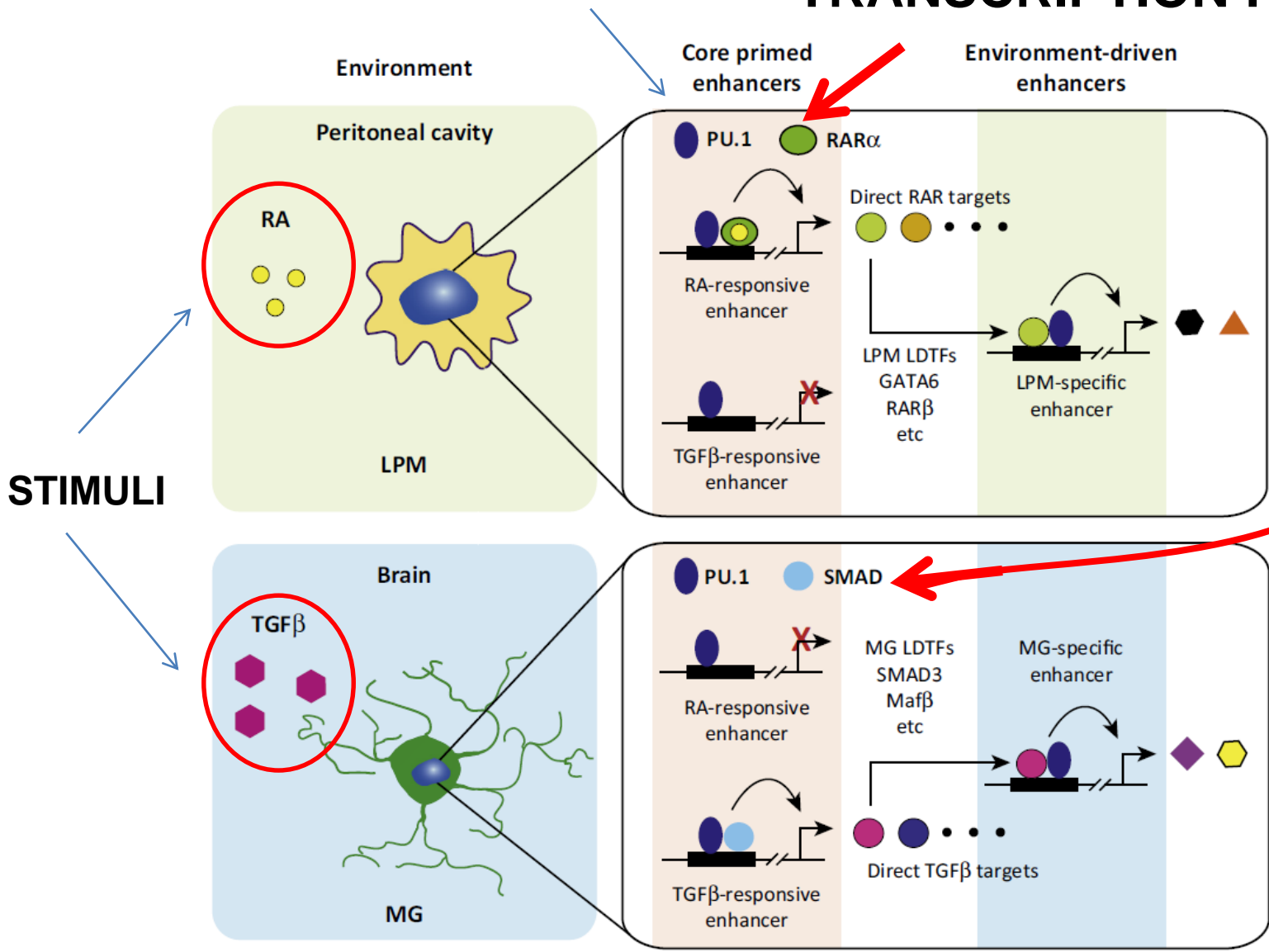
b

Active enhancer



# LDTF: LINEAGE –DETERMINING TRANSCRIPTION FACTORS

# SDTF: SIGNAL –DETERMINING TRANSCRIPTION FACTORS



# Enhancer Selection

- **The role of lineage-determining transcription factors.**
- **The role of signal-dependent transcription factors.**

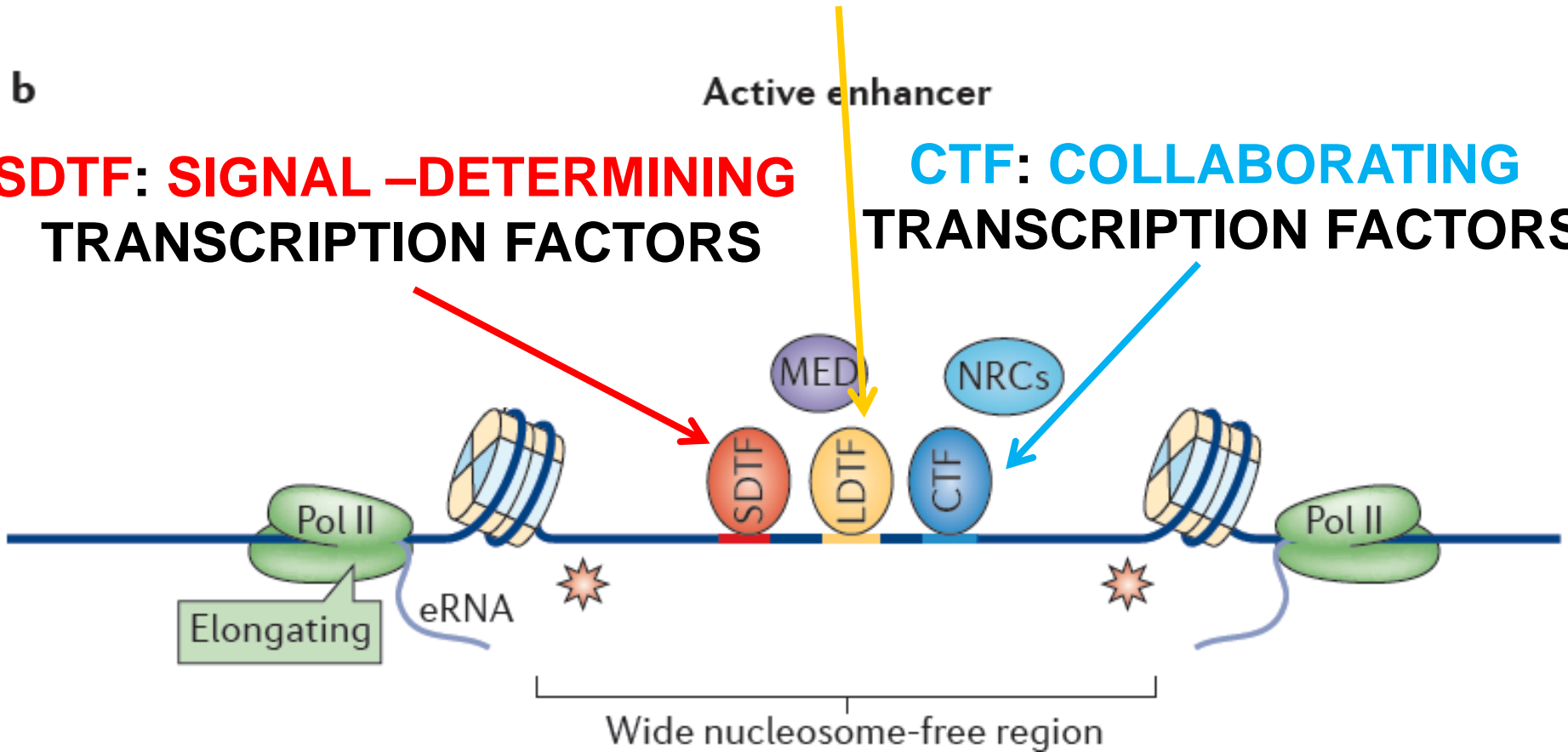


# LDTF: LINEAGE – DETERMINING TRANSCRIPTION FACTORS

b

**SDTF: SIGNAL – DETERMINING  
TRANSCRIPTION FACTORS**

**CTF: COLLABORATING  
TRANSCRIPTION FACTORS**

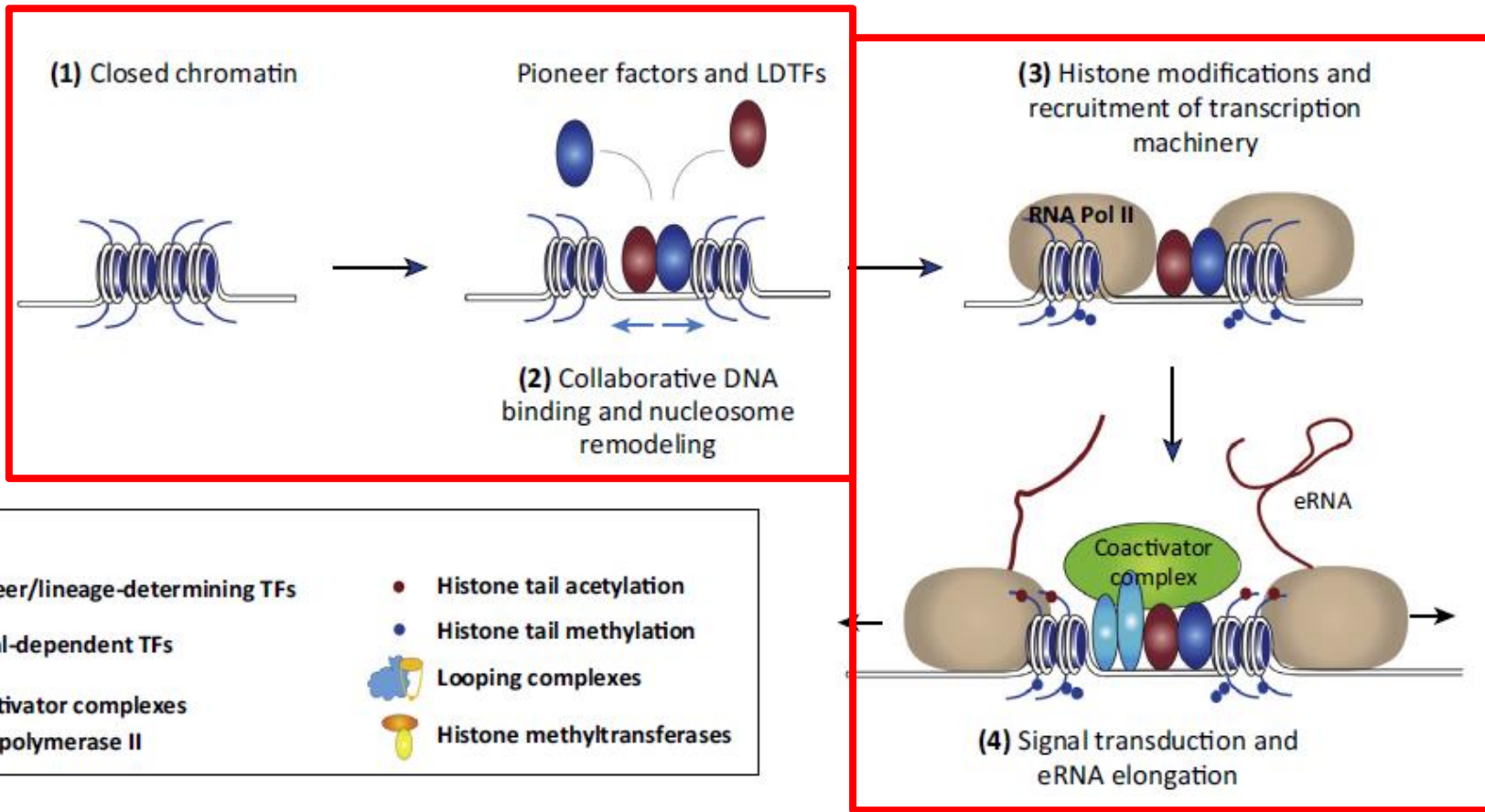




# Pioneer Factors and Lineage-determining Transcription Factors leads to nucleosome remodeling and increased chromatin accessibility



(A)



Pioneer Factors and Lineage-determining Transcription Factors leads to histone modifications and basal transcription machinery



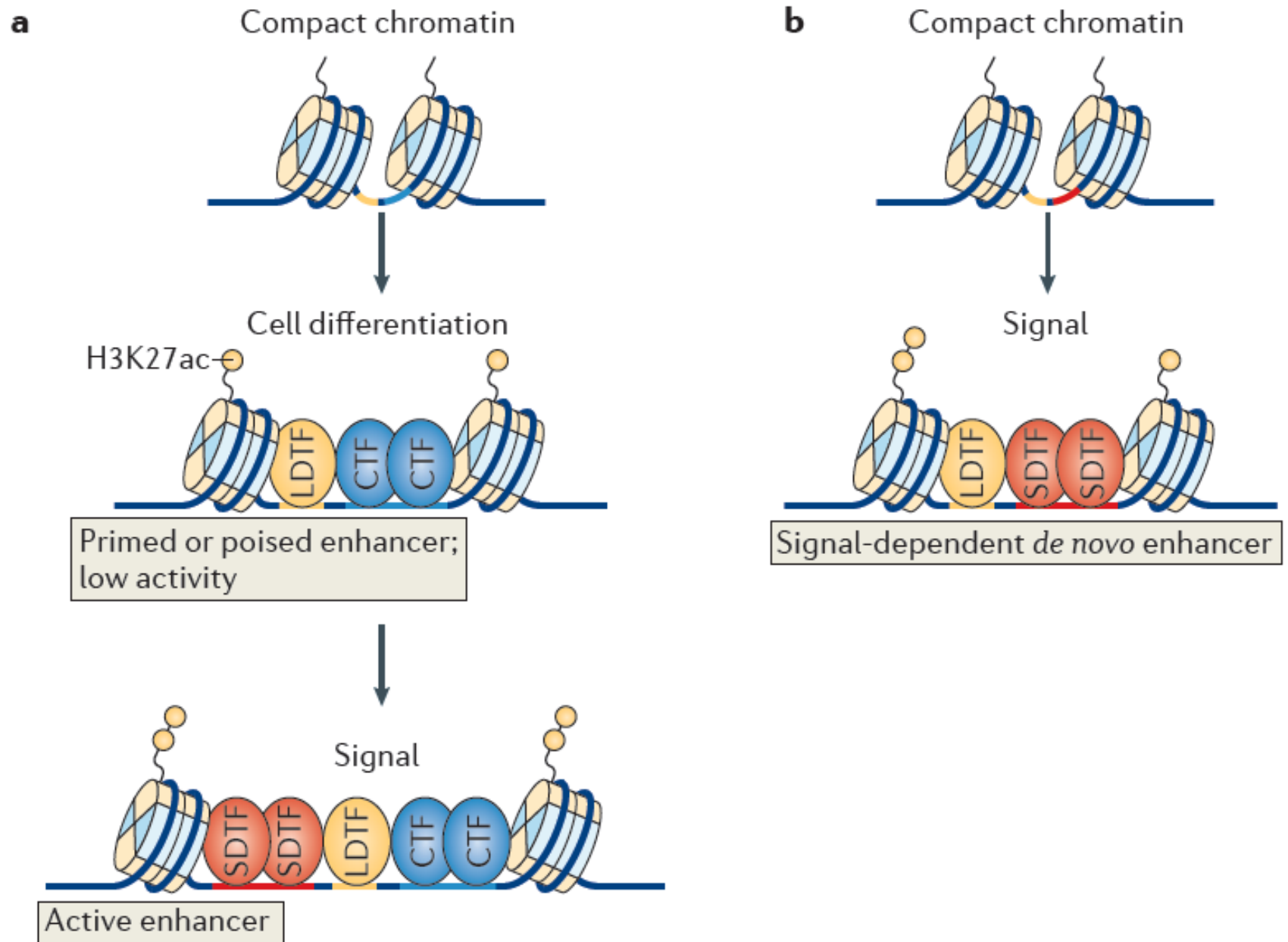
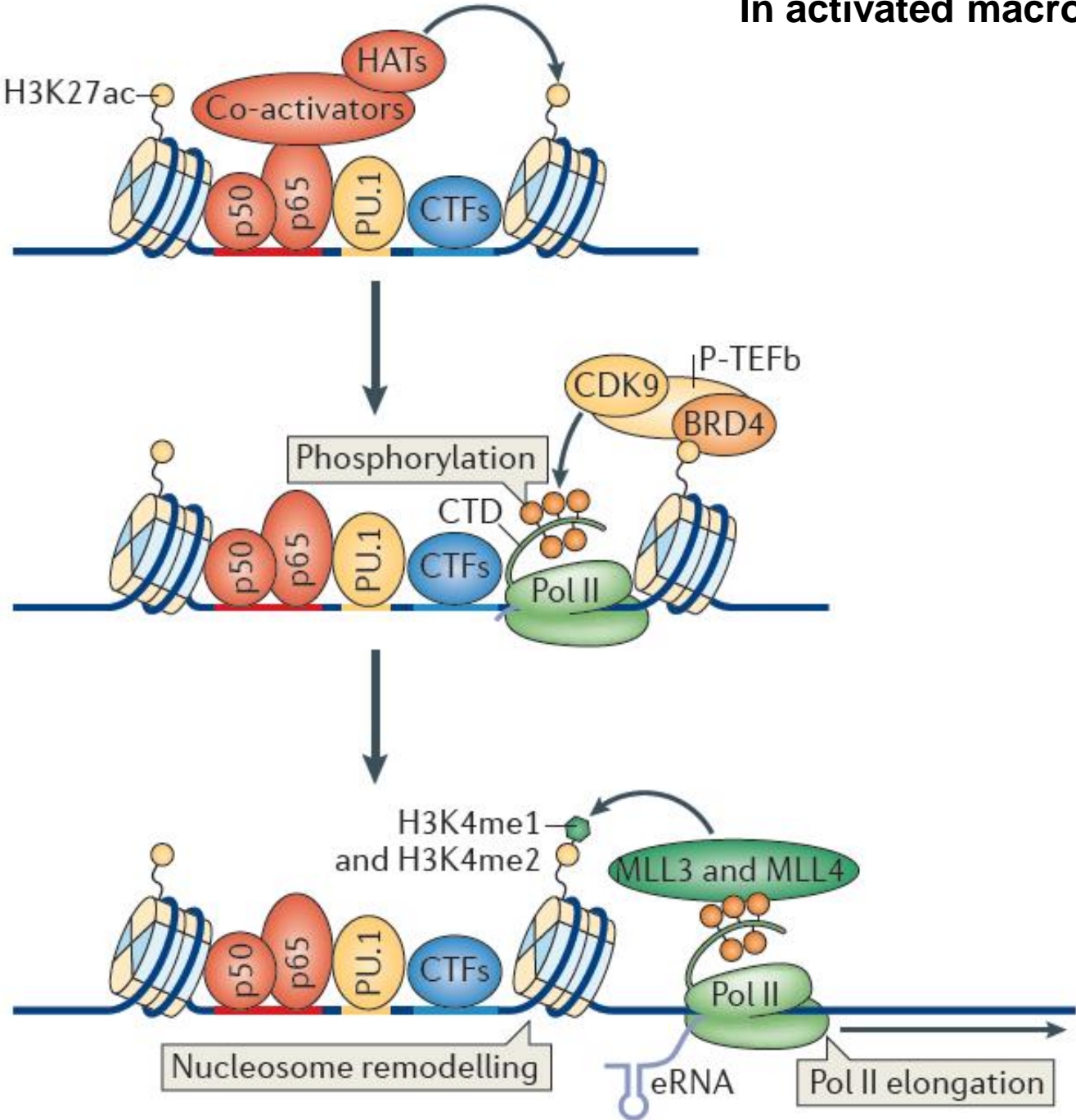


Figure 3 | **Cell type-specific enhancer selection and activation.** a | Collaborative

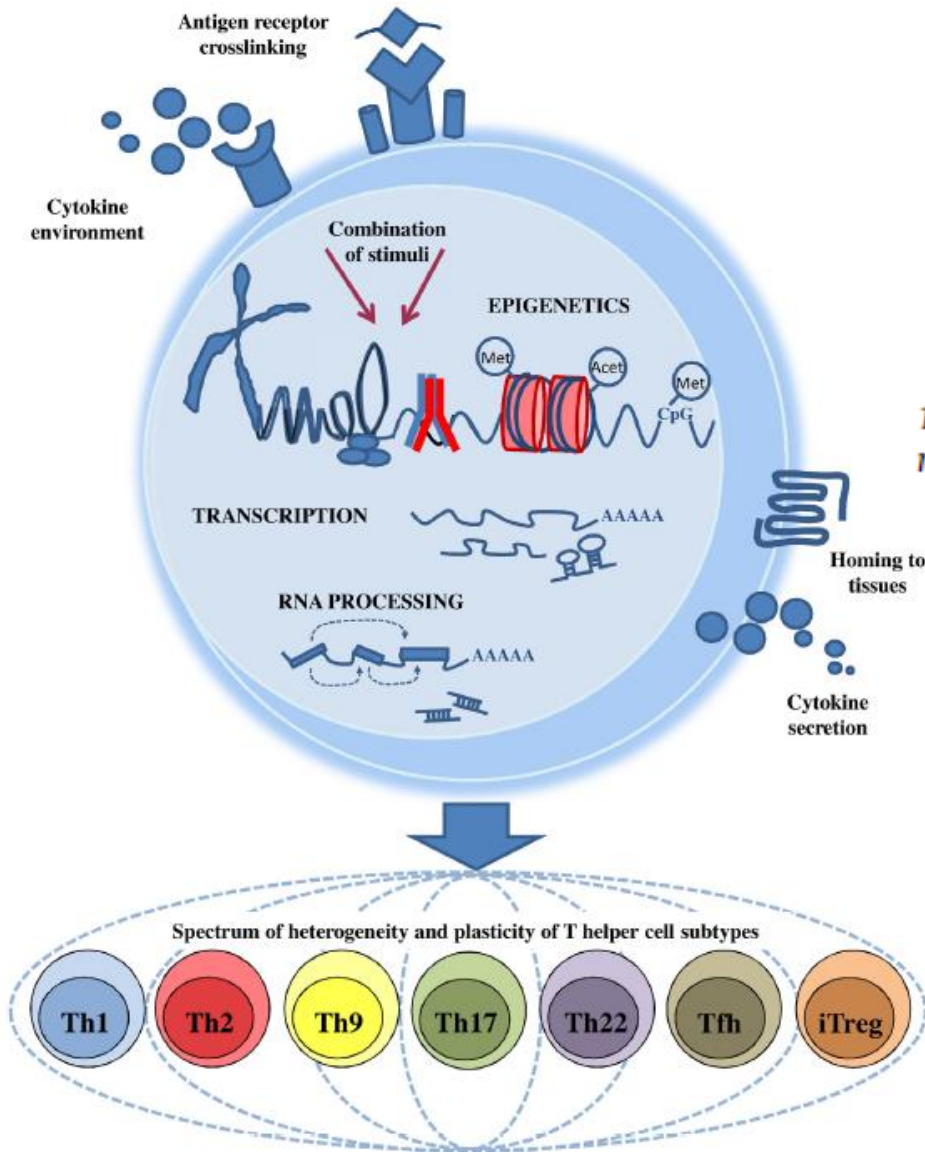


In activated macrophages



# Early T helper cell programming of gene expression in human

Soile Tuomela, Riitta Lahesmaa\*

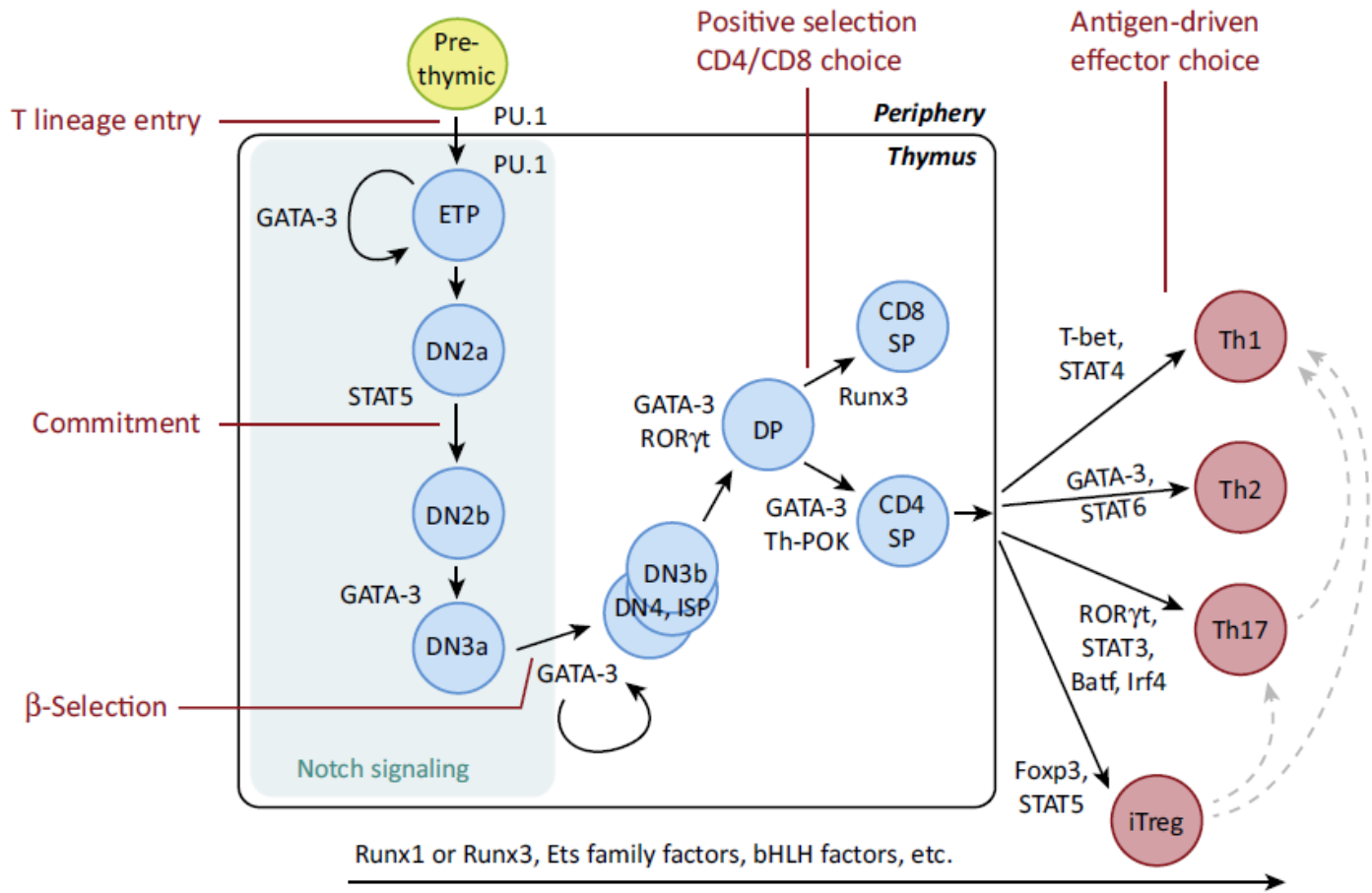


1.1. *Transcriptional regulation of human Th cell priming*

1.2. *Epigenetic regulation of Th cell priming in human*

1.3. *Regulation of Th cell differentiation by RNA processing and non-coding RNAs*

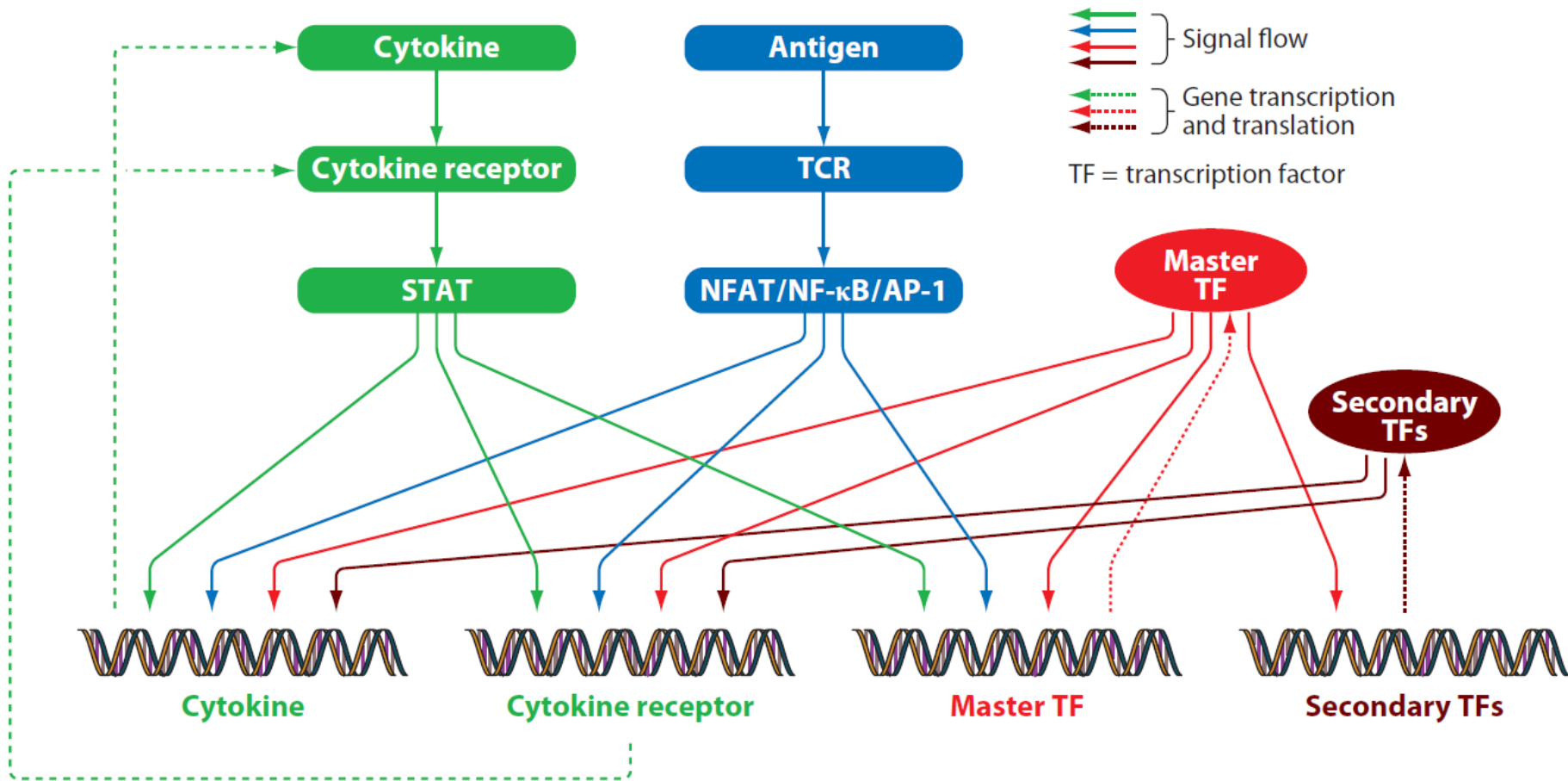






# Differentiation of Effector CD4 T Cell Populations\*

Jinfang Zhu, Hidehiro Yamane, and William E. Paul



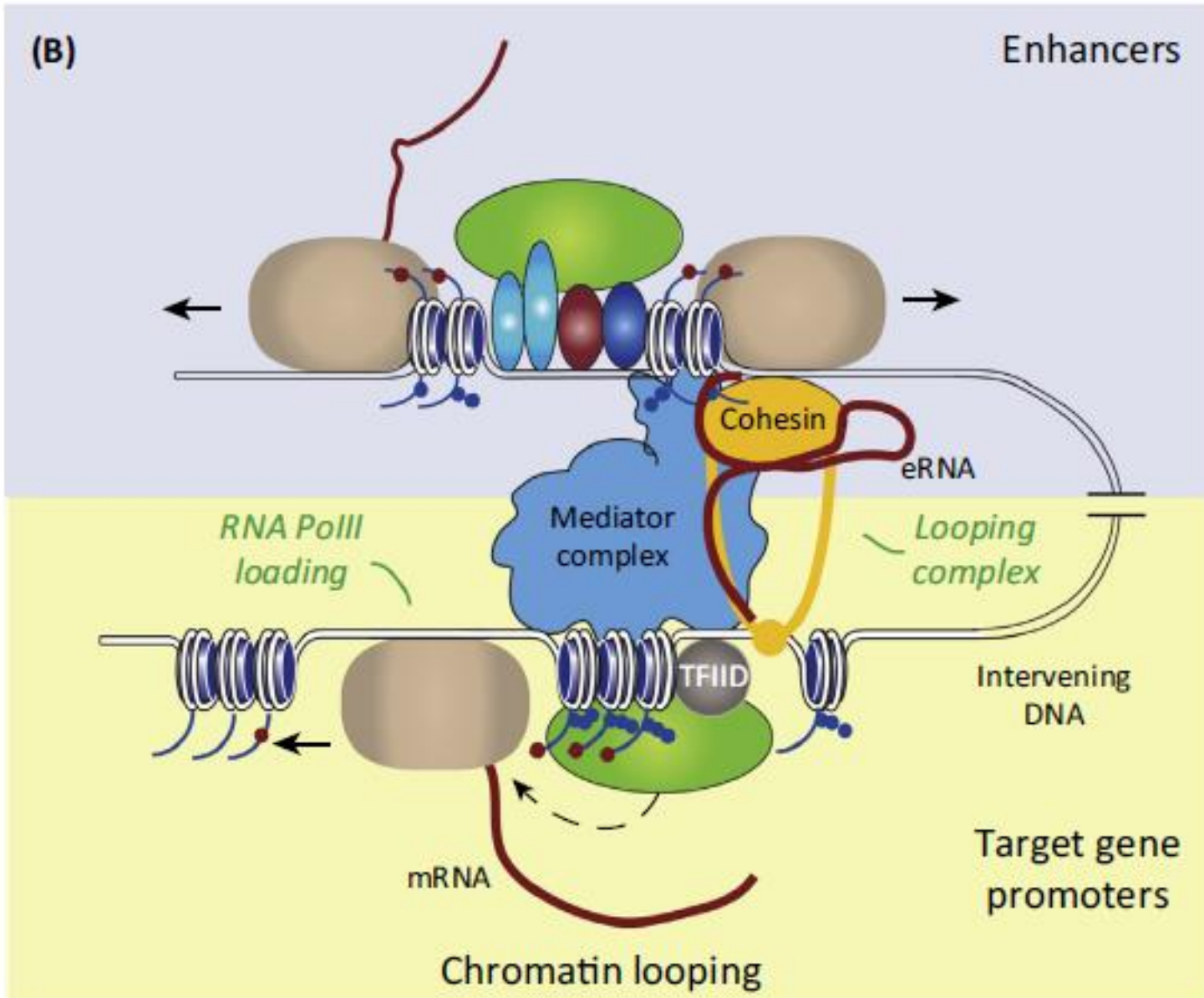
# Enhancer RNAs and regulated transcriptional programs

Michael T.Y. Lam<sup>1</sup>, Wenbo Li<sup>2</sup>, Michael G. Rosenfeld<sup>2</sup>, and Christopher K. Glass<sup>1,2</sup>

*Trends in Biochemical Sciences* April 2014, Vol. 39, No. 4

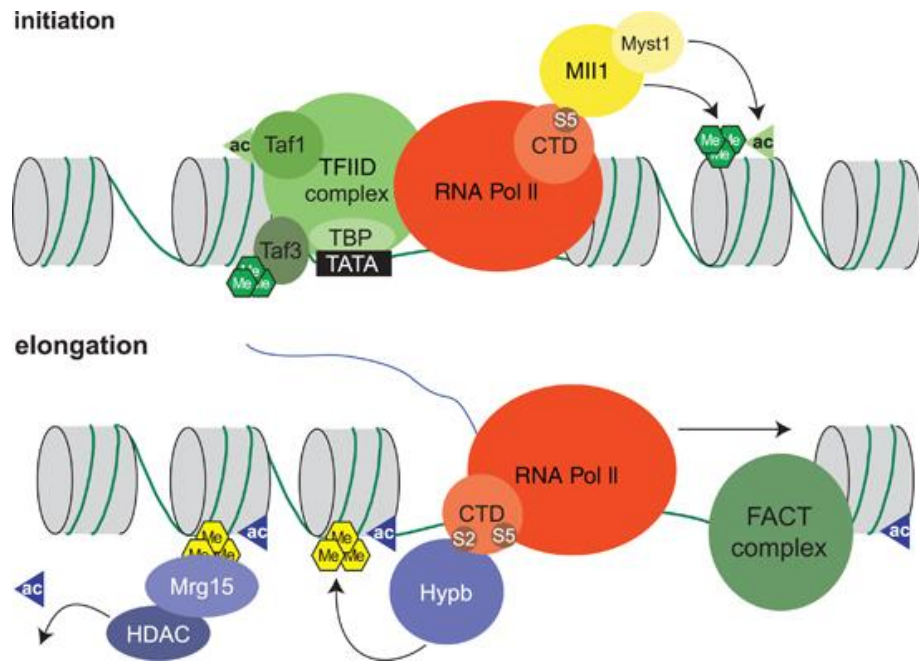


# eRNA mediates the long interactions

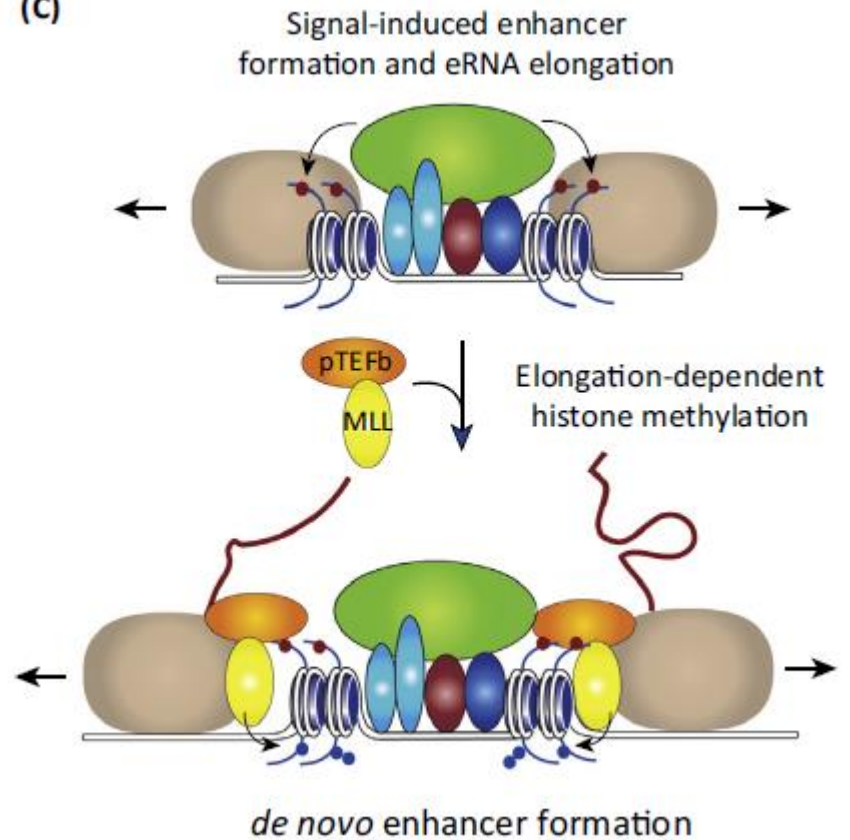


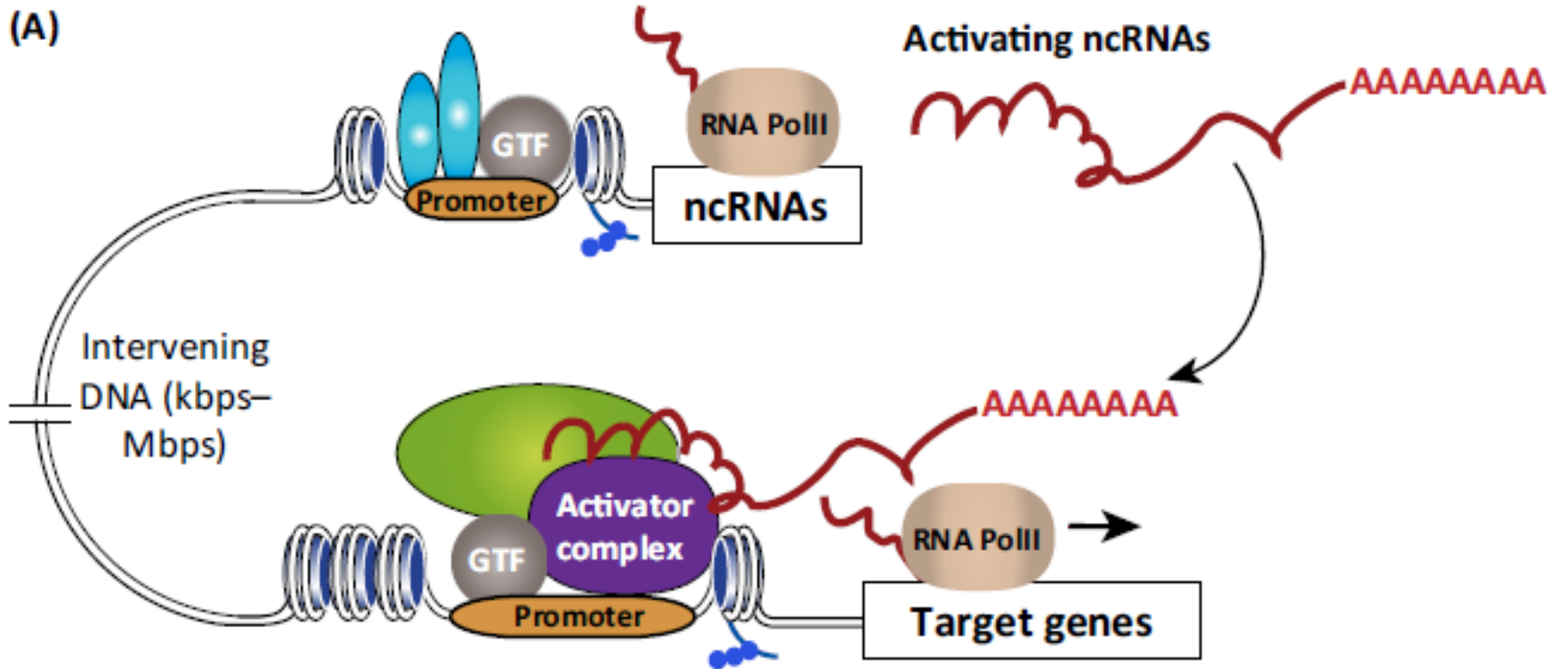


# Molecular mechanisms that underline enhancer activation



(c)

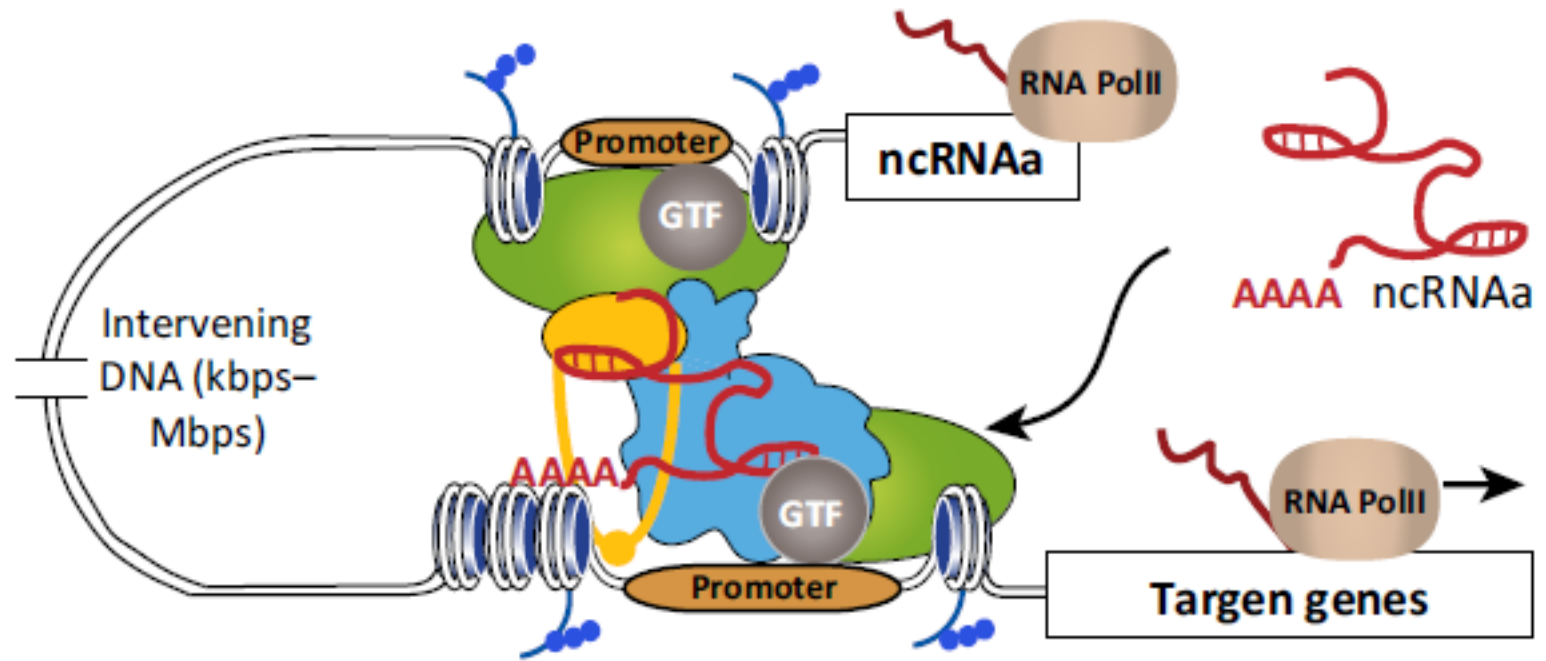




**Model 1: ncRNAs collaborate with transcriptional activators**



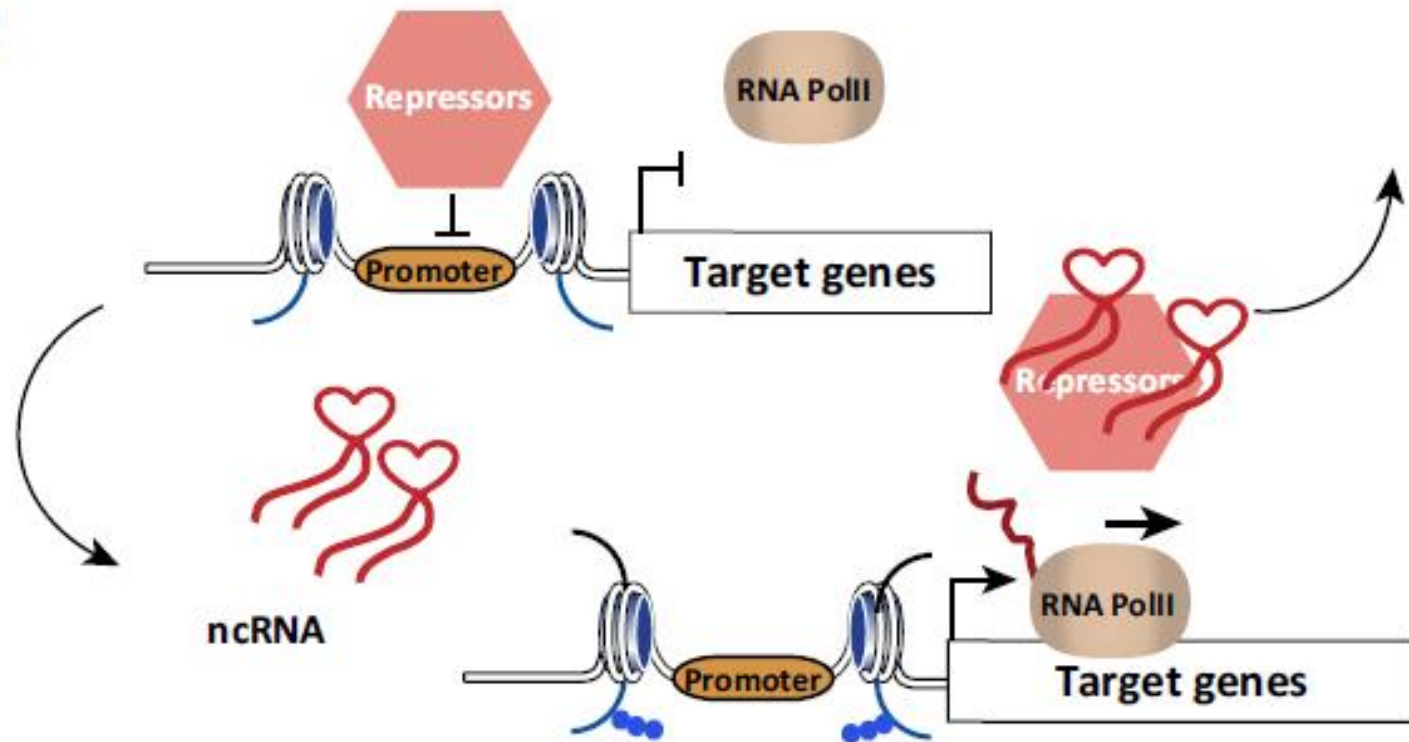
(B)



**Model 2: ncRNAs modulate chromatin loops**



(C)



### Model 3: ncRNAs evict transcriptional repressors

**Key:**

● Histone tail methylation



Signal dependent TFs



Looping complexes

● Coactivator complexes



General TFs



RNA polymerase II

● Transcriptional repressors

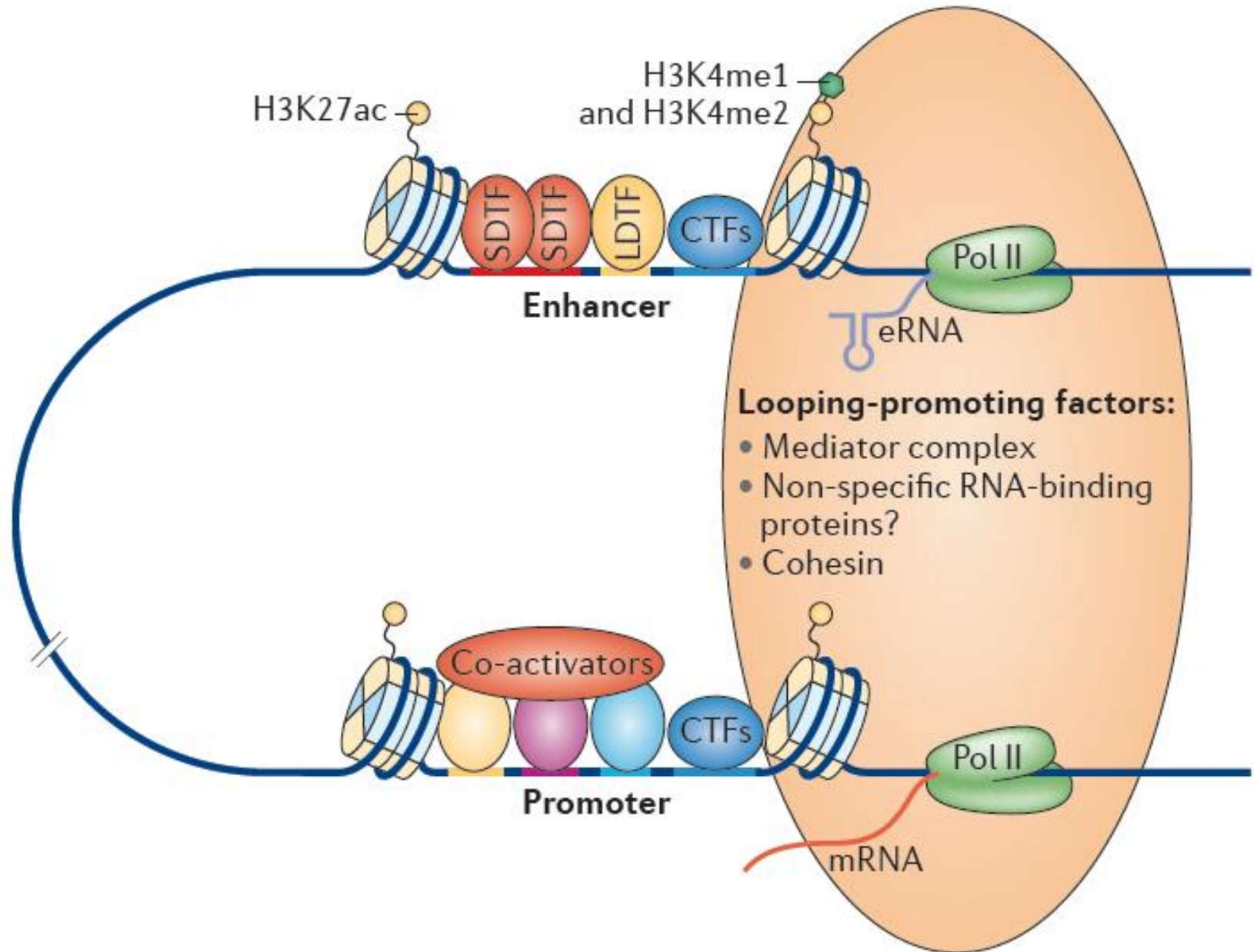


# Enhancer Function

- **Chromatin looping**
- **Super-enhancers, cluster of enhancers, key player in the cell identity and differentiation**



# CHROMATIN LOOPING





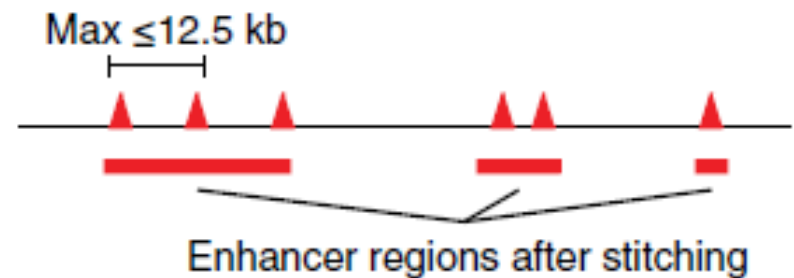
# Super-enhancers.

**a**

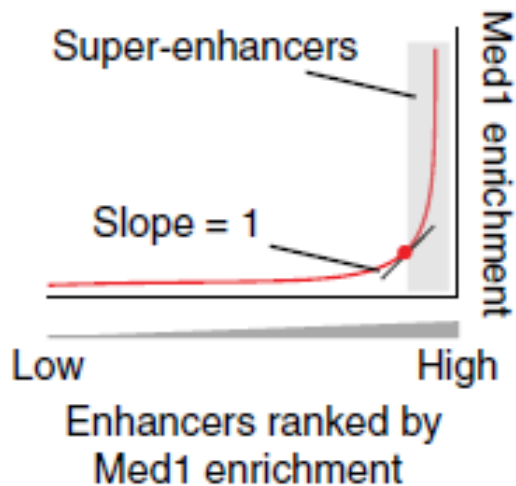
Step 1. Identification of enhancer locations



Step 2. Clustering of enhancers



Step 3. Identify super-enhancers



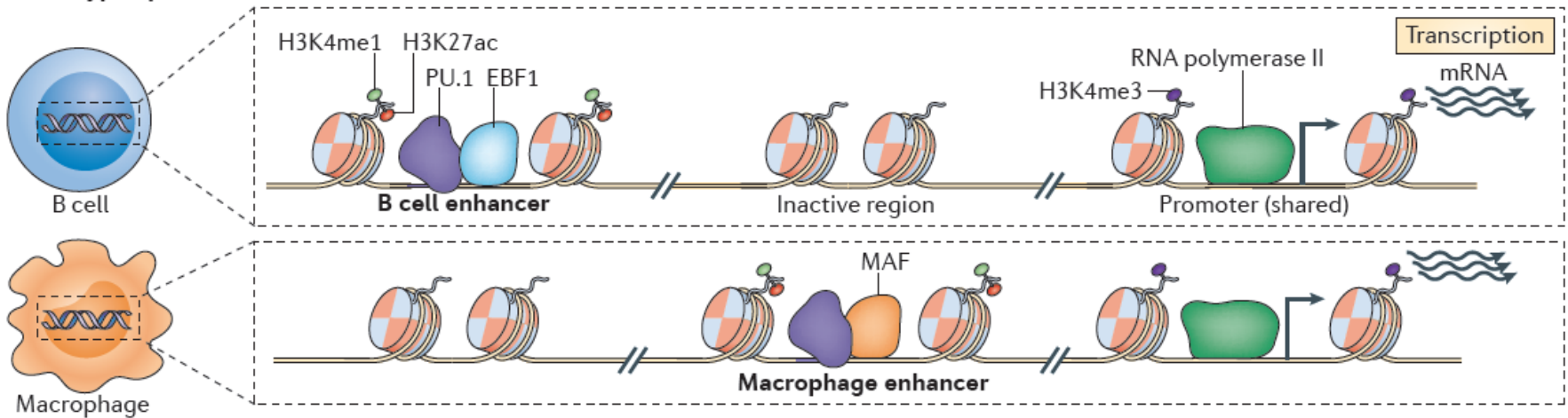
**b**

Factor used for step 1	Factor used for step 3	Reference
Oct4 + Sox2 + Nanog, Pu.1	Med1	Whyte <i>et al.</i>
MyoD, T-bet, C/EBP $\alpha$	MyoD, T-bet, C/EBP $\alpha$	Whyte <i>et al.</i>
H3K27ac	H3K27ac	Hnisz <i>et al.</i>
Med1	Med1	Loven <i>et al.</i>



# Cell-type-specific enhancers to regulate same genes

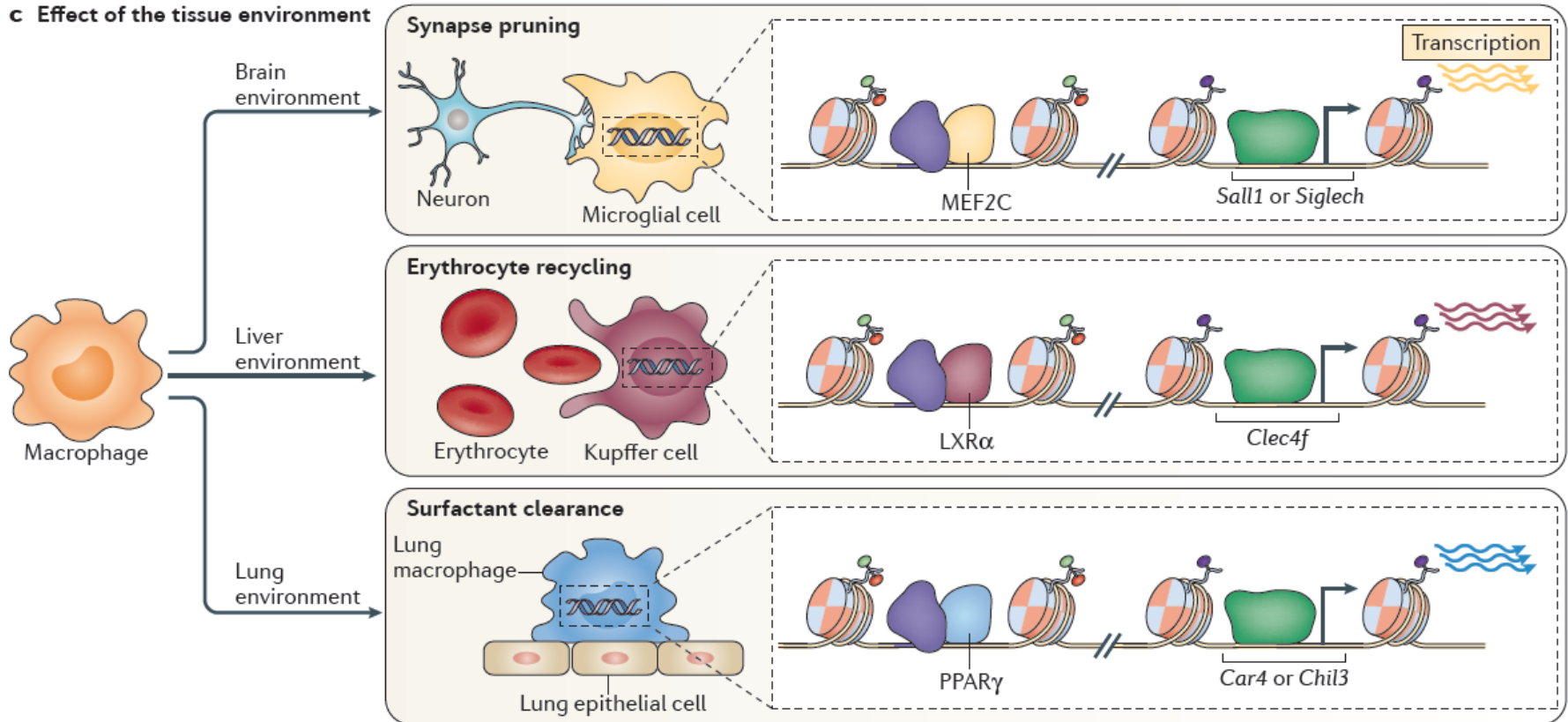
## a Cell-type-specific enhancers





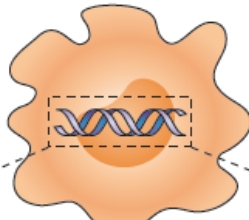
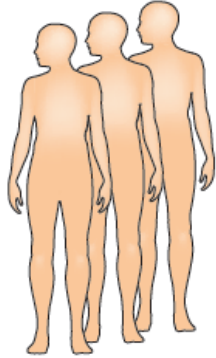
# Effect of the tissue environment

## c Effect of the tissue environment

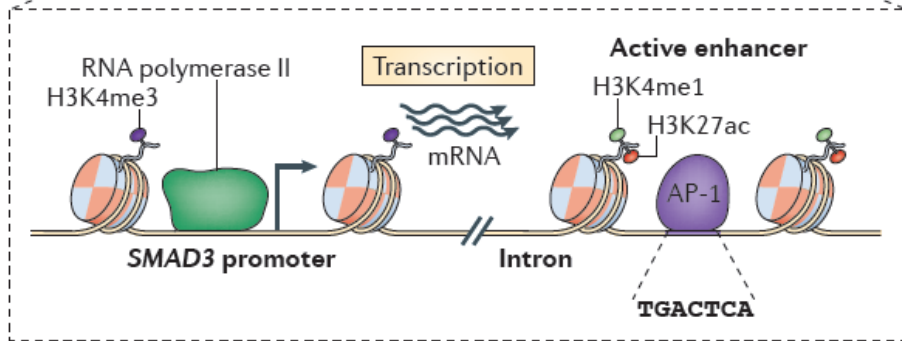


# Association of human chromatin data and susceptibility to immune disease

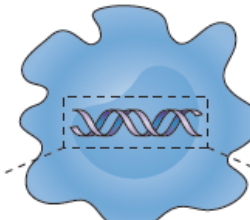
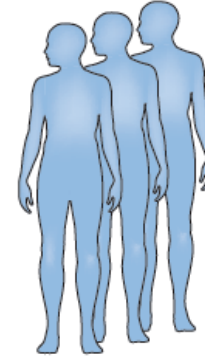
Healthy cohort



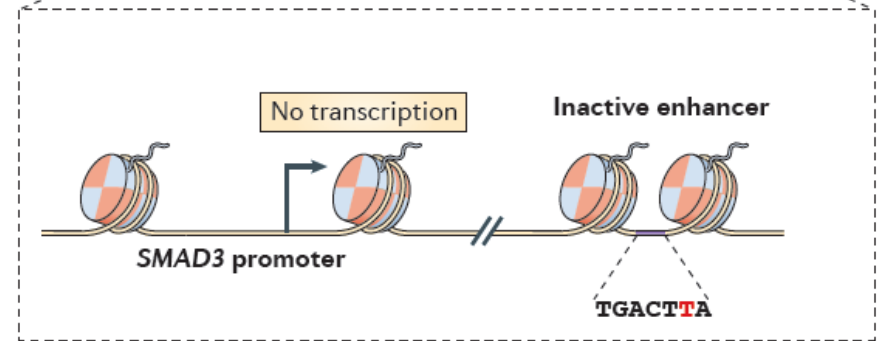
Monocyte



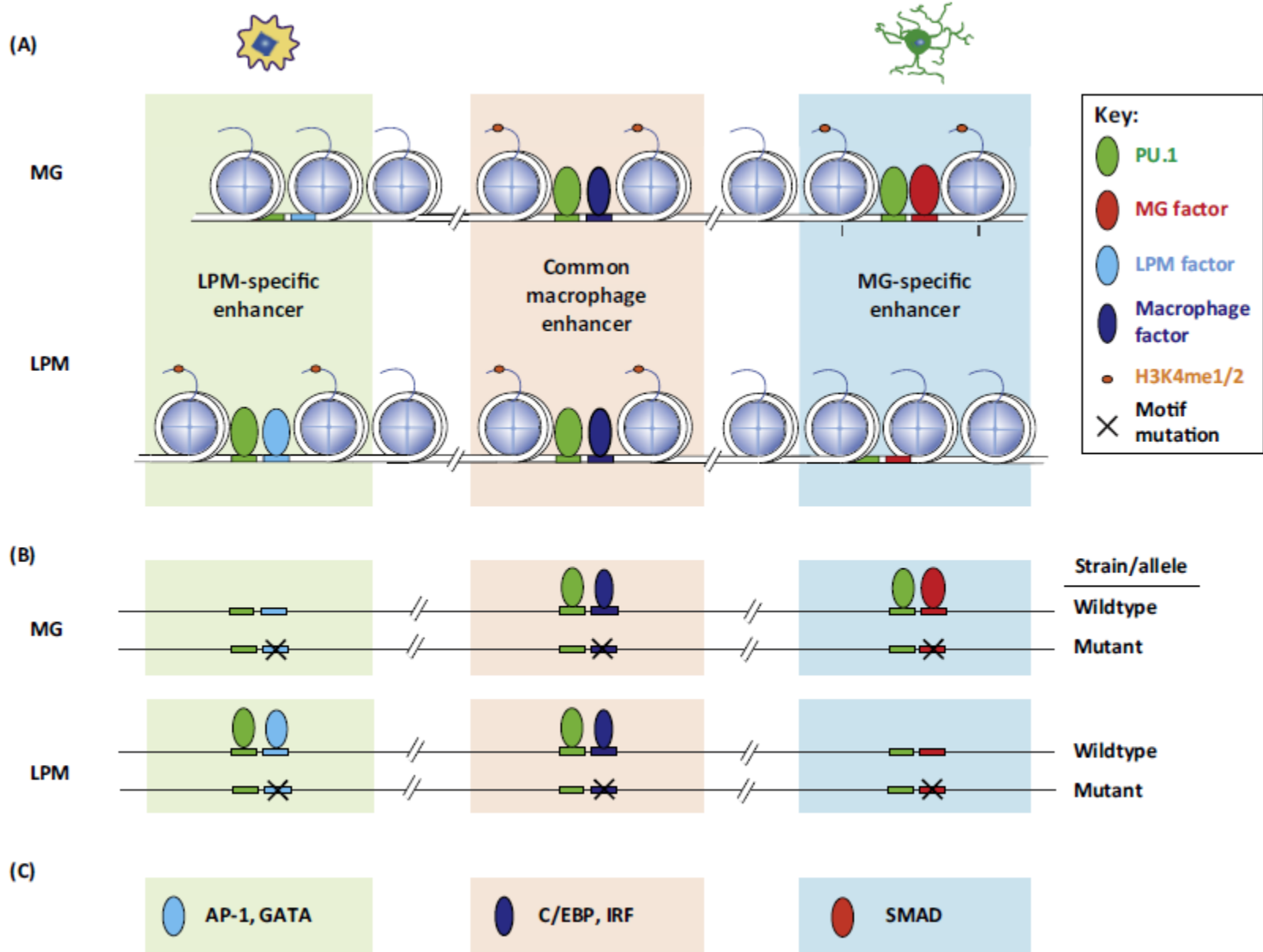
Disease cohort



Altered monocyte



# NATURAL GENETIC VARIATION IS ASSOCIATED WITH TF BINDING



## **SNPs in the genomic regulatory regions may affect:**

- **Enhancer Activation: loss of TFs interaction or TFs recruitment.**
- **Enhancer Selection: loss or association of LTDF**
- **Alteration of timing or specific tissues activation**
- **Long range interaction between genomic regulatory regions**



**How are SNPs studying in genome-wide manner?**



# Super-Enhancers in the Control of Cell Identity and Disease

Denes Hnisz,<sup>1,3</sup> Brian J. Abraham,<sup>1,3</sup> Tong Ihn Lee,<sup>1,3</sup> Ashley Lau,<sup>1,2</sup> Violaine Saint-André,<sup>1</sup> Alla A. Sigova,<sup>1</sup> Heather A. Hoke,<sup>1,2</sup> and Richard A. Young<sup>1,2,\*</sup>

<sup>1</sup>Whitehead Institute for Biomedical Research, 9 Cambridge Center, Cambridge, MA 02142, USA

<sup>2</sup>Department of Biology, Massachusetts Institute of Technology, Cambridge, MA 02139, USA

<sup>3</sup>These authors contributed equally to this work

\*Correspondence: [young@wi.mit.edu](mailto:young@wi.mit.edu)

<http://dx.doi.org/10.1016/j.cell.2013.09.053>

## SUMMARY

Super-enhancers are large clusters of transcriptional enhancers that drive expression of genes that define cell identity. Improved understanding of the roles that super-enhancers play in biology would be afforded by knowing the constellation of factors that constitute these domains and by identifying super-enhancers across the spectrum of human cell types. We describe here the population of transcription factors, cofactors, chromatin regulators, and transcription apparatus occupying super-enhancers in embryonic stem cells and evidence that super-enhancers are highly transcribed. We produce a catalog of super-enhancers in a broad range of human cell types and find that super-enhancers associate with genes that control and define the biology of these cells. Interestingly, disease-associated variation is especially enriched in the super-enhancers of disease-relevant cell types. Furthermore, we find that cancer cells generate super-enhancers at oncogenes and other genes important in tumor pathogenesis. Thus, super-enhancers play key roles in human cell identity in health and in disease.



## SUMMARY

Super-enhancers are large clusters of transcriptional enhancers that drive expression of genes that define cell identity. Improved understanding of the roles that super-enhancers play in biology would be afforded by knowing the constellation of factors that constitute these domains and by identifying super-enhancers across the spectrum of human cell types. We describe here the population of transcription factors, cofactors, chromatin regulators, and transcription apparatus occupying super-enhancers in embryonic stem cells and evidence that super-enhancers are highly transcribed. We produce a catalog of super-enhancers in a broad range of human cell types and find that super-enhancers associate with genes that control and define the biology of these cells. Interestingly, disease-associated variation is especially enriched in the super-enhancers of disease-relevant cell types. Furthermore, we find that cancer cells generate super-enhancers at oncogenes and other genes important in tumor pathogenesis. Thus, super-enhancers play key roles in human cell identity in health and in disease.

### DEFINITION

### AIM

1) Protein complexes

2) SE cell type-specific

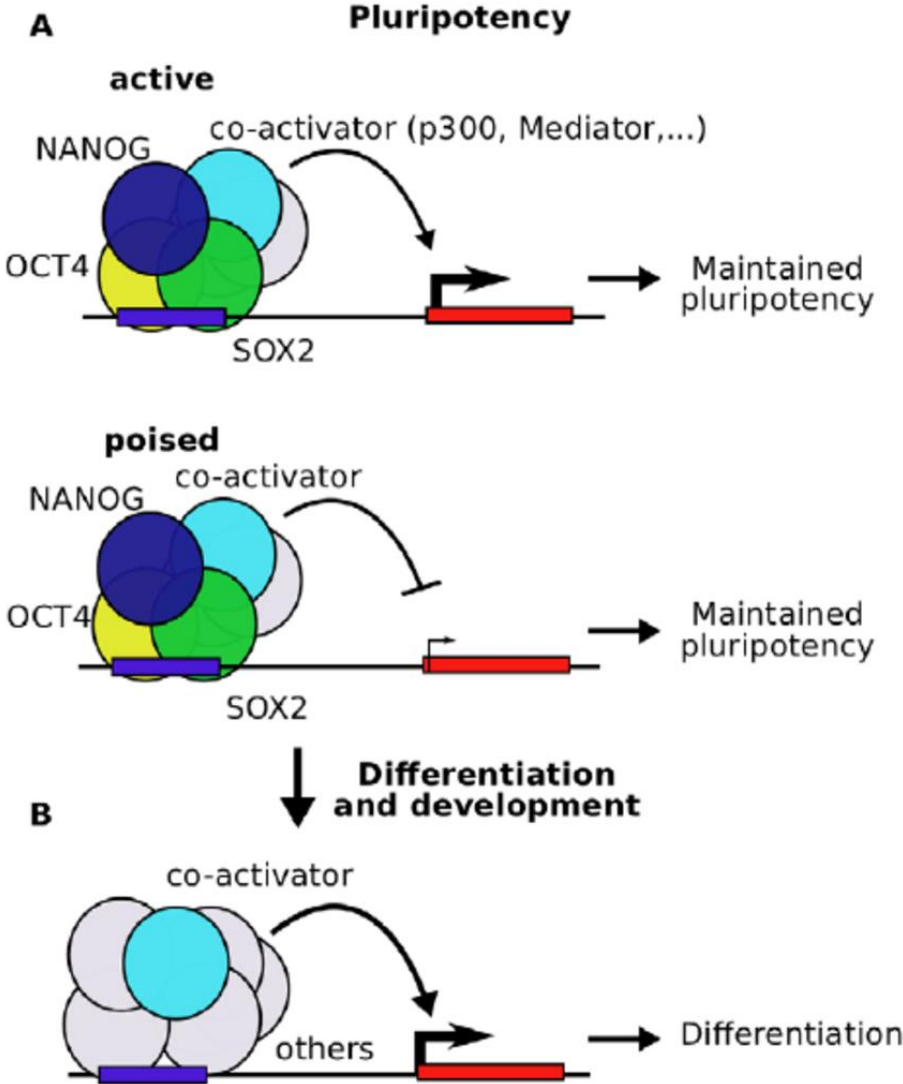
3) SNPs linked to disease in SE

### CONCLUSION



# SOX2, Nanog, OCT4: transcription factors that bind SE controlling genes for maintained pluripotency

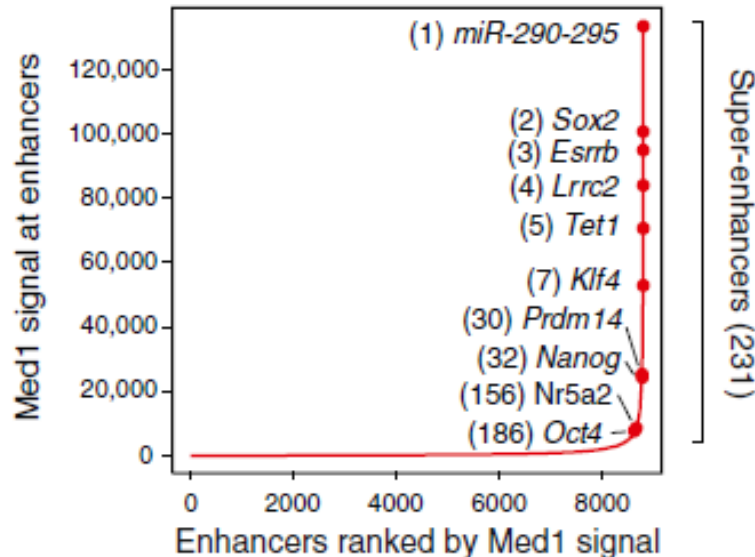
## Murine Embryonic stem cells (ESC)





# Mediator Coactivator Complexes and Master TFs are bound at Super-enhancers

## Transcription Factors in ESCs



Super-enhancers are clusters of enhancers—formed by binding of high levels of master transcription factors and Mediator coactivator—that drive high-level expression of genes encoding key regulators of cell identity (Figure 1A) (Whyte et al., 2013). Five ESC transcription factors were previously shown to occupy super-enhancers (Oct4, Sox2, Nanog, Klf4, and Esrrb) (Whyte et al., 2013), but there are many additional transcription factors that contribute to the control of ESCs (Ng and Surani, 2011; Orkin and Hochedlinger, 2011; Young, 2011). We compiled ChIP-seq data for 15 additional transcription factors in ESCs, for which high-quality ChIP-seq data were available, and investigated whether they occupy enhancers defined by Oct4, Sox2, and Nanog (OSN) co-occupancy (Whyte et al., 2013) (Table S1 avail-

(A) Distribution of Med1 ChIP-seq signal at enhancers reveals two classes of enhancers in ESCs. Enhancer regions are plotted in an increasing order based on their input-normalized Med1 ChIP-seq signal. Super-enhancers are defined as the population of enhancers above the inflection point of the curve. Example super-enhancers are highlighted along with their respective ranks and their associated genes.



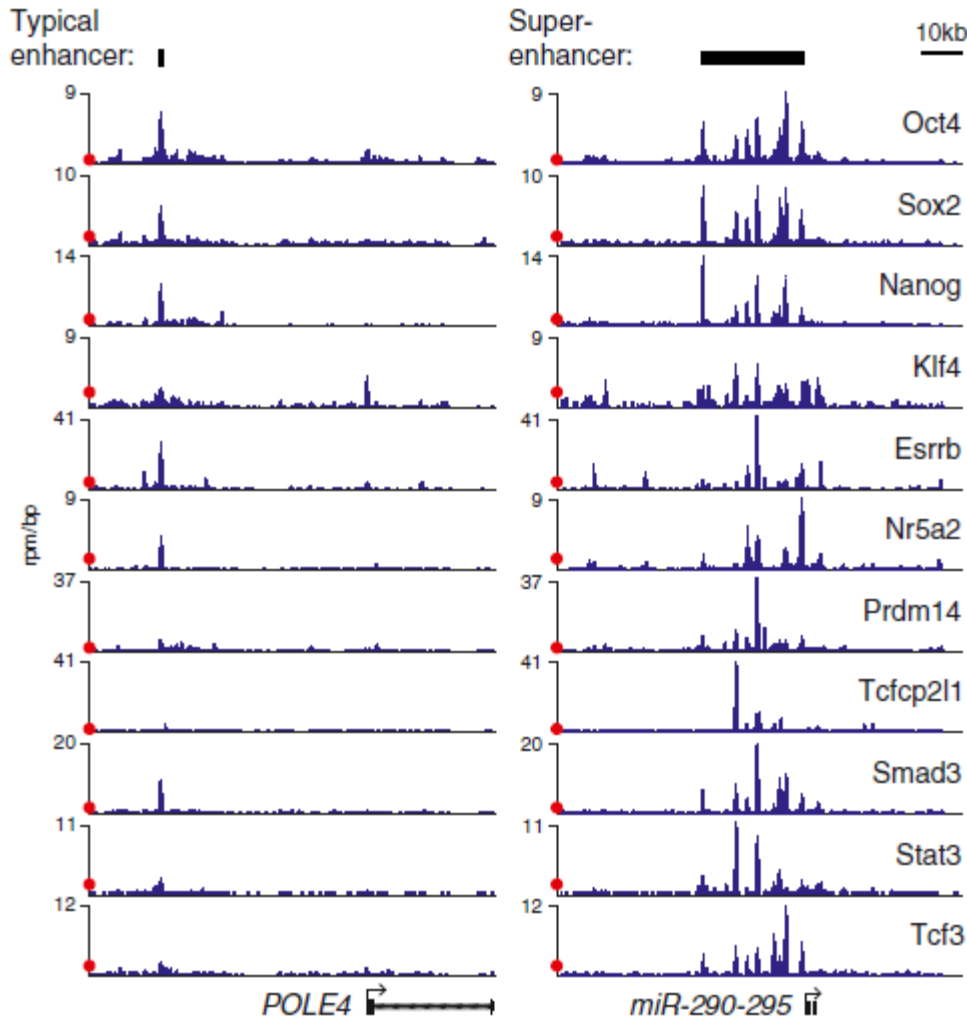
## **Bioinformatic analysis for the definition of SE:**

- Signal in proximity of the gene**
- signal extended in the genomic regions that identify SE**
- increased numbers of reads into SE respect to constituent, single enhancer in the SE**
- increased signal into SE respect to typical enhancer**



# Mediator Coactivator Complexes and Master TFs are bound at Super-enhancers

## Chromatin Immunoprecipitation Binding Profiles at target genes



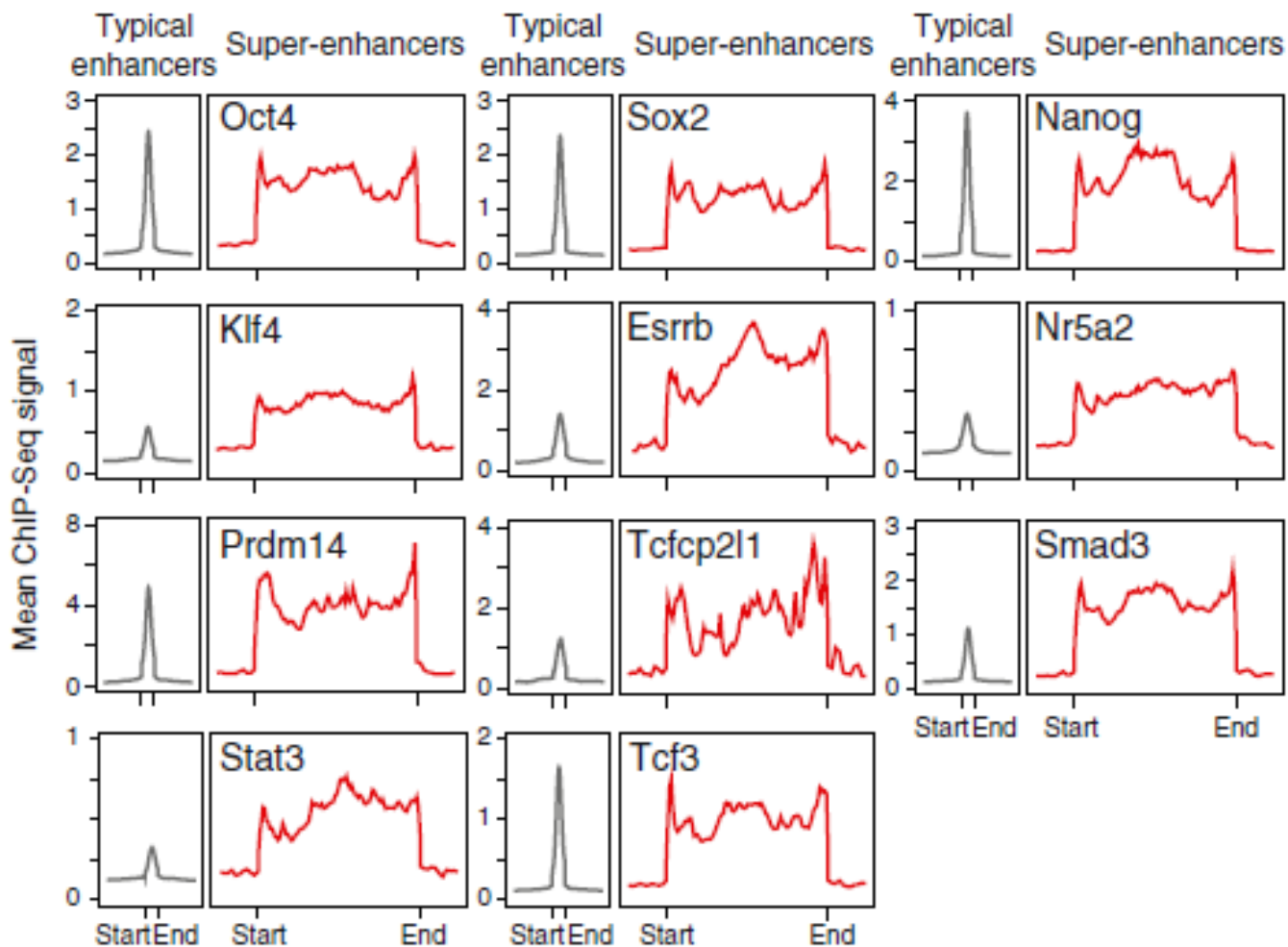
Transcription Factors

Specific Loci

(B) ChIP-seq binding profiles for the indicated transcription factors at the *POLE4* and *miR-290-295* loci in ESCs. Red dots indicate the median enrichment of all bound regions in the respective ChIP-seq data sets and are positioned at maximum 20% of the axis height. rpm/bp, reads per million per base pair.



# ChIP-seq signal across SE domains



(C) Metagene representations of the mean ChIP-seq signal for the indicated transcription factors across typical enhancers and super-enhancer domains. Metagenes are centered on the enhancer region, and the length of the enhancer reflects the difference in median lengths (703 bp for typical enhancers, 8,667 bp for super-enhancers). Additional 3 kb surrounding each enhancer region is also shown.

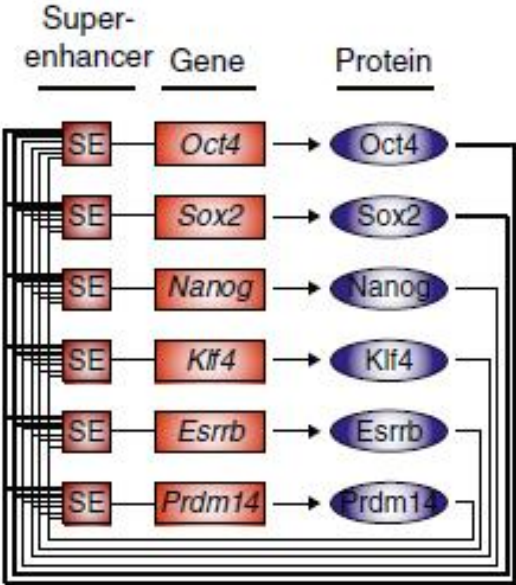


# TFs motif enrichment are used to associate gene target

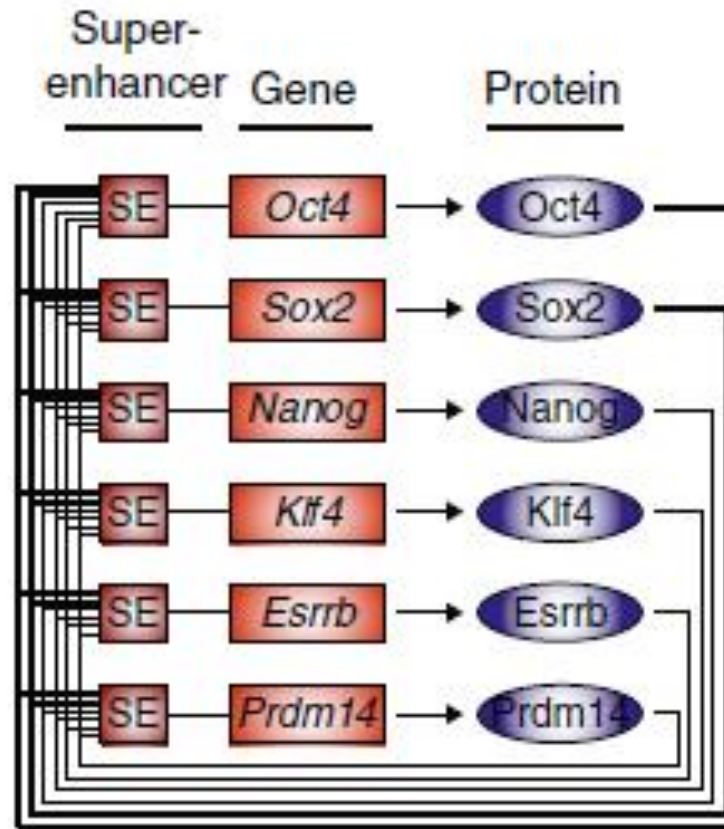
F

Transcription factor	Motif	P-value	Transcription factor	Motif	P-value
Oct4		$9.19 \times 10^{-64}$	Prdm14	n.a.	n.a.
Sox2		$3.01 \times 10^{-67}$	Tcfcp2l1		$6.83 \times 10^{-11}$
Nanog		$9.46 \times 10^{-17}$	Smad3		$9.31 \times 10^{-11}$
Klf4		$4.33 \times 10^{-6}$	Stat3		$2.90 \times 10^{-10}$
Esrrb		$2.55 \times 10^{-84}$	Tcf3		$5.46 \times 10^{-27}$
Nr5a2	n.a.	n.a.			

G



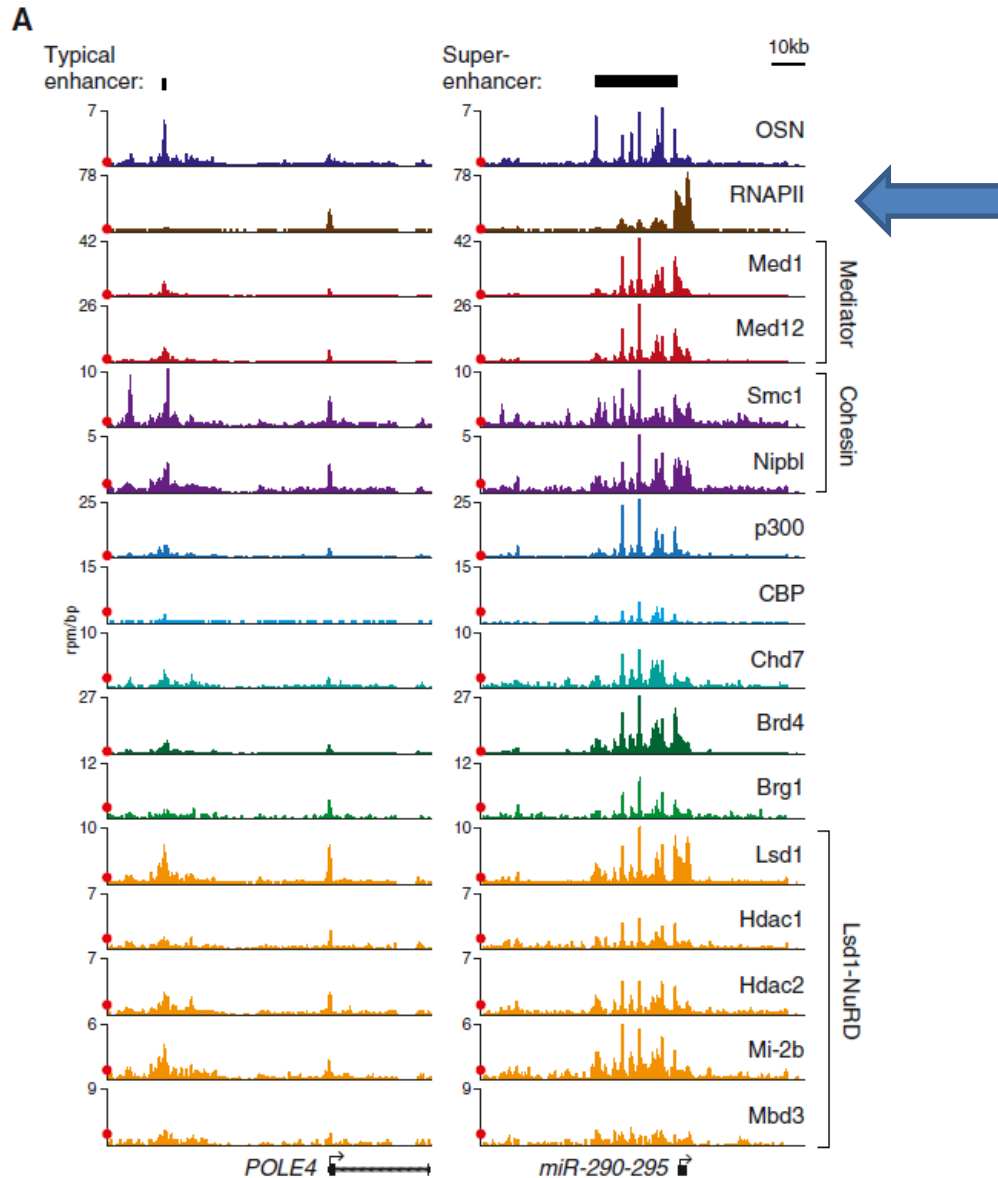
# Core Transcriptional Regulatory Circuit of ESCs



(G) Revised model of the core transcriptional regulatory circuitry of ESCs. The model contains an interconnected autoregulatory loop consisting of transcription factors that meet three criteria: (1) their genes are driven by super-enhancers, (2) they co-occupy their own super-enhancers as well as those of the other transcription factor genes in the circuit, and (3) they play essential roles in regulation of ESC state and iPSC reprogramming. The layout of the circuit model was adapted from [Whyte et al. \(2013\)](#).

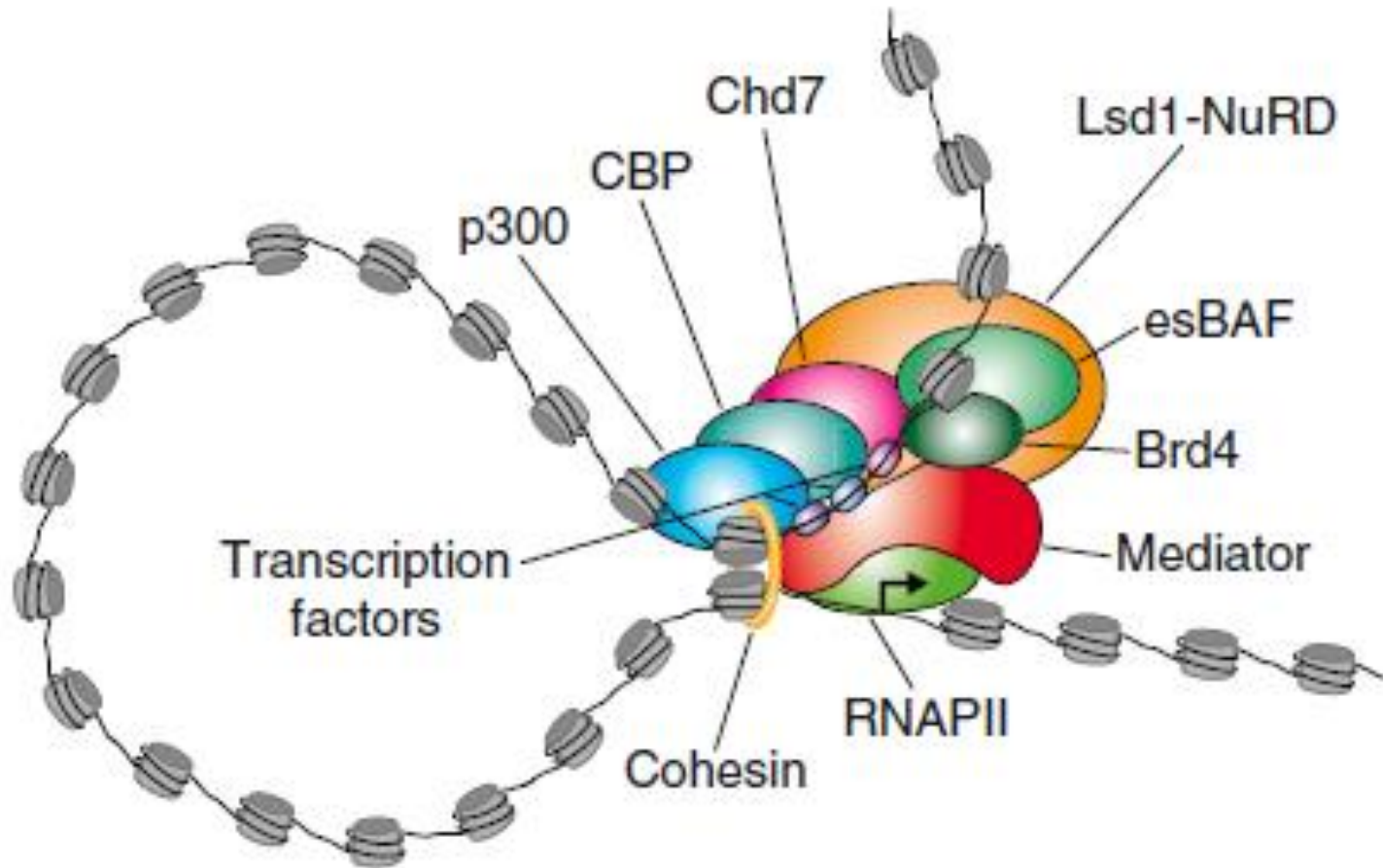


# Super-enhancers are occupied by a large portion of the enhancer-associated RNA polymerase II



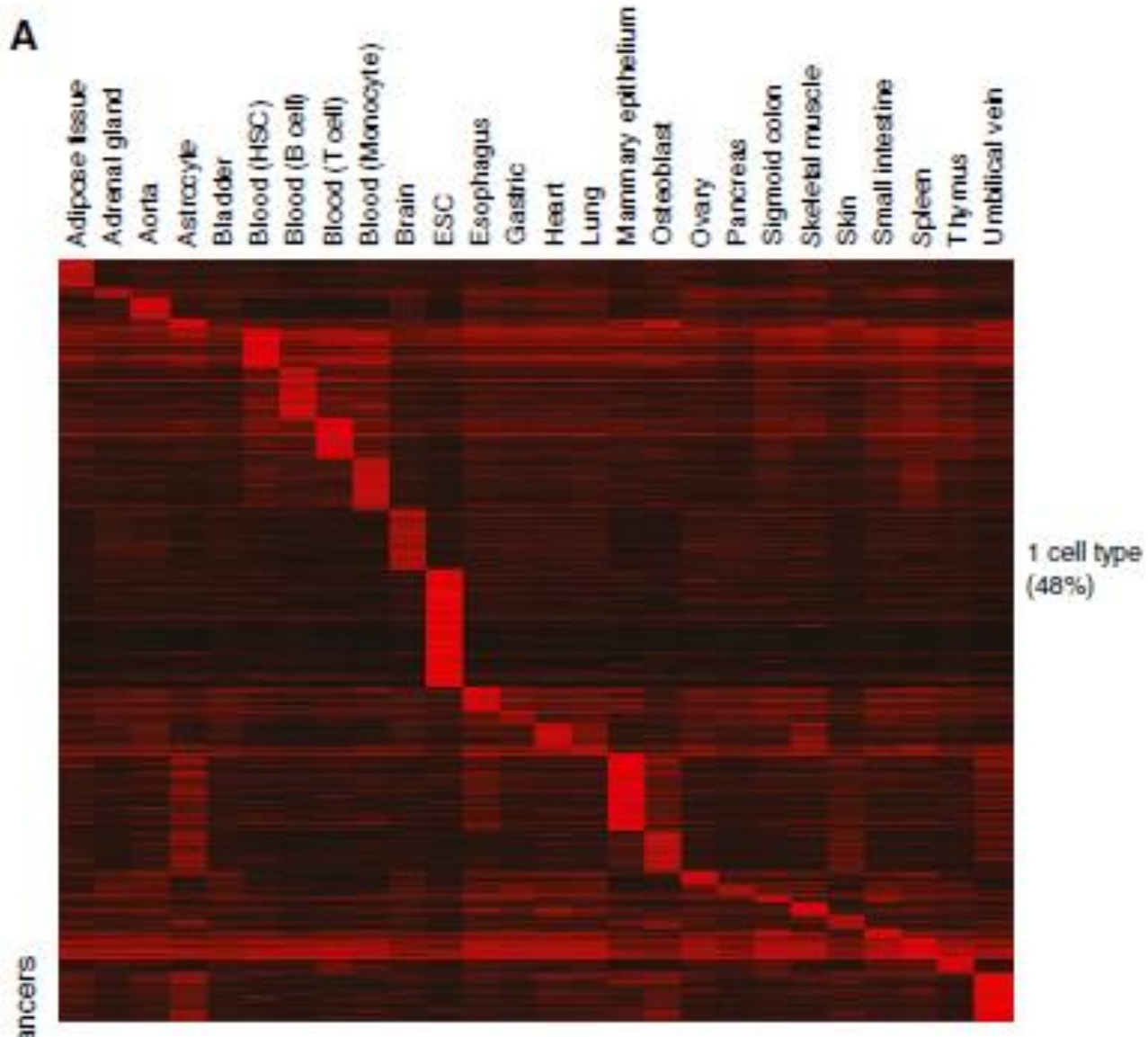


**Model showing RNAPII, transcriptional cofactors, and chromatin regulators that are found in ESC super-enhancers.** The indicated proteins are responsible for diverse enhancer-related functions, such as enhancer looping, gene activation, nucleosome remodeling, and histone modification.

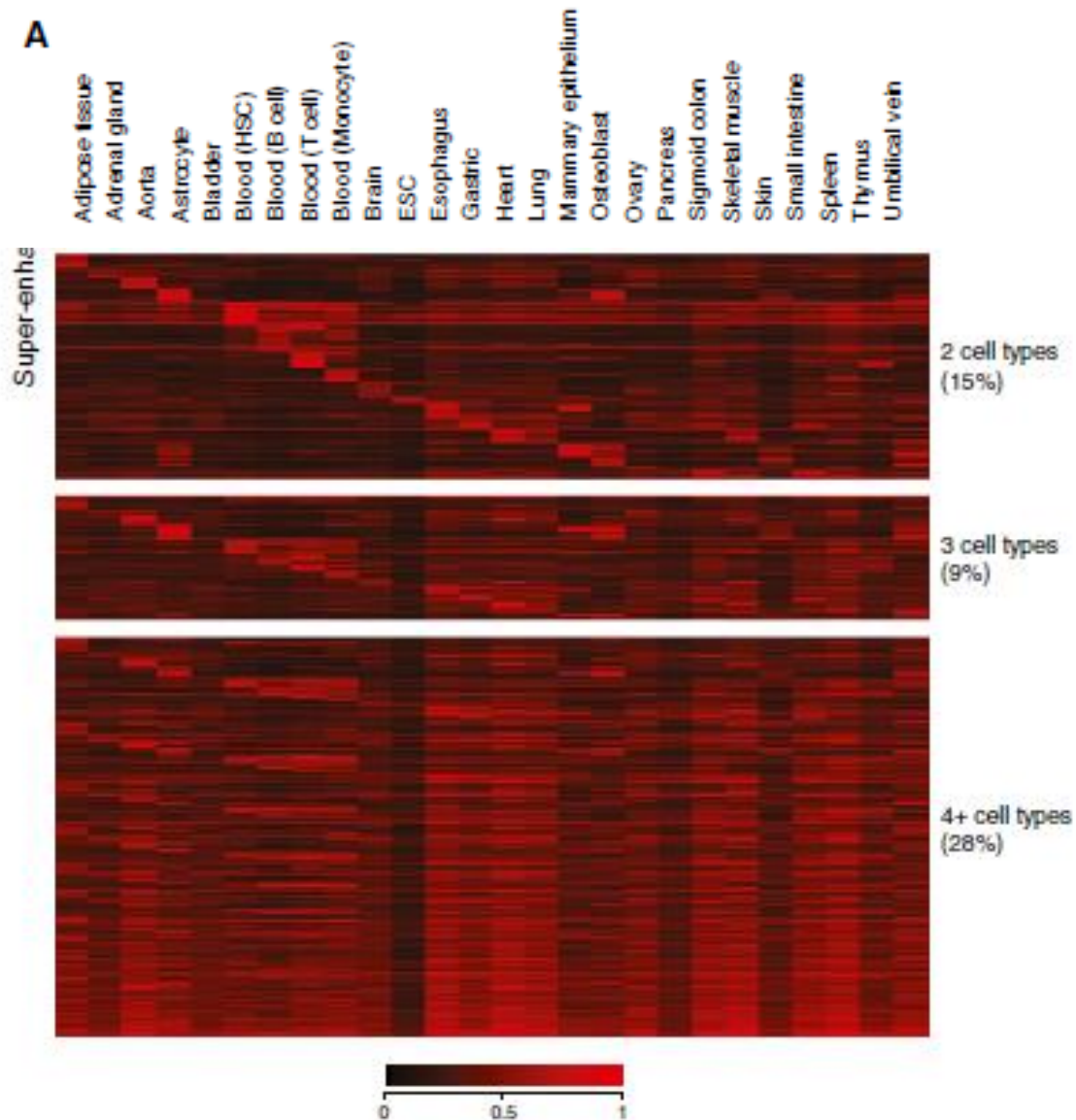




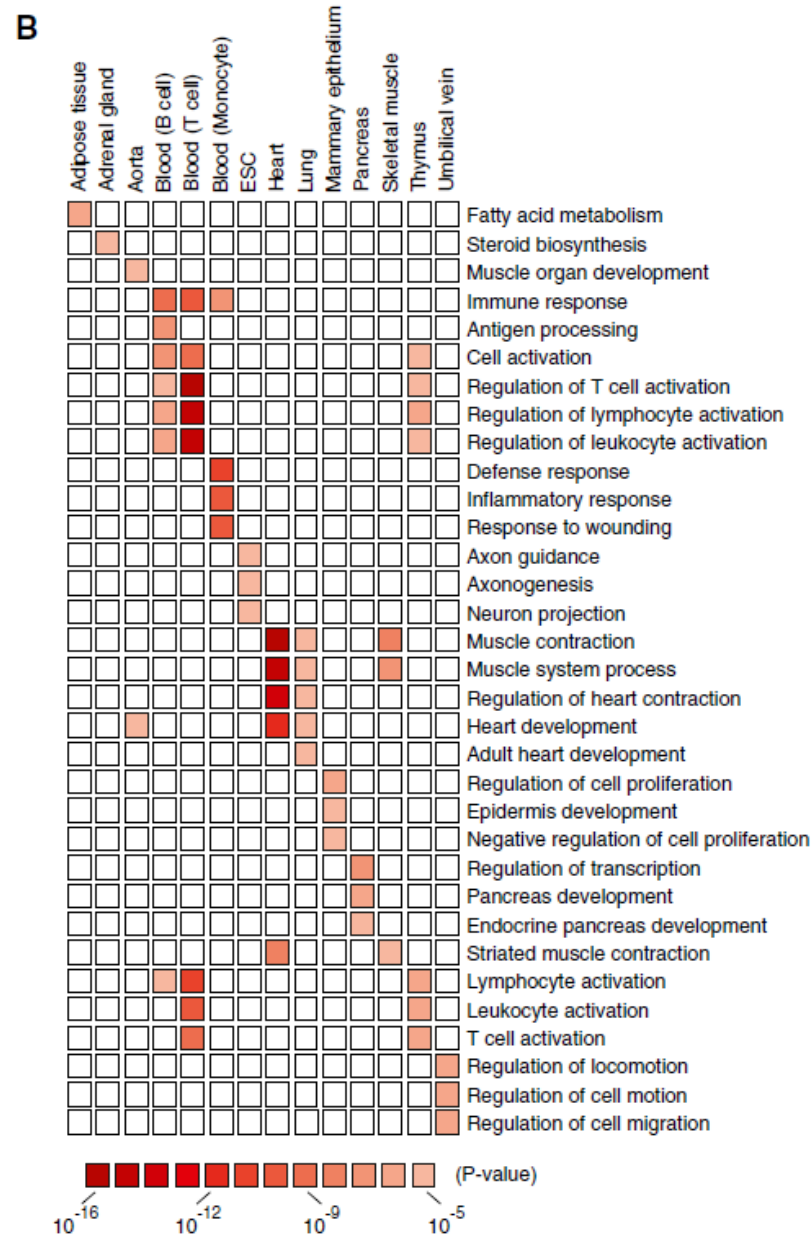
# SUPERENHANCER SHARED BETWEEN SEVERAL CELL TYPES



# SUPERENHANCER SHARED BETWEEN SEVERAL CELL TYPES



# GENE ASSOCIATED TO SUPERENHANCER IN SEVERAL CELL TYPES: GENE ONTOLOGY



# MASTER TRANSCRIPTION FACTORS IN SIX CELL TYPES

**C**

<u>Brain</u>	<u>Heart</u>	<u>Skeletal muscle</u>	<u>Lung</u>	<u>Adipose tissue</u>	<u>B cell</u>
NKX2-2	TBX20	MYOD1	NFIB	PPARG	IKZF3
OLIG1	TBX5	PITX2	TBX5	CEBPB	PAX5
BRN2	MEF2A	SIX1	CEBPA	CEBPD	BACH2
SOX10	NKX2-5	TEAD4	TBX2	CREB1	OCT2
SOX2	GATA4		TBX3		IKZF1
					IRF8

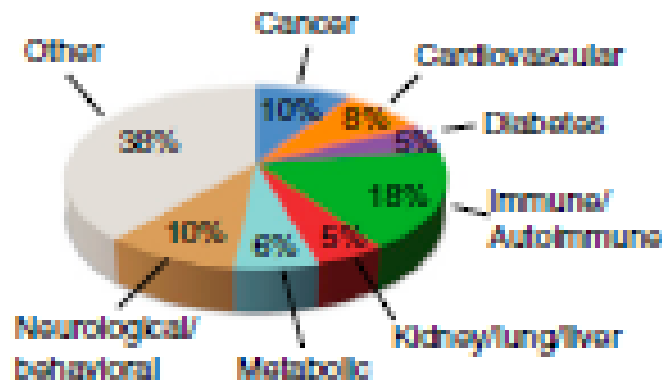
(C) Candidate master transcription factors identified in six cell types. All of these transcription factors were previously demonstrated to play key roles in the biology of the respective cell type or facilitate reprogramming to the respective cell type.



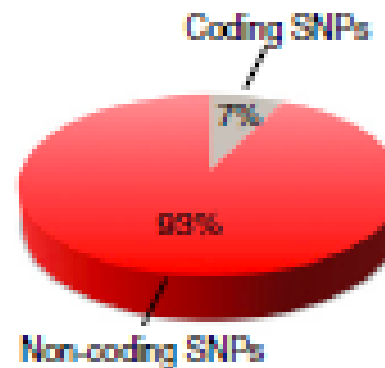
# SINGLE NUCLEOTIDE MUTATIONS LINKED TO DISEASE (GWAS) ASSOCIATED TO SUPERENHANCERS

**A**

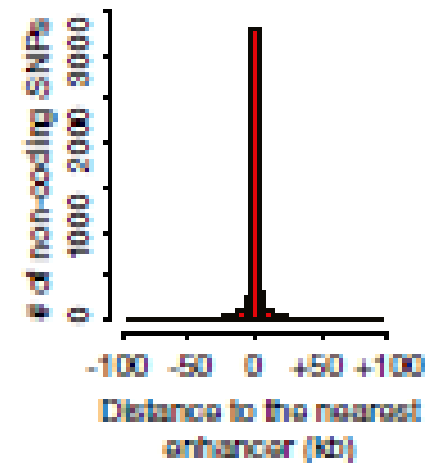
5,303 SNPs from 1,675 GWAS studies



Coding vs. non-coding SNPs

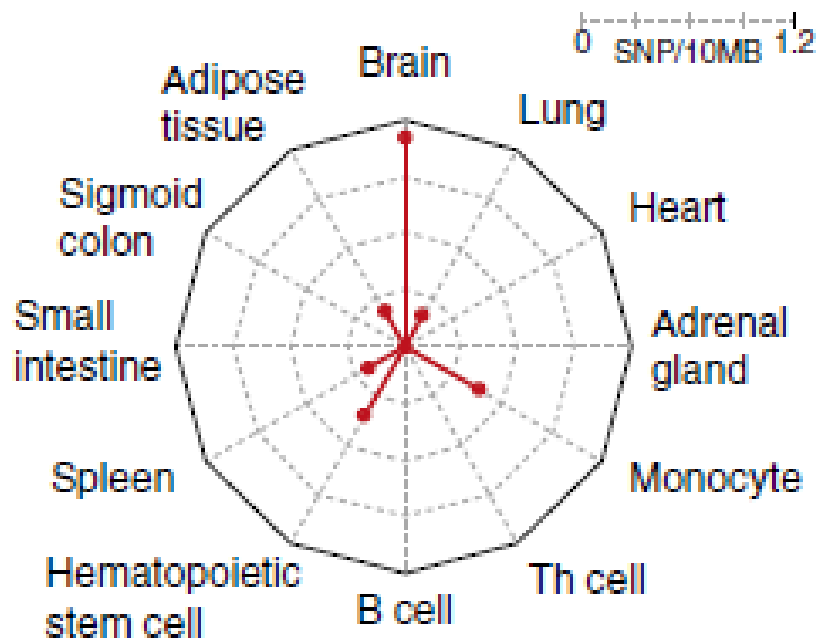


Proximity to enhancers

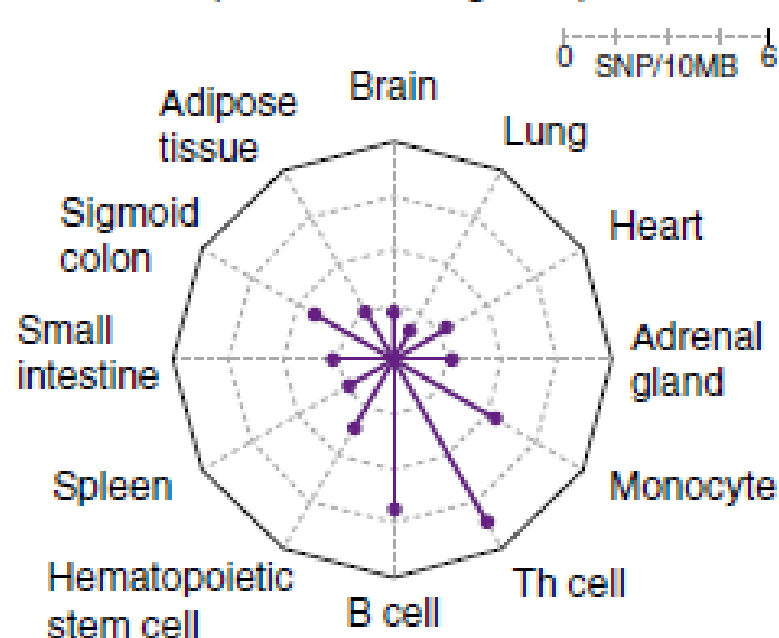


# SINGLE NUCLEOTIDE MUTATIONS LINKED TO DISEASE (GWAS) ASSOCIATED TO SUPERENHANCERS

Alzheimer's disease  
(27 non-coding SNP)



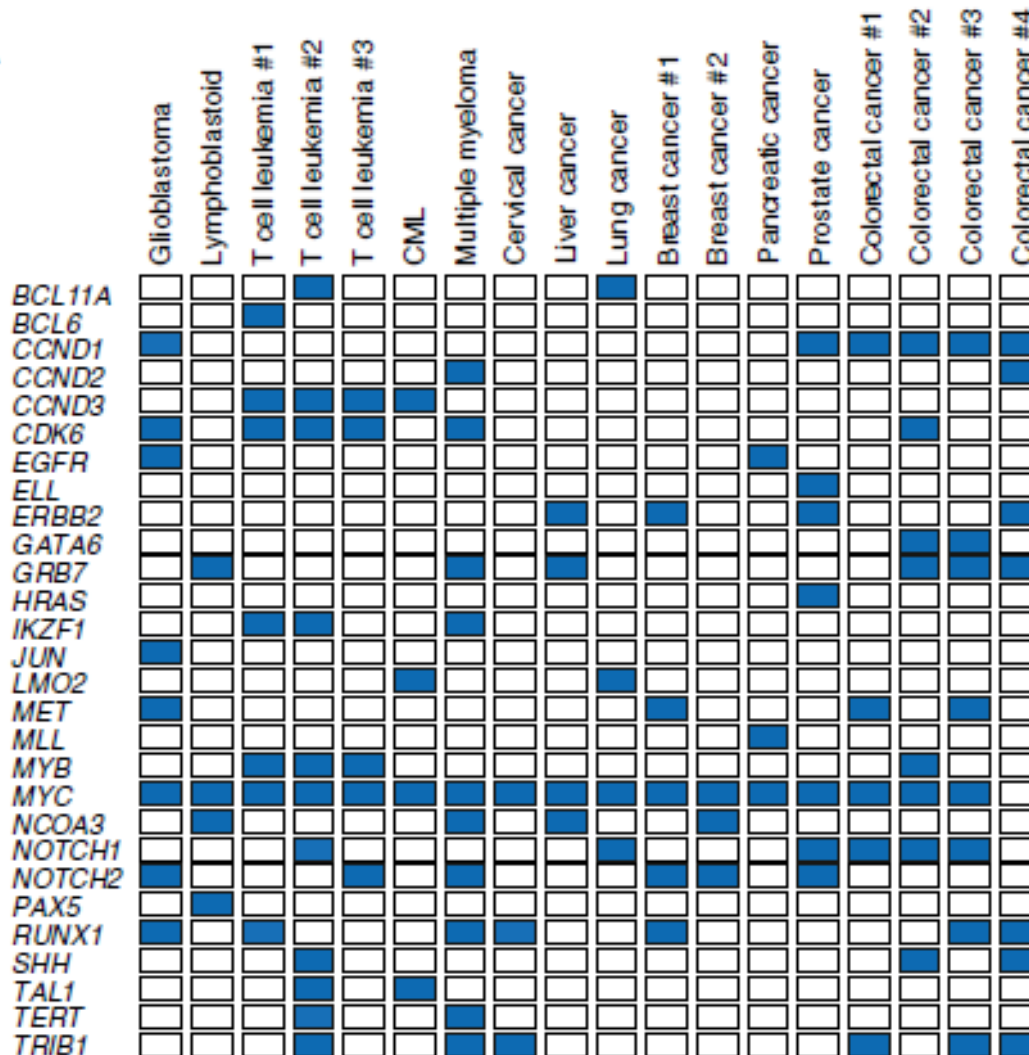
Multiple sclerosis  
(108 non-coding SNP)



# Super-enhancers in Cancer

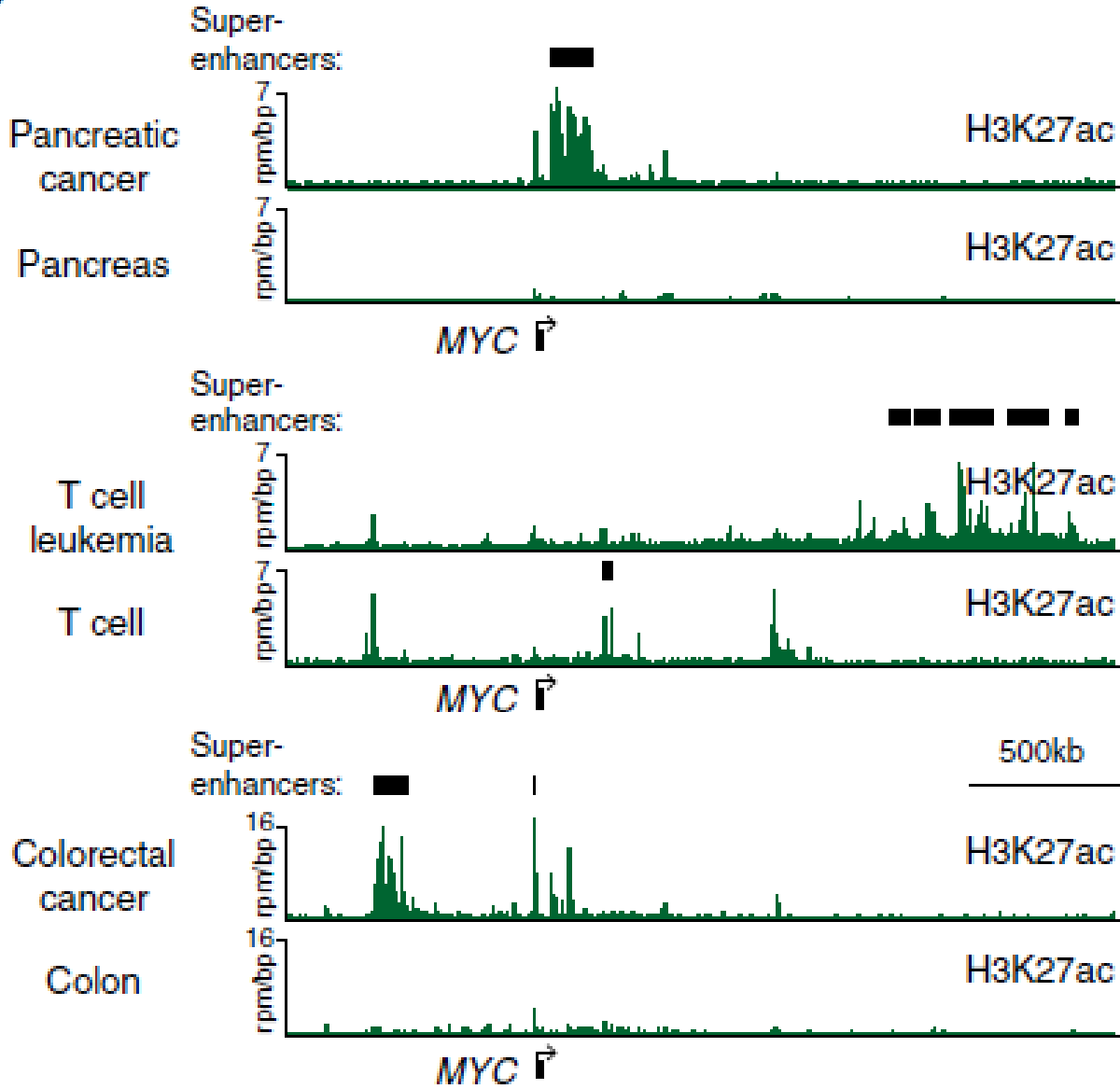
Genes associated with SE and involved in cancer progression

A



# The super-enhancers formed in the MYC locus were tumor type specific

**B**





# Super-enhancers are associated with genes that act as hallmarks in colonrectal cancer

Colorectal cancer

