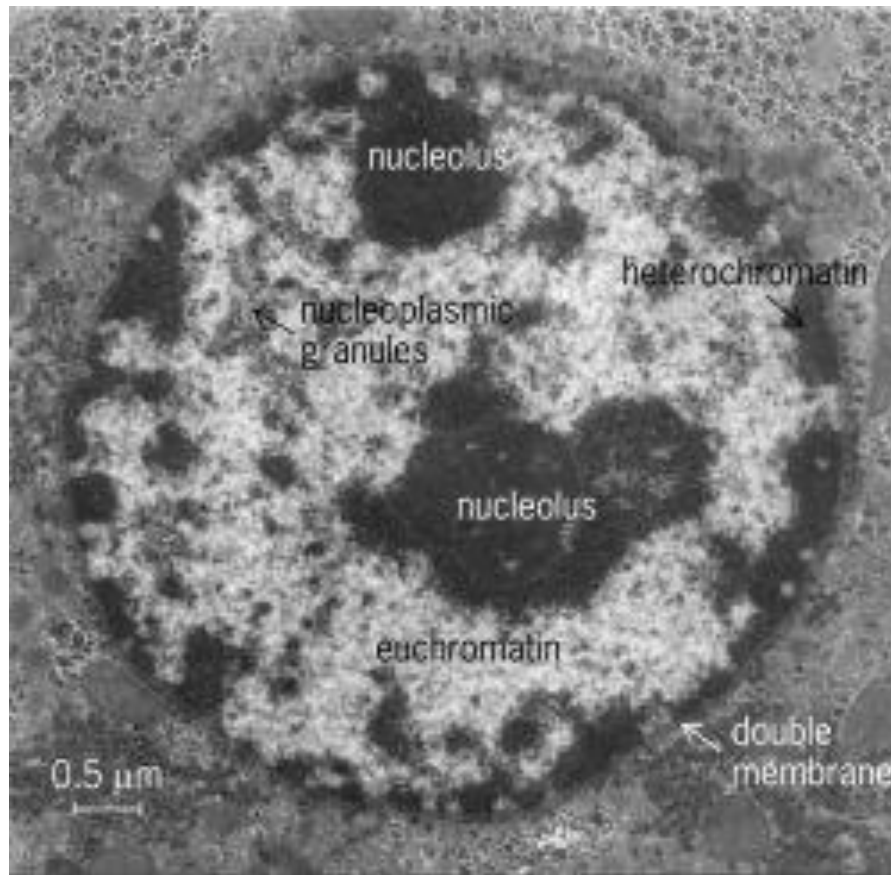


# Ch 1 - L3.1

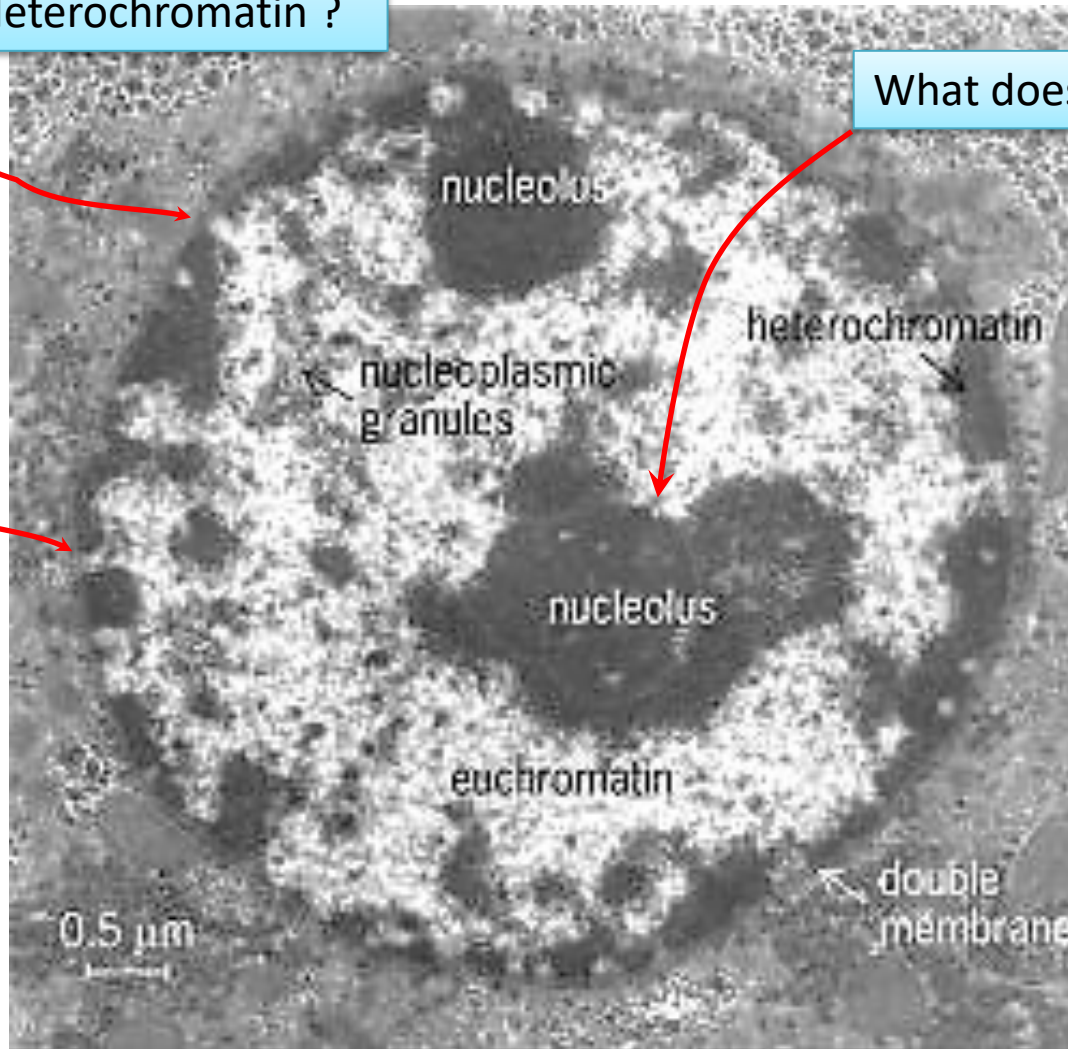
Chromatin Structure  
Nucleosome positioning  
Accessibility

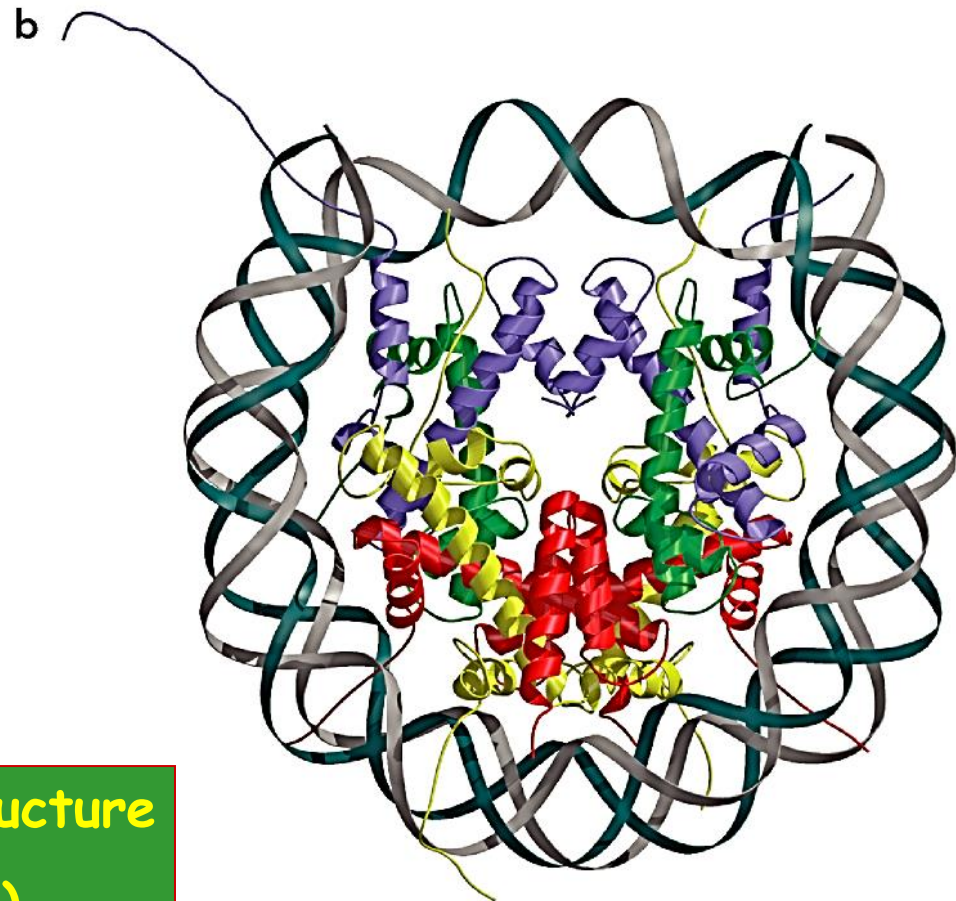
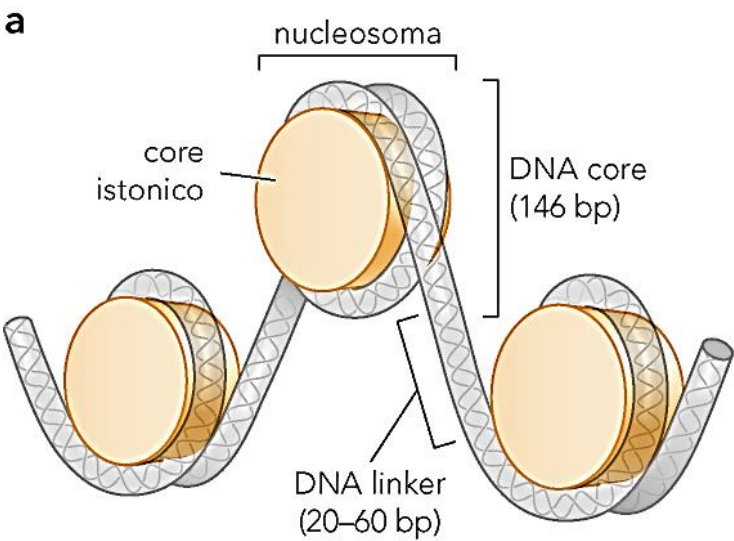


A rat liver cell nucleus at  
Transmission E.M.

What we find in peripheral spaces not occupied by Heterochromatin ?

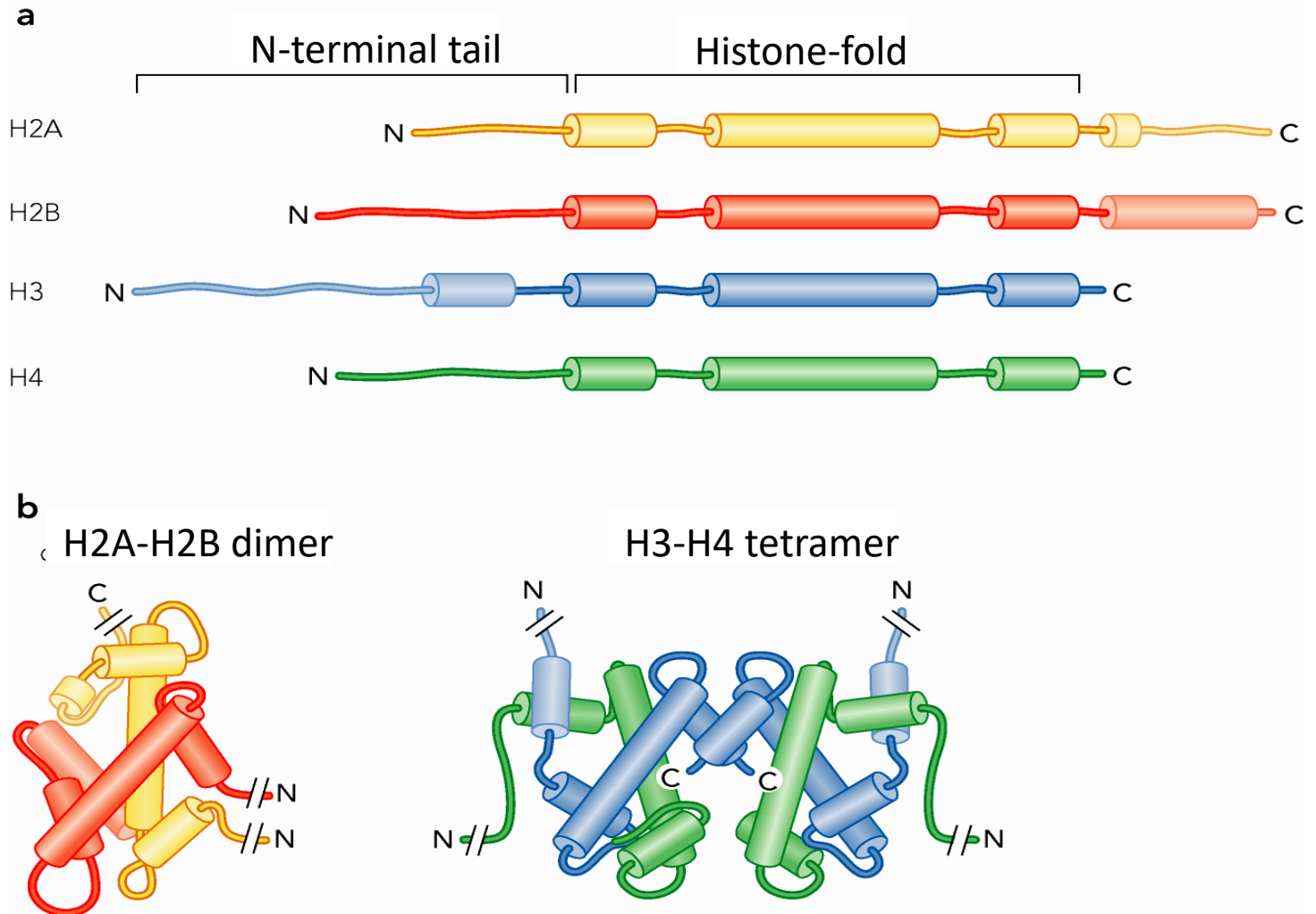
What does the nucleolus do ?

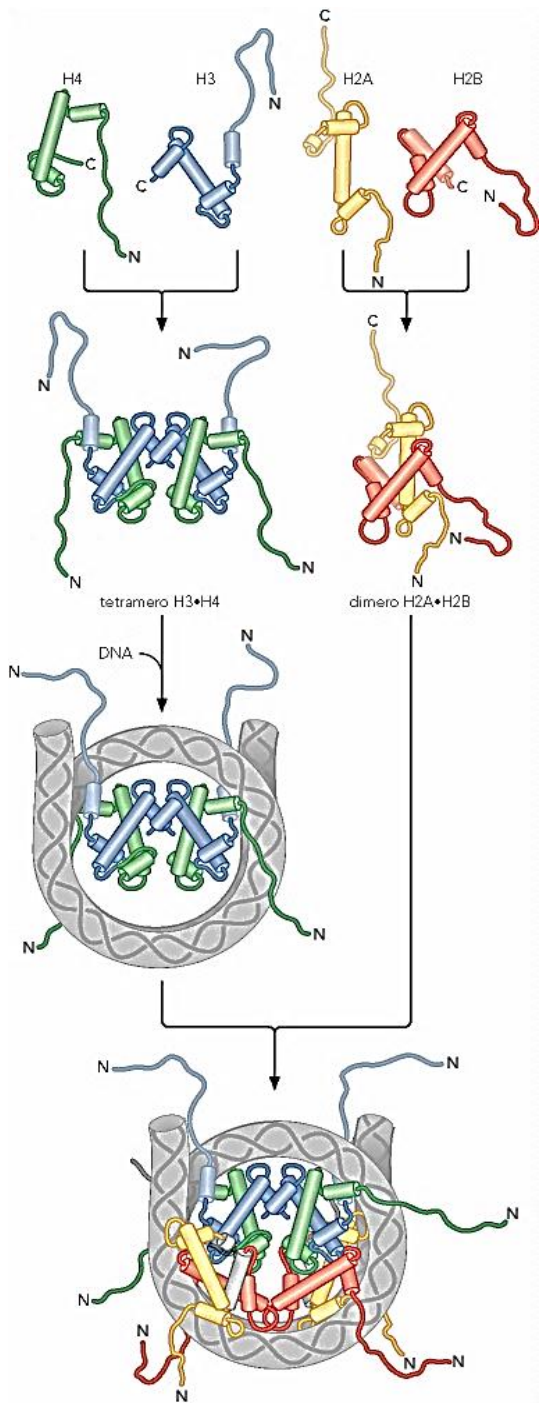




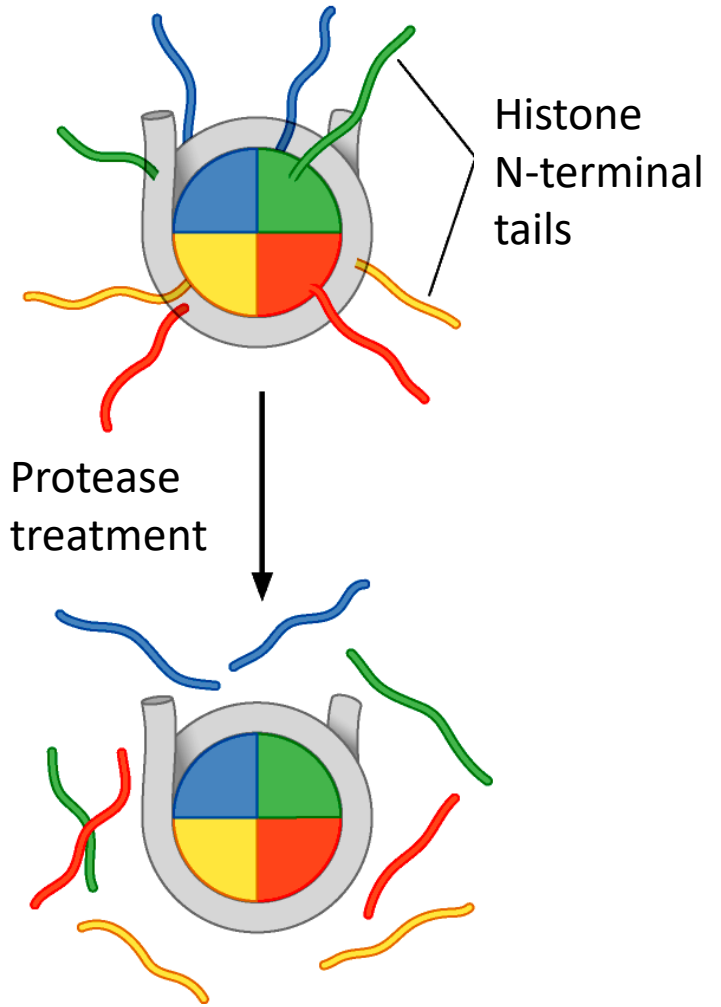
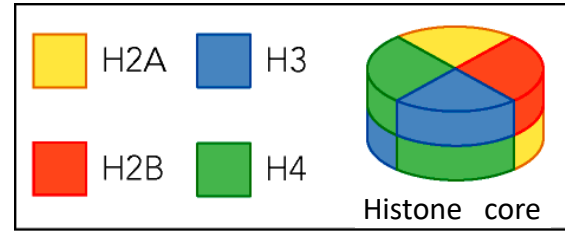
**Nucleosome DNA structure  
topology (supercoiling)  
topoisomerases**

# Histones

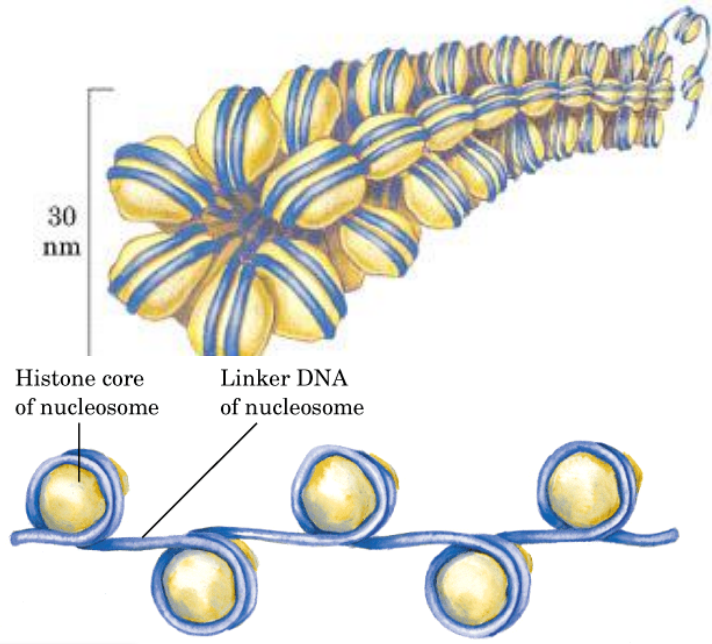
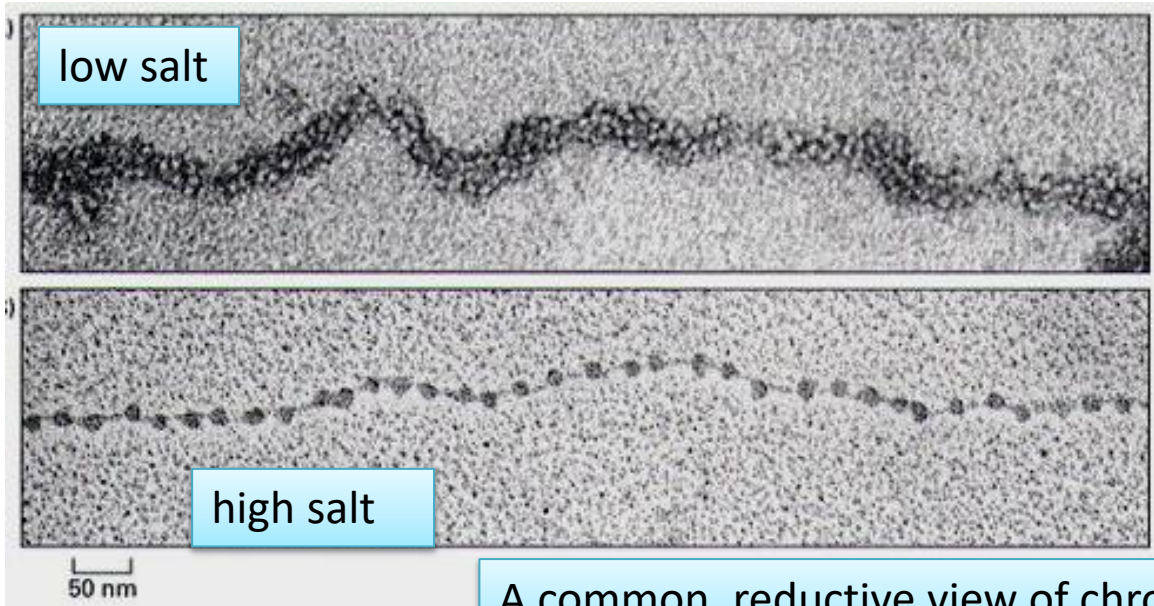




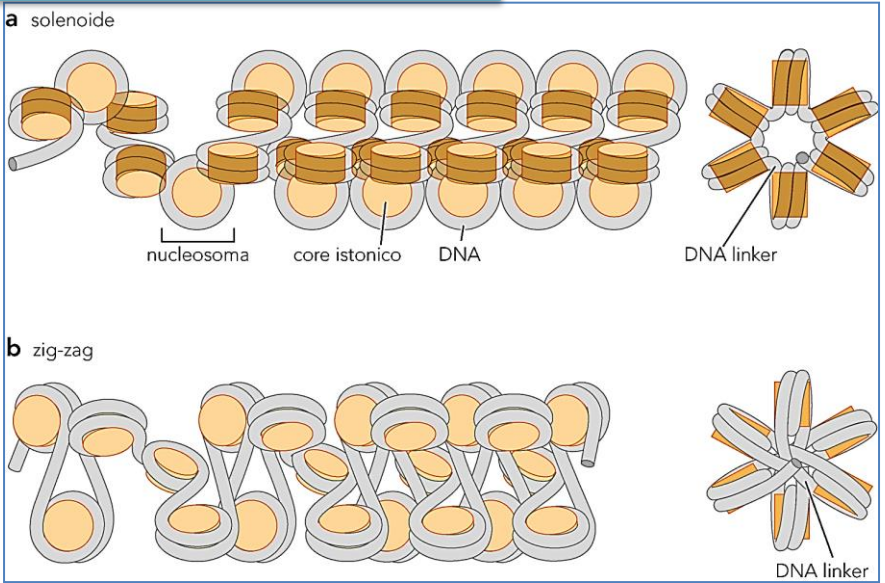
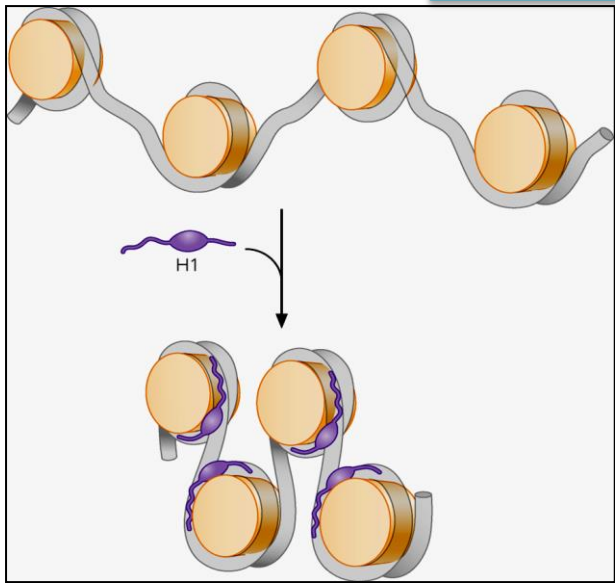
**Nucleosome assembly & N-terminal tails**



# Basic chromatin structure



A common, reductive view of chromatin



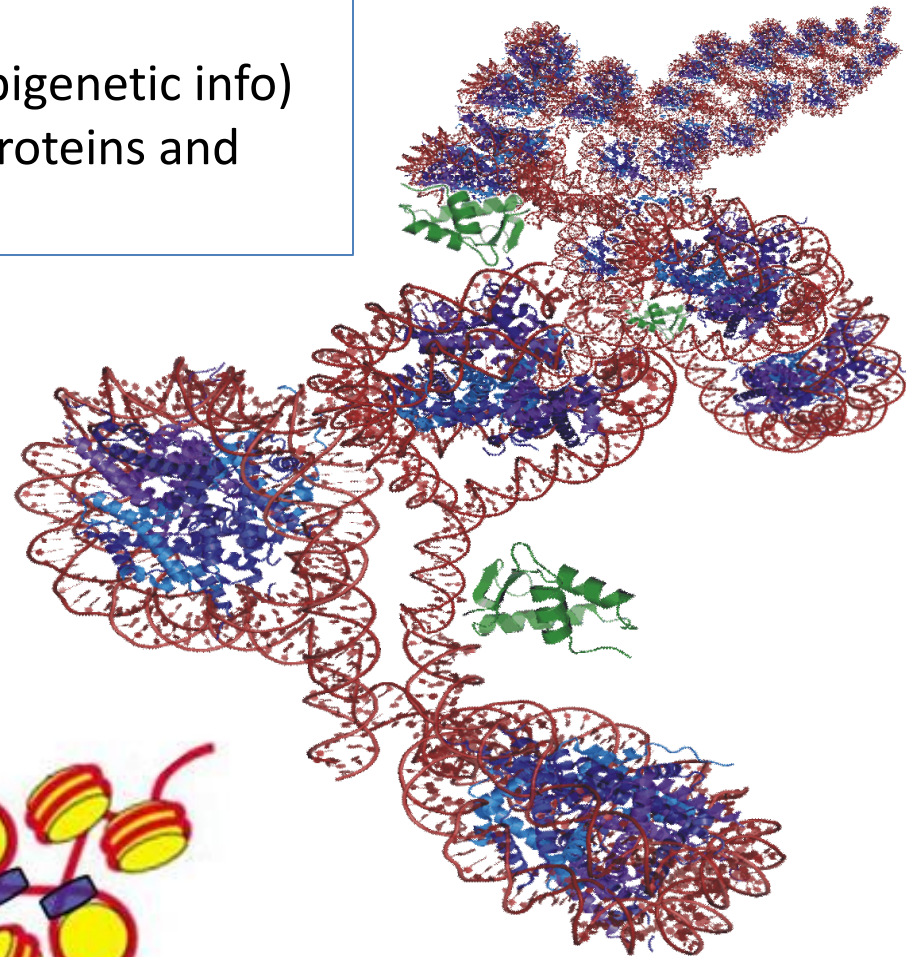
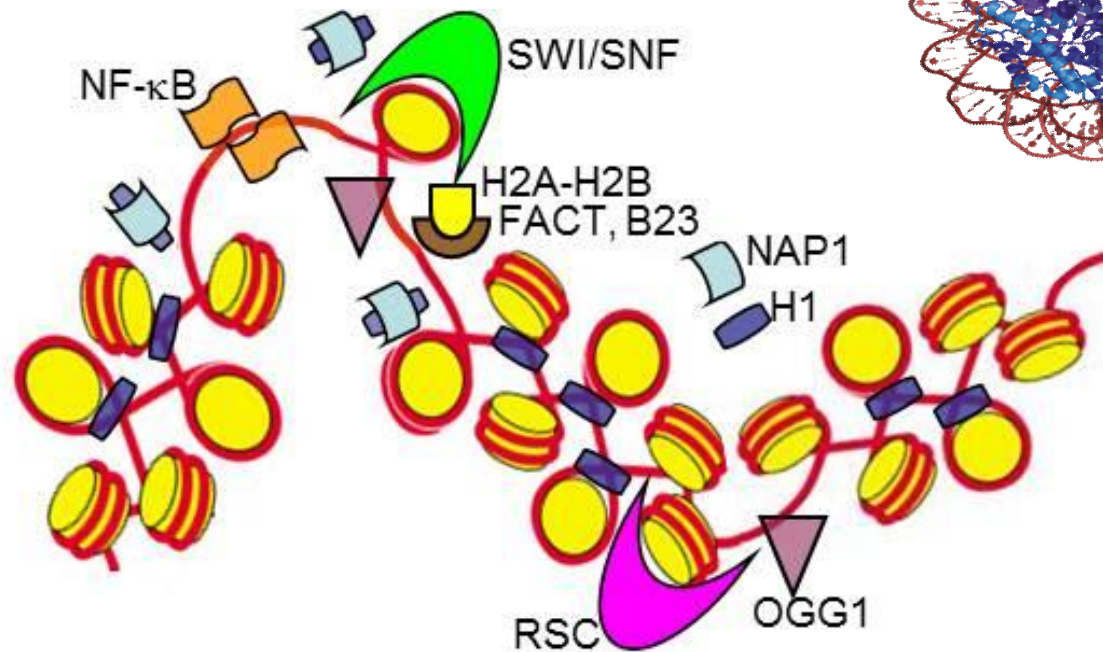
## **Heterochromatin**

- **Facultative**
- **Constitutive at centromeres, telomeres  
( repetitive sequences )**



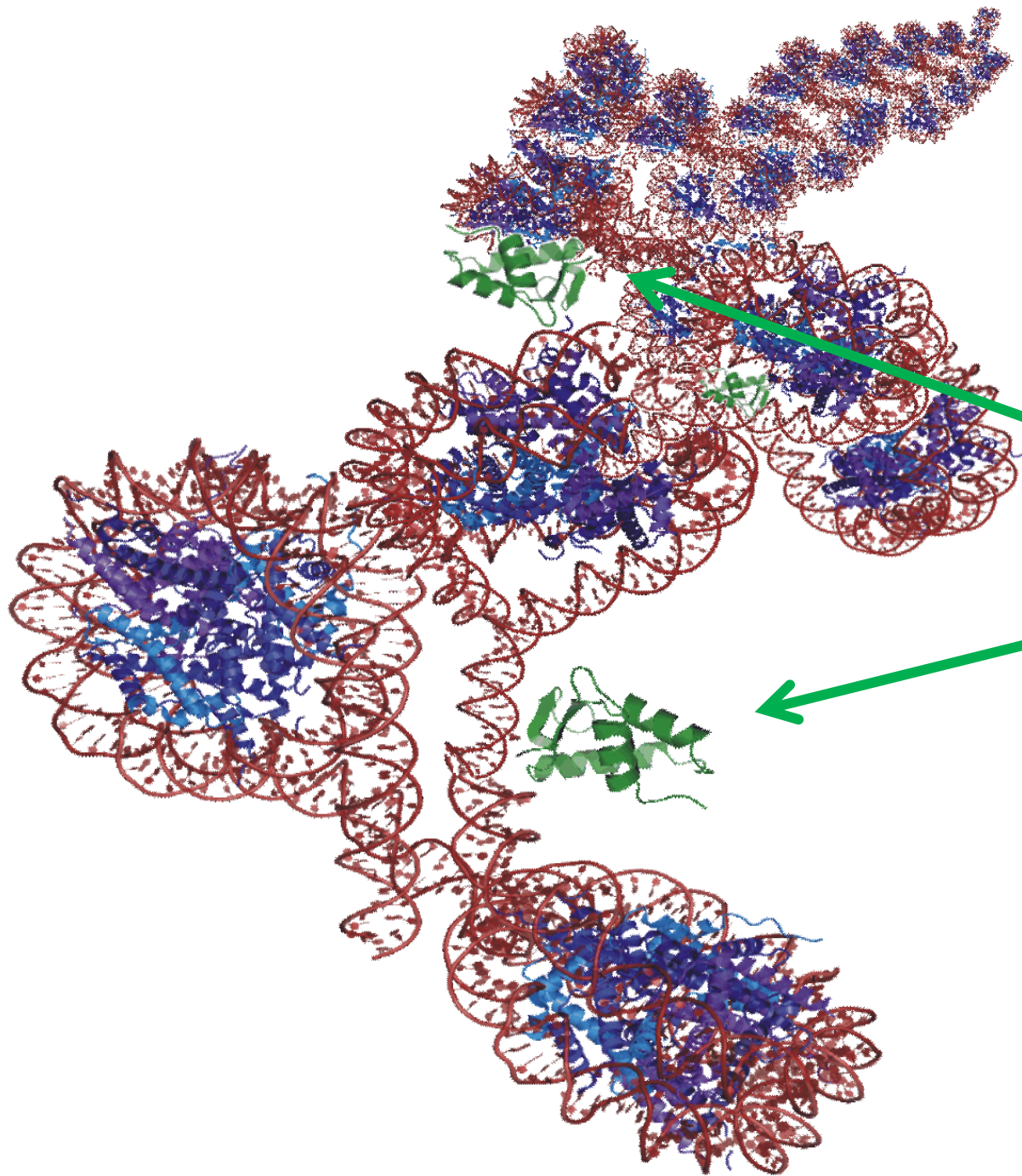
**Chromatin** is composed of

- DNA (genetic information)
- variously modified histone proteins (epigenetic info)
- DNA binding factors, enzymes, other proteins and noncoding RNAs (regulatory).



ENS, LYON

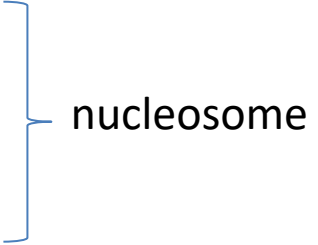
(Proteins in this cartoon are just examples)



Protein factors (e.g. TFs) bind to **chromatin**, not to DNA

## Molecular organization of **heterochromatin** versus **euchromatin**

Features distinguishing HC from EC

1. Accessibility
  2. Nucleosome positioning
  3. Post-translational Modifications to Histones (PTM)
  4. Histone-binding proteins and modifying enzymes
  5. Histone variants
  6. **DNA methylation**
  7. **Noncoding RNA**
  8. **Transcription factors**
- 
- nucleosome

## 1. Accessibility

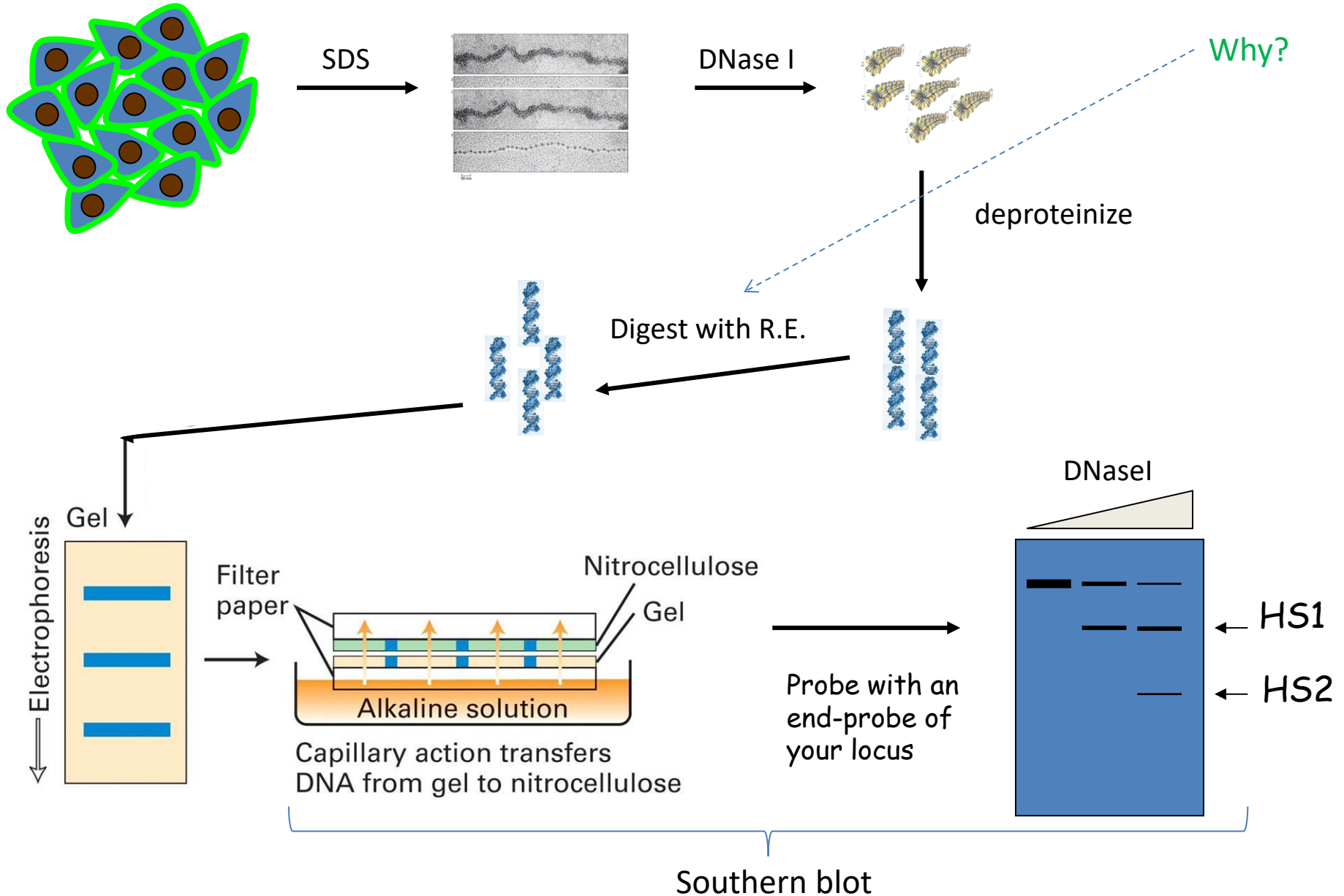
General principle: **enzyme accessibility**.

Dnase I assay → Dnase HS sites (hypersensitive)

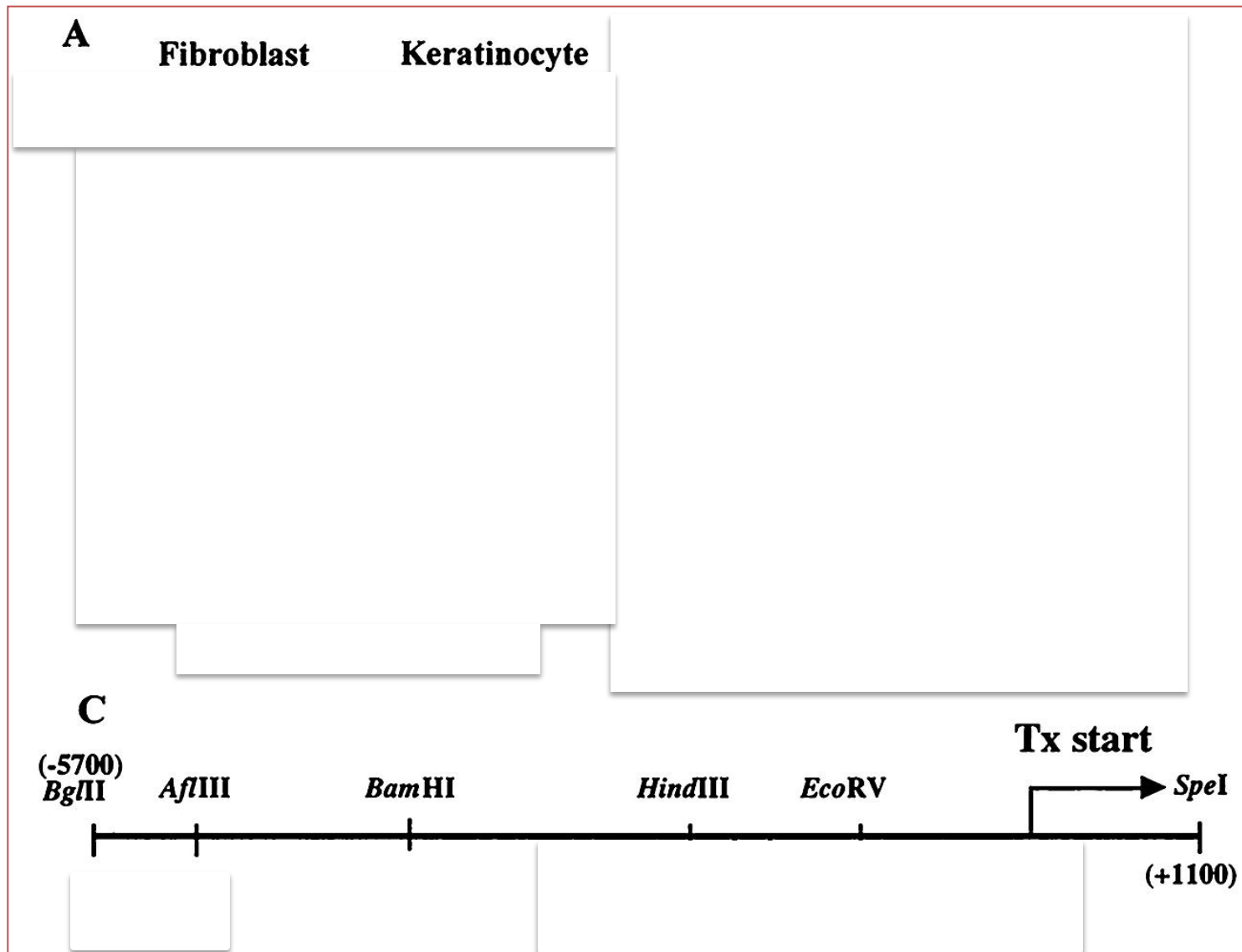
Let's see :

1. a scheme
2. an example taken from a quite old paper.

# The classical assay to detect the gross organization of chromatin at a specified locus: the **DNase I Hypersensitivity Assay**

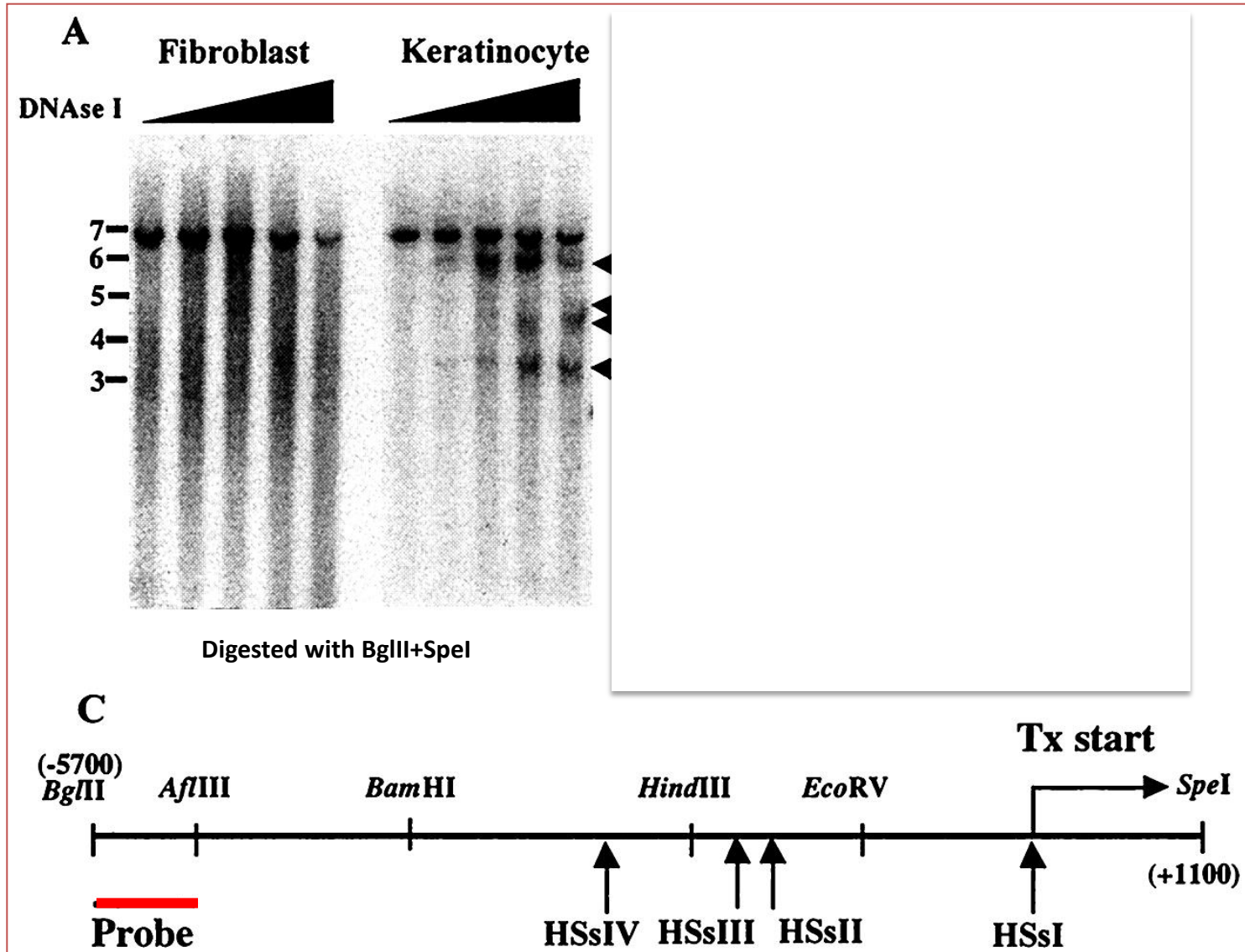


# Keratin K14 gene, analysis of the 5'-flanking region



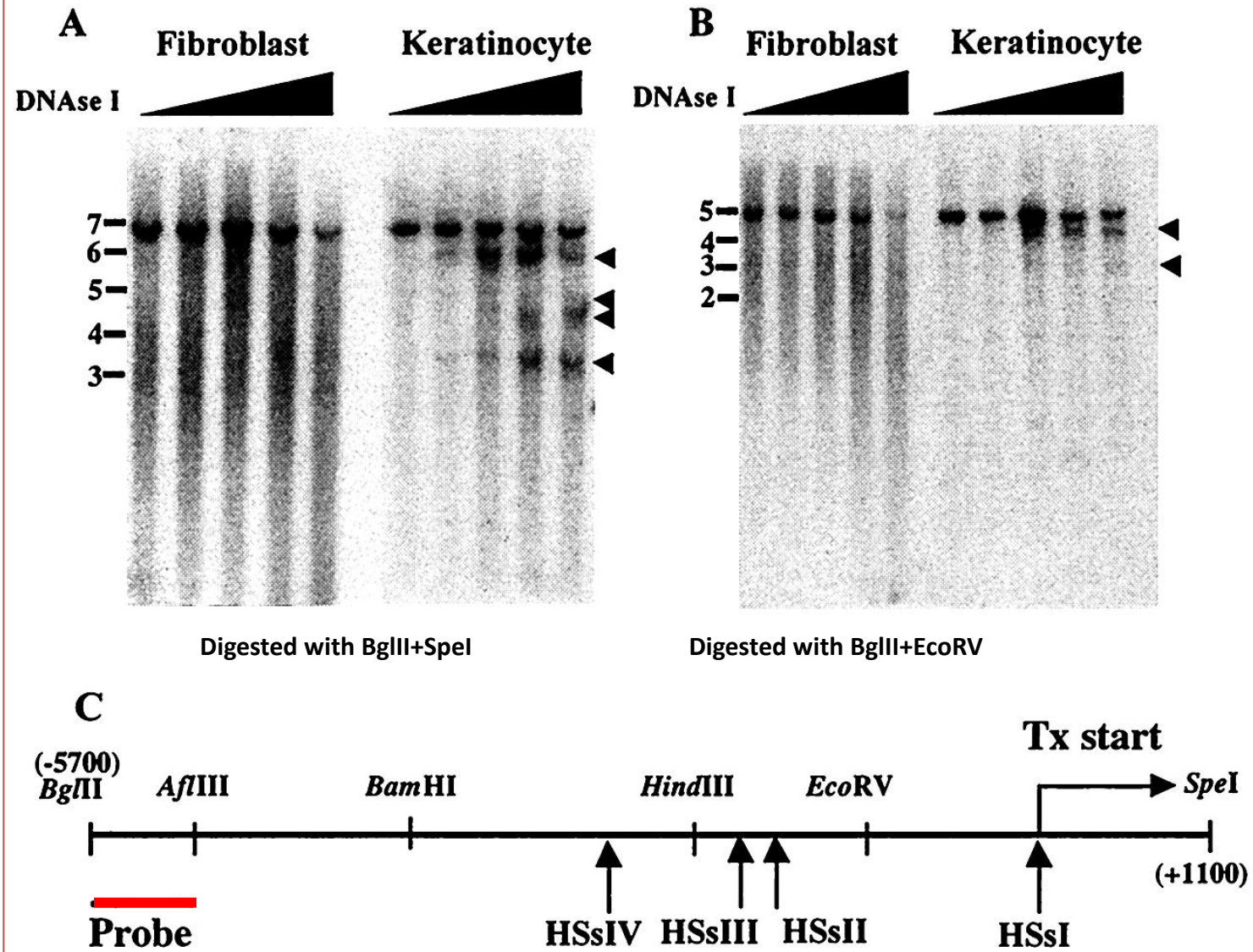
Tratto da: Sinha et al., (2000), Mol Cell Biol, 20: 2543-2555.

# Keratin K14 gene, analysis of the 5'-flanking region



Tratto da: Sinha et al., (2000), Mol Cell Biol, 20: 2543-2555.

# Keratin K14 gene, analysis of the 5'-flanking region



Tratto da: Sinha et al., (2000), Mol Cell Biol, 20: 2543-2555.



If accessibility is not the same in all parts of chromatin, may it depend (also) from different distribution of nucleosomes ?

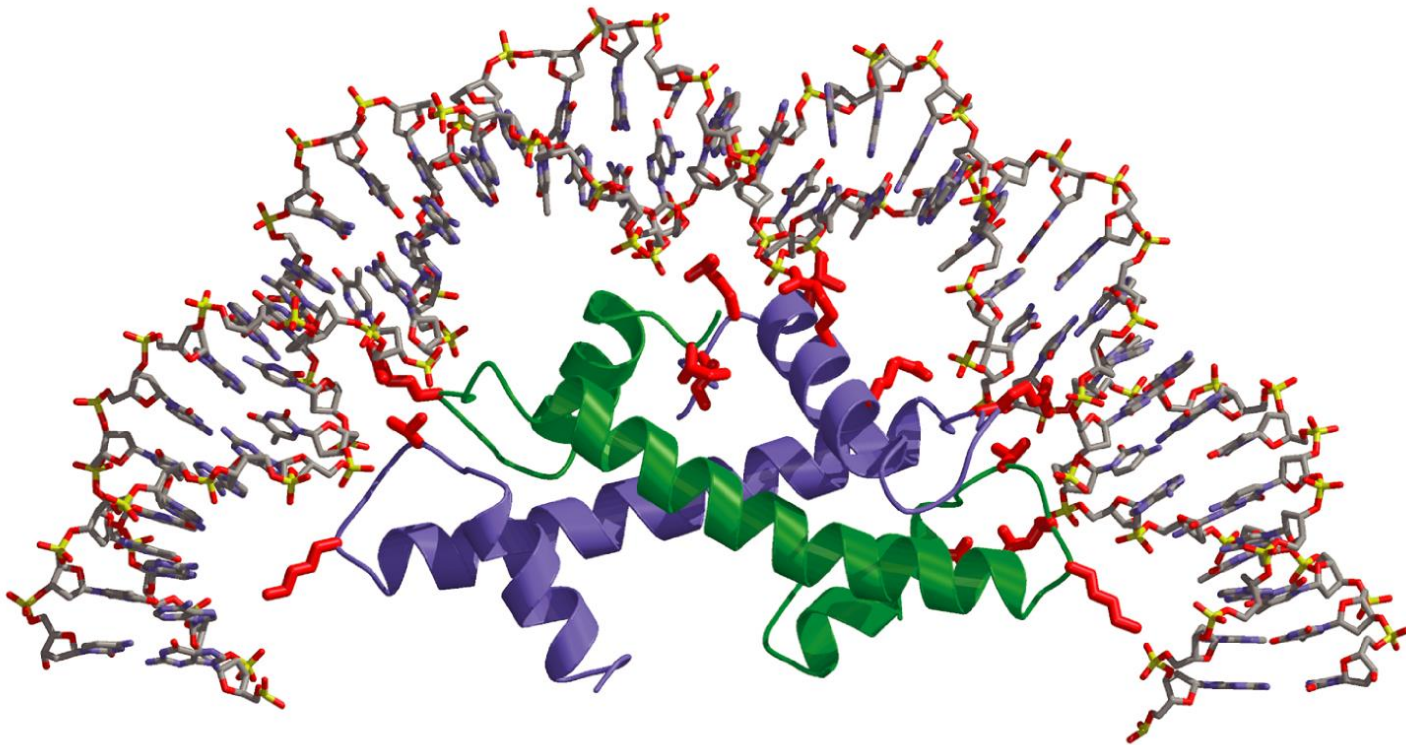
## 2. Nucleosome positioning

Questions that are relevant to genome organization:

- Do nucleosome display preferred positions in the genome ?  
(are they just arrayed regularly spaced as they appear in pictures?)
- Do nucleosomes display any kind of **specificity** for nucleotide sequence?
- Are nucleosome mobile ? Can they translate position over DNA ?  
(are they stable on the same sequence they were assembled to ?)
- Is nucleosome position **functional** to gene/genome regulation ?

«Passive» positioning

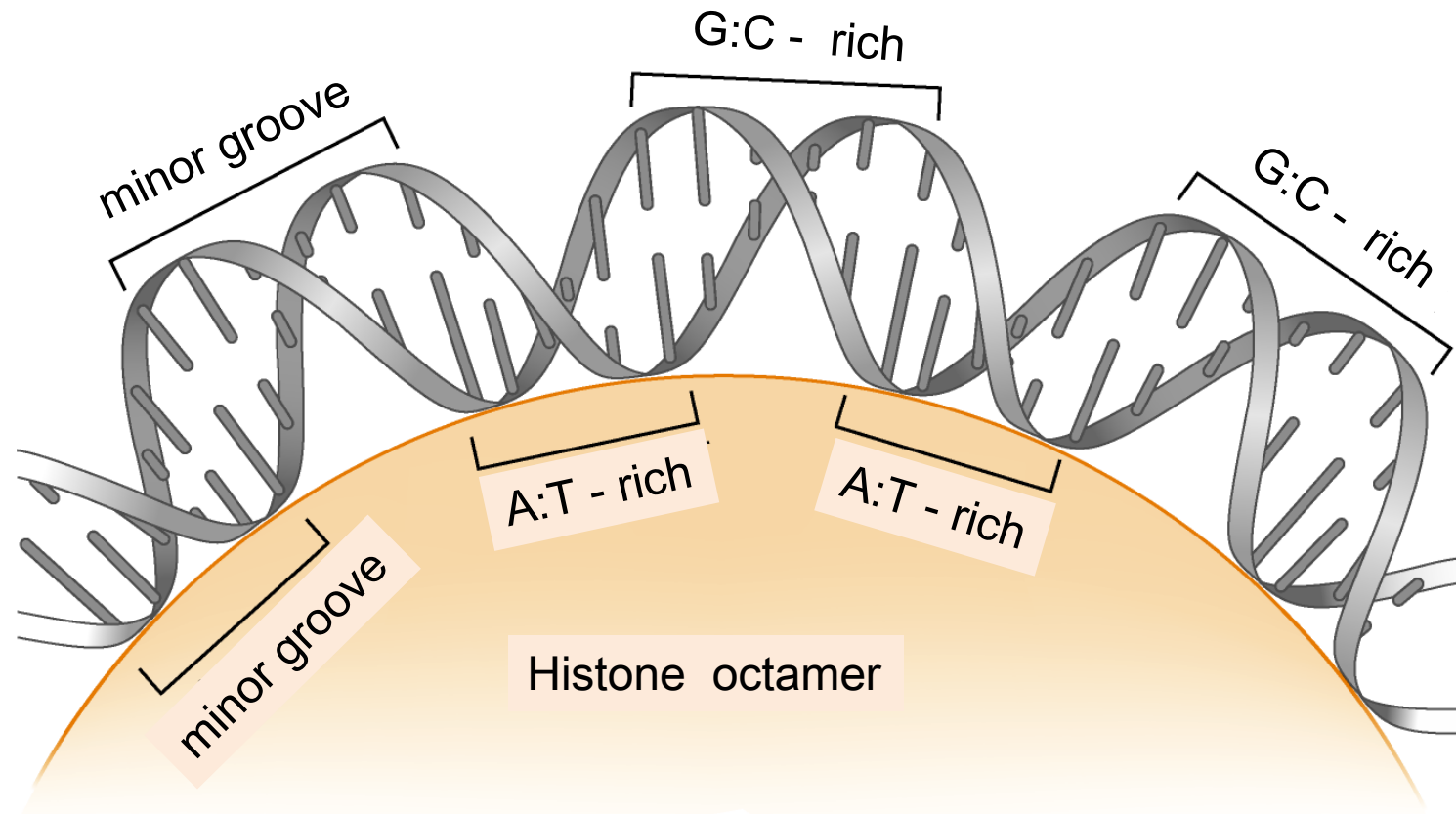
Protein-DNA contacts are mainly due to basic aminoacids – backbone phosphates



## «Passive» positioning

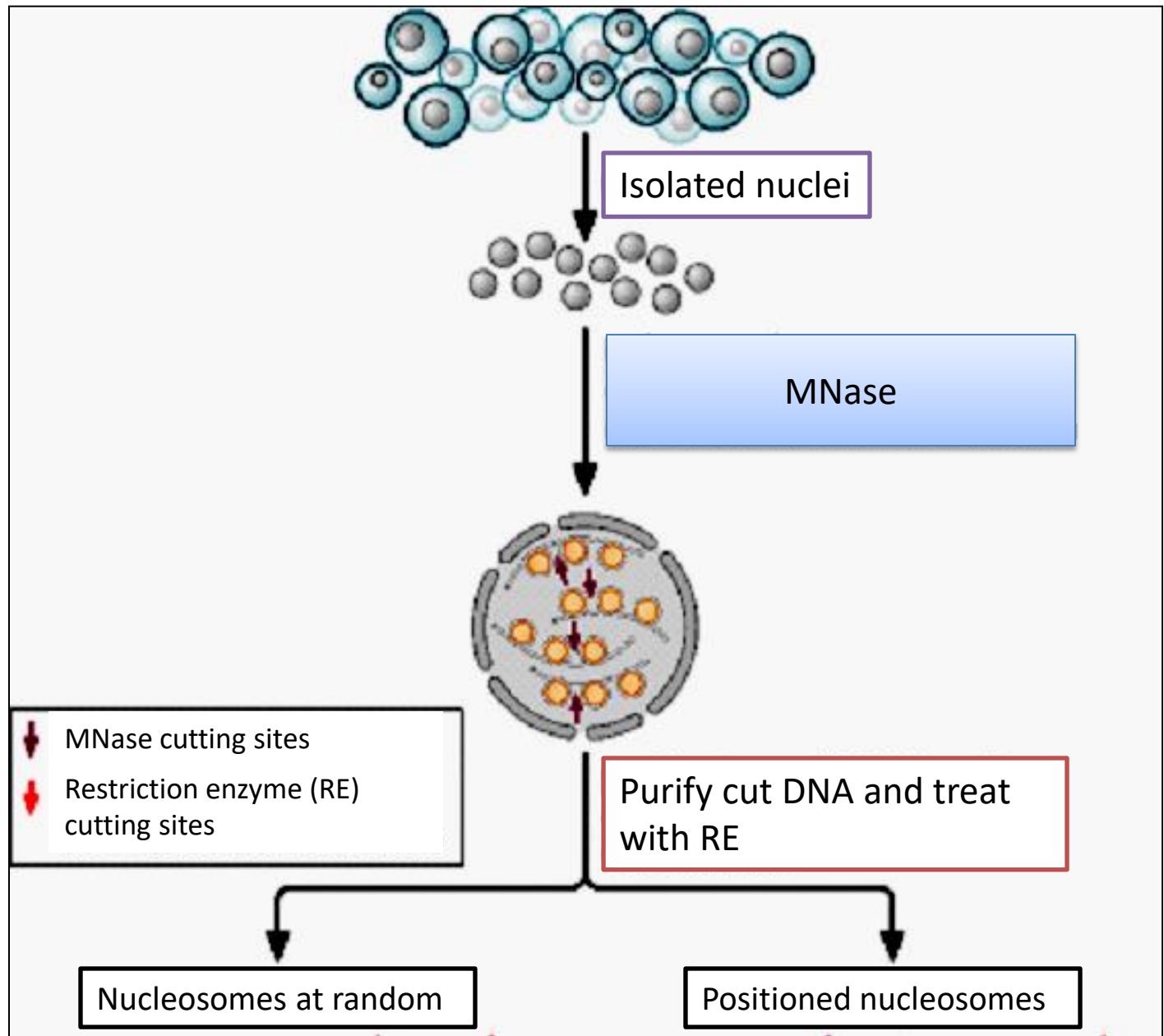
Weak sequence specificity: in general, A/T rich sequences are in touch, whereas C/G rich stay outside.

This is due to intrinsic bending of DNA axis in different sequence contexts.



# Nucleosome positioning analysis

1st step: digestion

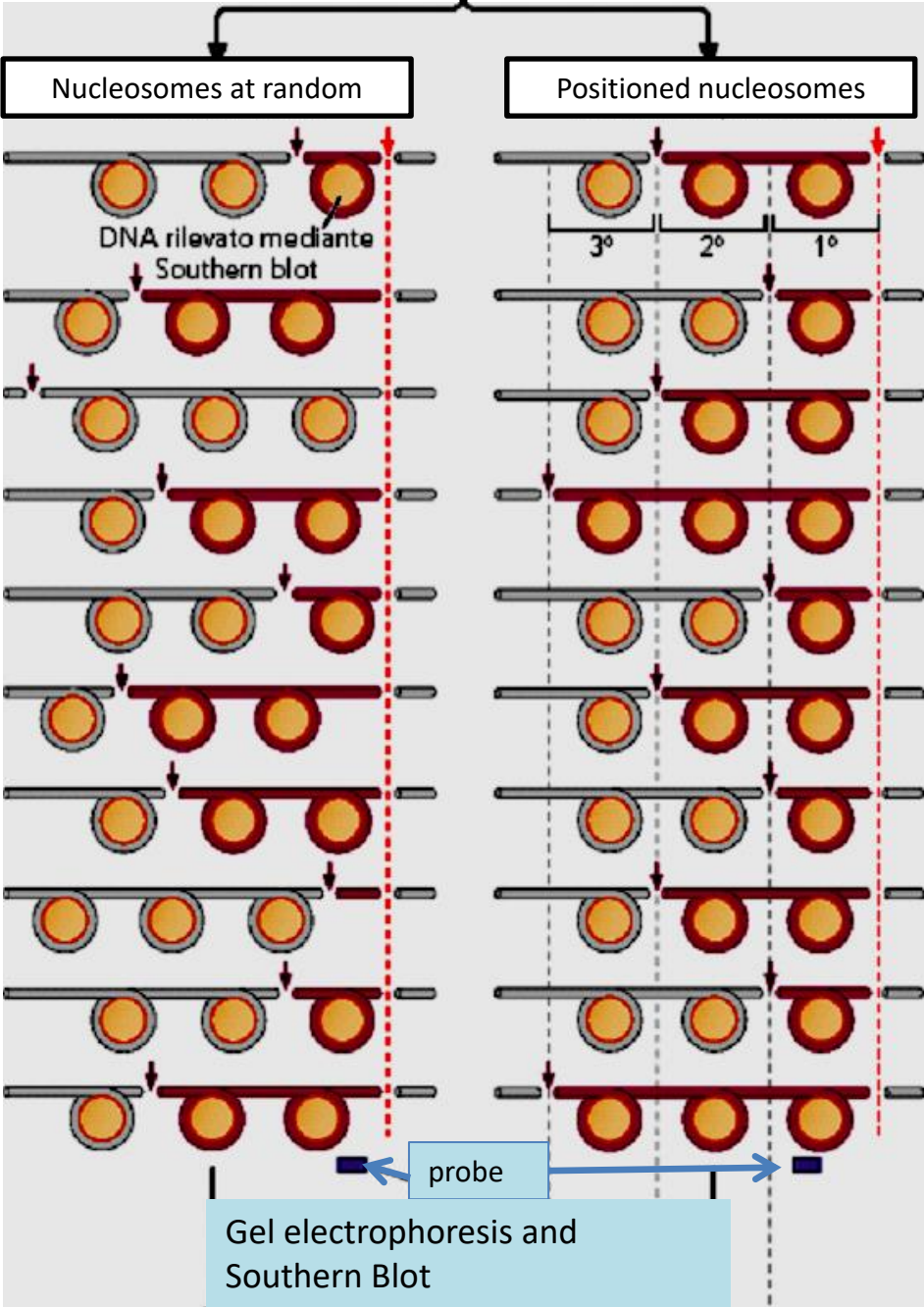


Micrococcal Nuclease I is a mixed endo-exonuclease.

MNase induces single-strand breaks and subsequently double stranded ones by cleaving the complementary strand in close proximity to the first break. MNase continues to digest the exposed DNA ends until it reaches an obstruction, such as a nucleosome or a very stably bound Transcription Factor. In appropriate condition, then, MNase releases fragments of approximately one nucleosome length (~147 bp), which are typically selected for sequencing in MNase-Seq experiments.

2 different cases:  
mobile nucleosomes  
*versus*  
positioned nucleosomes

each row is one  
molecule of DNA  
extracted from cells



↓ MNase cuts  
↓ R.E. cuts

Gel electrophoresis and  
Southern Blot

mobile

versus

positioned

