Building blocks and primary structure

# Why is the structure so important?

- Interpretation of the mechanism of function of individual proteins
- 2. Approaches to the "protein folding problem"
- 3. Patterns of molecular evolution
- 4. Predictions of the structures of closely related proteins homology modelling
- **5**. Protein engineering:
  - Modifications to probe mechanisms of function
  - 2. Attempts to enhance thermostability
  - **3.** Clinical applications
- 6. Drug design

### Sequence = specific folding

Is Translated to a Sequence of

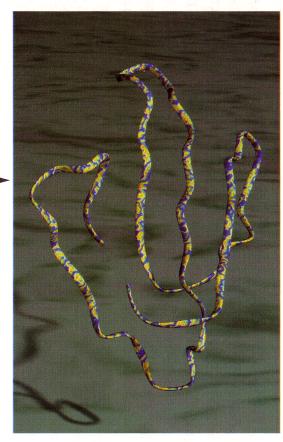
Amino Acids in a Protein...

A Sequence of Bases in DNA...

one strand **Triplets of bases read from** 

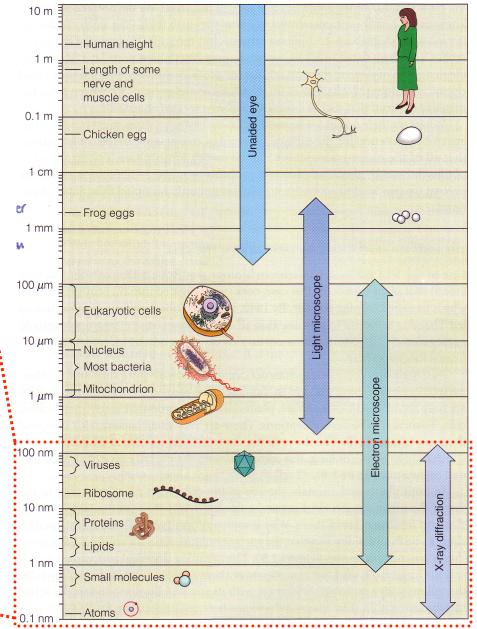
**Three Bases** UUU F UCU S UAU Y UGU C UUC F UCC S UAC Y UGC C UCA S UUA L **UAA Stop UGA Stop** UCG S UUG L **UGG W UAG** Stop CCU P CUU L CAU H CGU R CUC L CCC P CAC H CGC R CCA P CUA L CAA Q CGA R CCG P CUG L CAG Q CGG R ACU T AUU I AAU N AGU S ACC T AAC N AGC S AUC I ACA T AAA K AUA I AGA R ACG T AAG K AGG R AUG M GCU A GUU V GAU D **GGU G** GCC A GAC D **GGC G** GUC V GUA V GCA A GAA E **GGA G GUG V** GCG A GAG E **GGG G One Amino Acid Genetic Code** 'Translation Table'

Which Folds Spontaneously to a Precise Three-Dimensional Structure

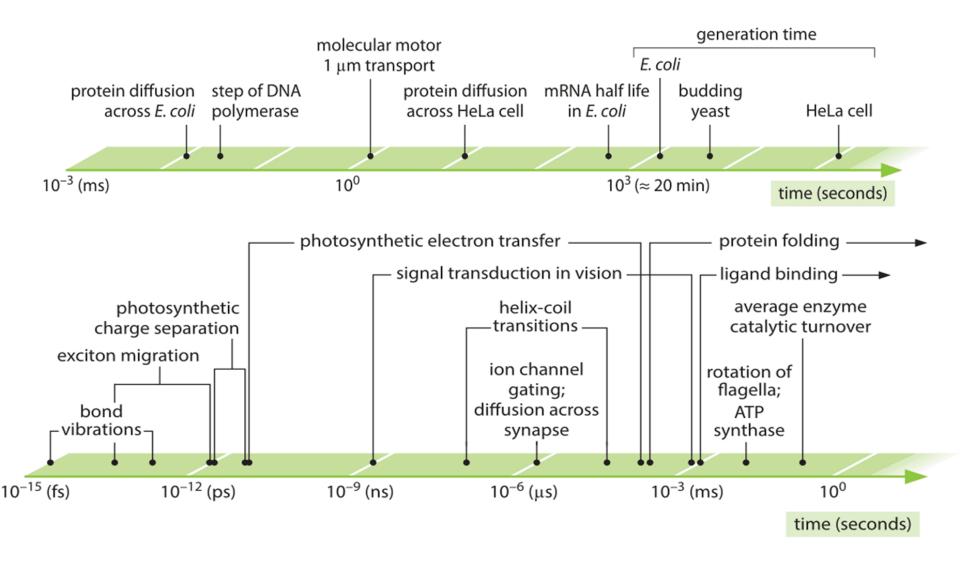


## Range and sizes in biochemistry

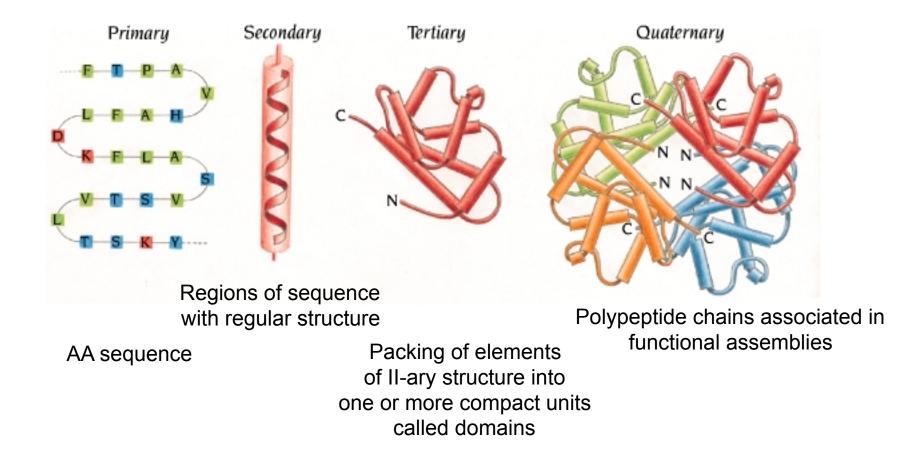
- Techniques for study in biochemistry / structural biology:
  - Cryo-electron microscopy (Cryo-EM)
  - Atomic Force Microscopy (AFM) and Scanning Tunneling Microscopy (STM)
  - Crystallography and X-ray diffraction
  - Nuclear Magnetic Resonance (NMR)
  - Circular dichroism
  - Fluorescence
  - Raman scattering, Electron spin resonance, Mossbauer spectroscopy, Infra-red spectroscopy

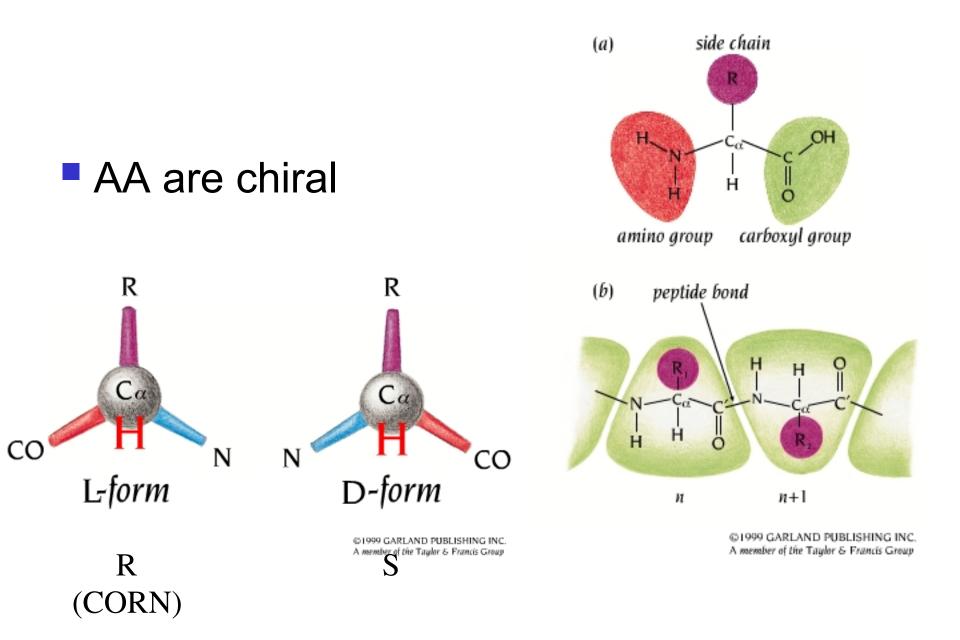


## Time in biochemistry

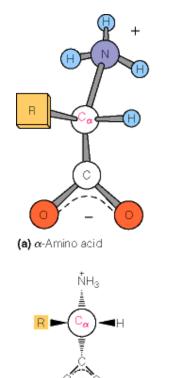


# Proteins are polymers of aminoacids

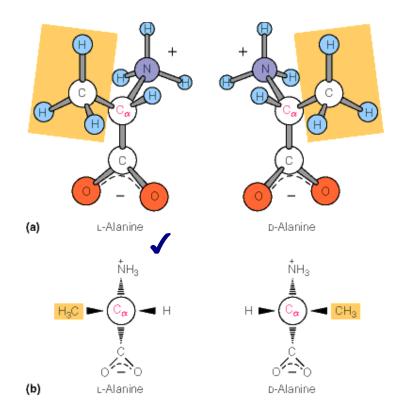




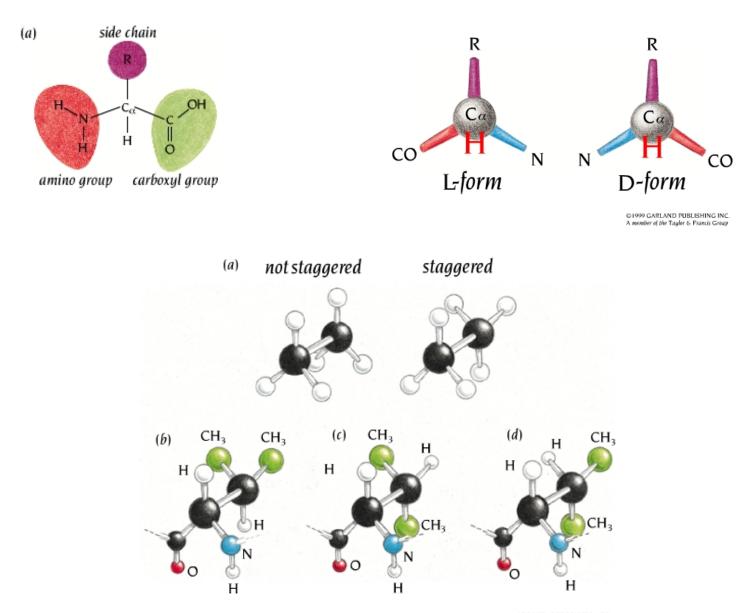
# Stereochemistry of $\alpha$ -amino acids



(b) Compact representation



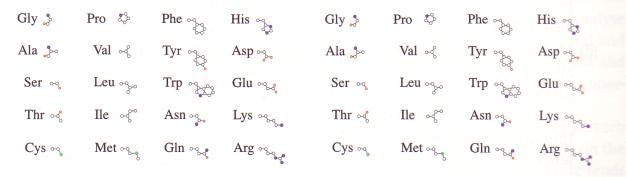
#### Aminoacids: classification and properties.



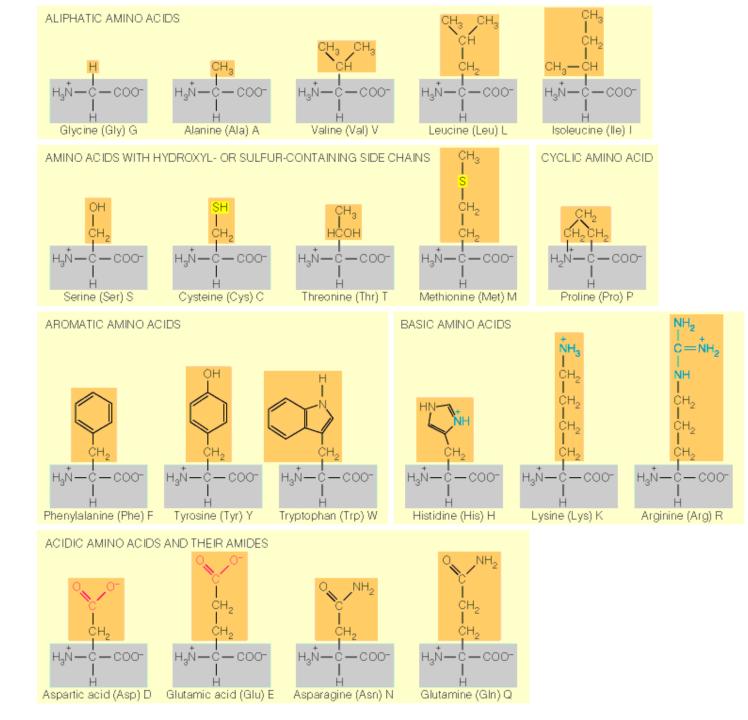
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# Different side-chains = different properties

#### I letter and 3 letters codes

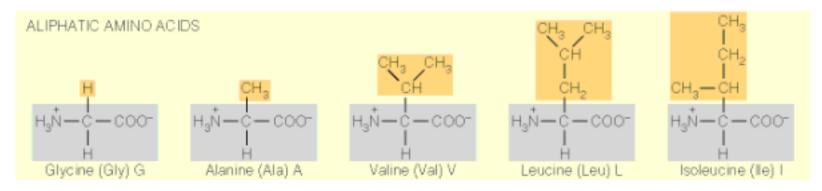


Gly 🗗 Pro 🚫 Phe 📢 His 🥠 Gly 🕐 Pro 🛇 Phe 📢 His 📢 Ala 🚺 Val 🕻 Tyr 📢 Asp 📢 Ala 🚺 Val 🕻 Tyr 🏠 Asp 😱 Ser 🗘 Leu 🚺 Trp 🕵 Glu 😍 Ser 🗘 Leu 🚺 Trp 🕵 Glu 🔍 Thr C Ile C Asn Lys C Thr C Ile C Asn 😱 Lys ( Cys 🜔 Met 🜓 Gln 🔍 Arg 🔍 Cys 😱 Met C Gln 🕘 Arg 🤇



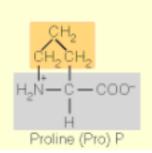
## Properties of aa side-chains

#### Aliphatic aa:



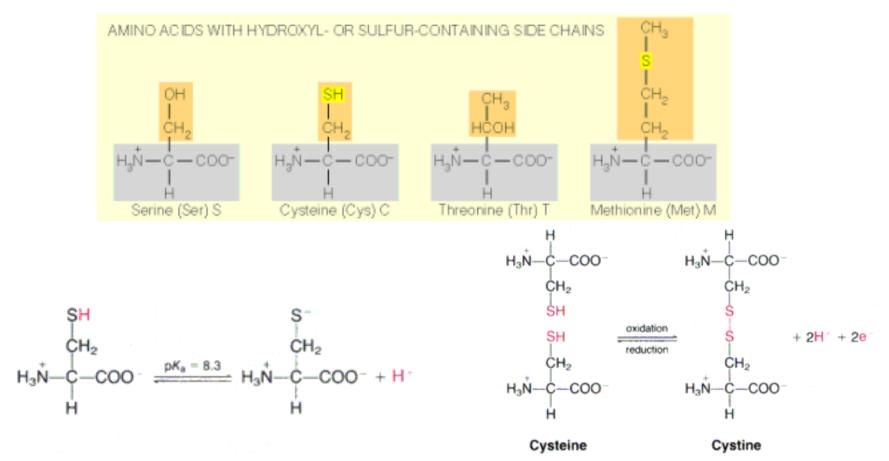
- Flexible and hydrophobic, the most lle;
- Found in the hyodrophobic core of proteins
- Pro: exception, rigid ring as side chain, often a structure breaker

#### CYCLIC AMINO ACID

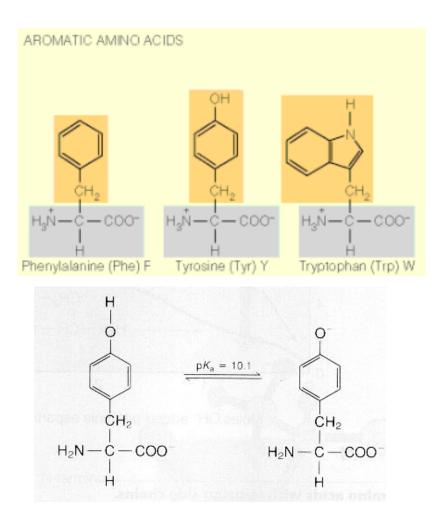


## Hydroxyl-, sulfur- side chains

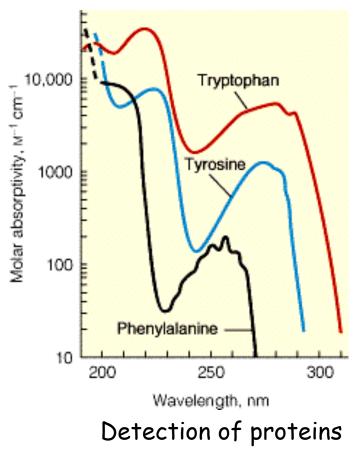
#### Weakly polar side chains



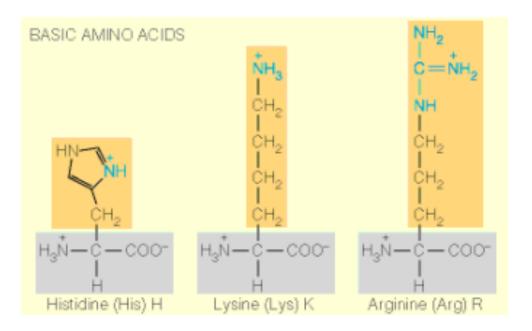
#### **Aromatic side-chains**

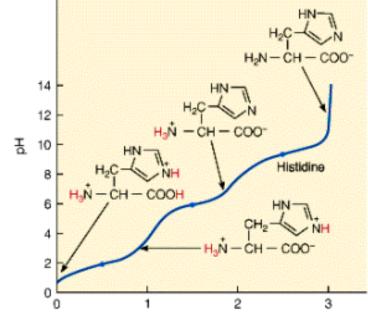


#### Absorbance



### Basic side-chains





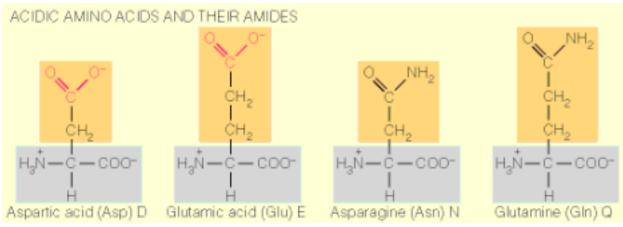
(a)

Moles OH<sup>-</sup> added per mole histidine

Group Type	Typical p <i>K</i> a Range <sup>a</sup>
α-Carboxyl	3.5-4.0
Side chain carboxyls	4.0-4.8
of aspartic and	
glutamic acids Imidazole (histidine)	6.5-7.4
Cysteine (-SH)	8.5-9.0
Phenolic (tyrosine)	9.5-10.5
α-Amino	8.0-9.0
Side chain amino (lysine)	9.8-10.4
Guanidinyl (arginine)	~12

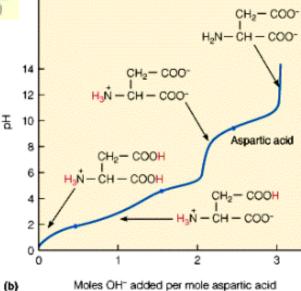
<sup> $^{\circ}$ </sup>Values outside these ranges are observed. For example, side chain carboxyls have been reported with  $pK_a$  values as high as 7.3.

# Acidic side-chains and amides



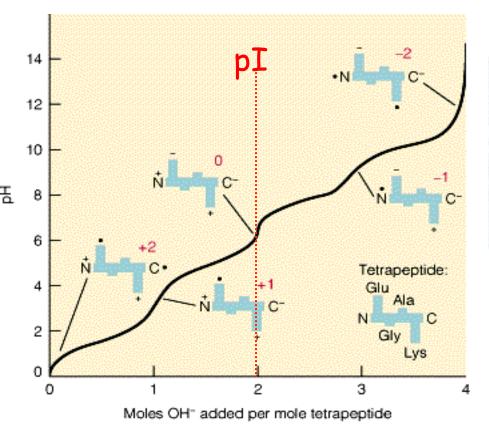
Group Type	Typical pK <sub>a</sub> Range <sup>a</sup>
$\alpha$ -Carboxyl	3.5-4.0
Side chain carboxyls	4.0 - 4.8
of aspartic and	
glutarnic acids	
Imidazole (histidine)	6.5-7.4
Cysteine (—SH)	8.5-9.0
Phenolic (tyrosine)	9.5-10.5
α-Amino	8.0-9.0
Side chain amino	9.8-10.4
(lysine)	
Guanidinyl (arginine)	~12

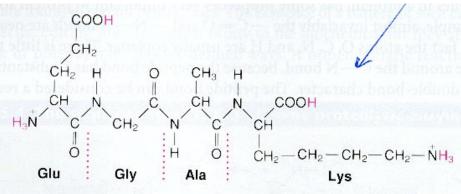
<sup>e</sup>Values outside these ranges are observed. For example, side chain carboxyls have been reported with pK<sub>a</sub> values as high as 7.3.



### Proteins are polyampholites

#### pl: isoelectric point



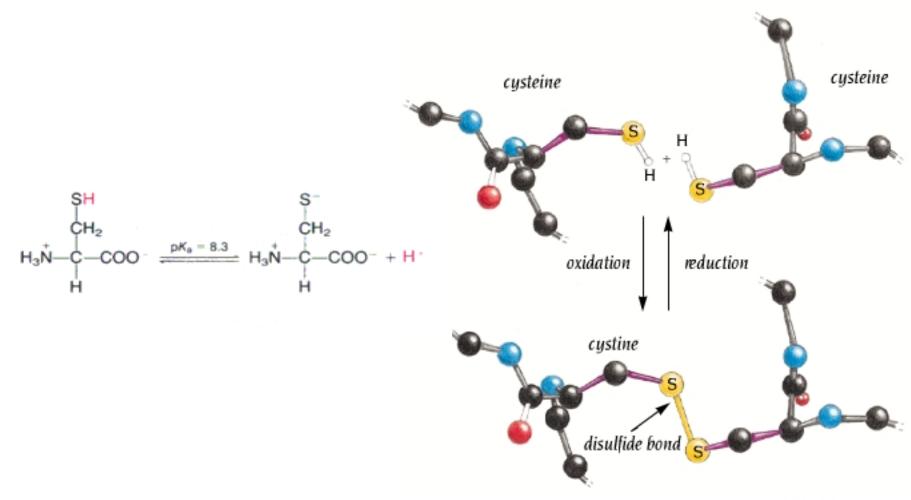


Name	Abbreviations	pK <sub>a</sub> of α-COOH Group	$pK_a$ of $\alpha$ -NH <sub>3</sub> <sup>+</sup> Group	pK <sub>a</sub> of Ionizing Side Chain"	Residue <sup>b</sup> Mass (daltons)	Occurrence <sup>c</sup> in Proteins (mol %)
Alanine	A, Ala	2.3	9.7	_	71.08	9.0
Arginine 米	R, Arg	2.2	9.0	12.5 🗮	156.20	4.7
Asparagine	N, Asn	2.0	8.8	_	114.11	4.4
Aspartic acid 米	D, Asp	2.1	9.8	3.9 🗮	115.09	5.5
Cysteine 💥	C, Cys	1.8	10.8	8.3	103.14	2.8 🗮
Glutamine	Q, Gln	2.2	9.1	_	128.14	3.9
Glutamic acid	E, Glu	2.2	9.7	4.2	129.12	6.2
Glycine	G, Gly	2.3	9.6	—	57.06	7.5
Histidine 🗡	H, His	1.8	9.2	6.0	137.15	2.1 🗮
Isoleucine	I, Ile	2.4	9.7	—	113.17	4.6
Leucine	L, Leu	2.4	9.6	_	113.17	7.5
Lysine	K, Lys	2.2	9.0	10.0	128.18	7.0
Methionine	M, Met	2.3	9.2	_	131.21	1.7
Phenylalanine	F, Phe	1.8	9.1	_	147.18	3.5
Proline	P, Pro	2.0	10.6	_	97.12	4.6
Serine	S, Ser	2.2	9.2	—	87.08	7.1
Threonine	T, Thr	2.6	10.4	_	101.11	6.0
Tryptophan 💥	W, Trp	2.4	9.4	_	186.21	1.1 💥
Tyrosine	Y, Tyr	2.2	9.1	10.1	163.18	3.5
Valine	V, Val	2.3	9.6	—	99.14	6.9

"Approximate values found for side chains on the free amino acids.

<sup>b</sup> To obtain the mass of the amino acid itself, add the mass of a mole of water, 18.02 g. The values given are for neutral side chains; slightly different values will apply at pH values where protons have been gained or lost from the side chains.

#### $2(-CH_2-SH) + 1/2O_2 \Leftrightarrow -CH_2-S-S-CH_2 + H_2O$



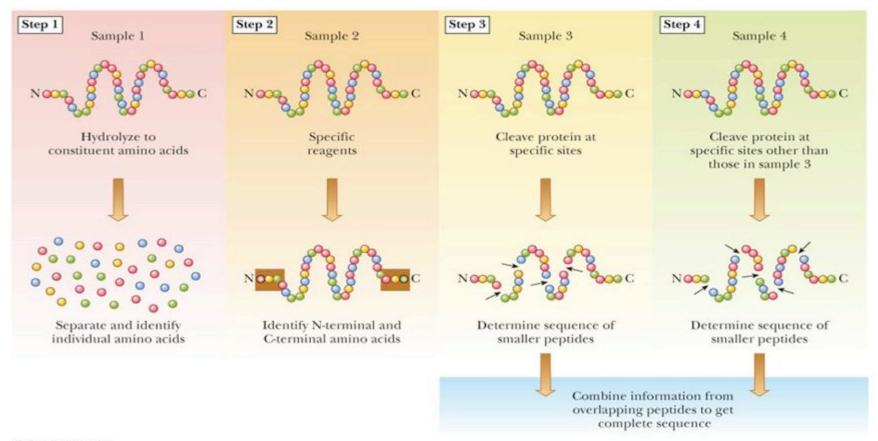
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# Analysis of proteins' primary structure:

### Primary sequnce analysis

Protein, sequence to be determined





## **Chemical degradation**

HCI: Complete degradation, 6M HCI at 100-110 °C for 12-36 h

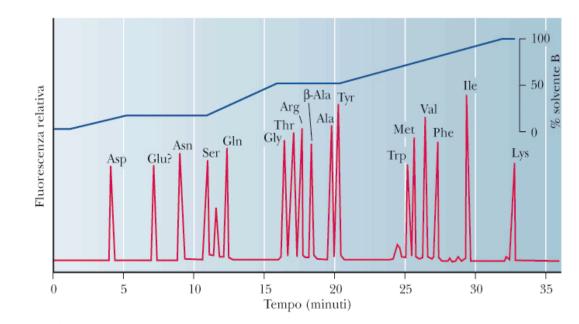


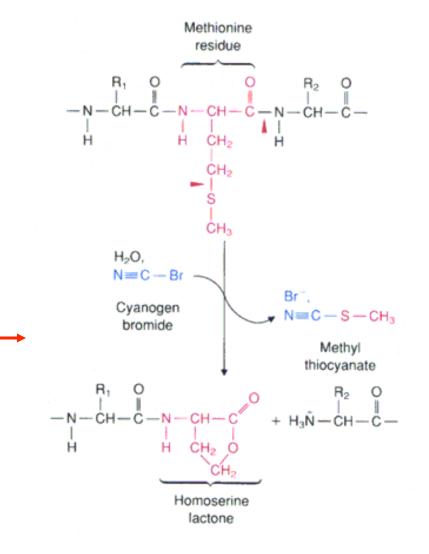
FIGURA 5.15 Cromatogramma HPLC relativo alla separazione di una miscela di amminoacidi.

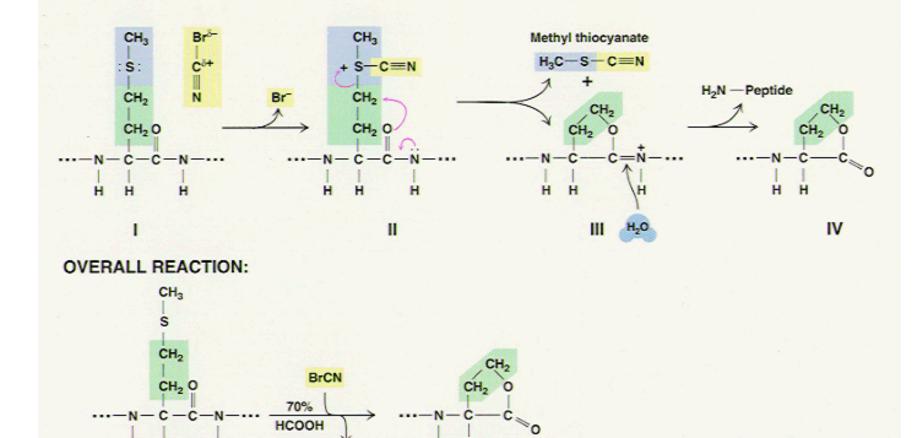


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### **Chemical degradation**

#### Cyanogen bromide





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Peptide with C-terminal

homoserine lactone

н

Overhead transparencies to accompany Garrett/Grisham: Biochemistry Transparency 14 Figure 4.25

н

н

Polypeptide

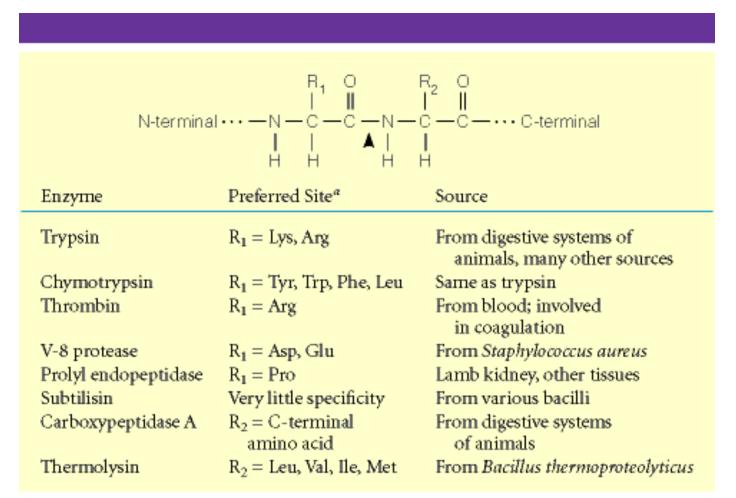
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Protein hydrolysis by cyanogen bromide takes place where methionine residues are

H<sub>2</sub>N - Peptide

#### Enzymatic degradation: Proteases:



"The residues indicated are those next to which cleavage is most likely. Note that in some cases preference is determined by the residue on the N-terminal side of the cleaved bond ( $R_1$ ) and sometimes by the residue to the C-terminal side ( $R_2$ ). Generally, proteases do not cleave where proline is on the other side of the bond. Even prolyl endopeptidase will not cleave if  $R_2$  = Pro.



A

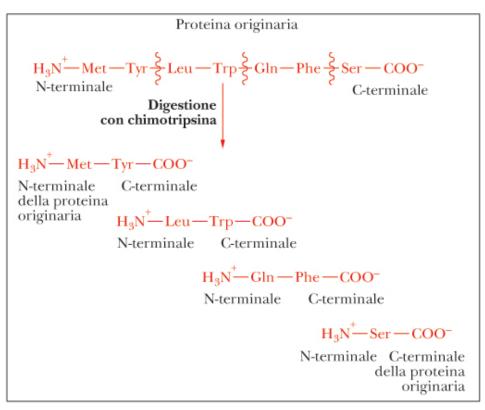
HN  $CH_2$  $CH_2$  $CH_2$ OH CH<sub>2</sub> CH<sub>2</sub> COO CH<sub>2</sub> O  $CH_2$ CH<sub>2</sub> O CH<sub>3</sub> O CH<sub>2</sub> O 0 CH - C - NCH – C CH CH - C -NCH Ala Arg Ser Lys Asp Ĥ Ĥ Η Н Η Tripsina Tripsina B N-Asp-Ala-Gly-Arg-His-Cys-Lys-Trp-Lys-Ser-Glu-Asn-Leu-Ile-Arg-Thr-Tyr-C Tripsina Asp-Ala-Gly-Arg His-Cys-Lys Trp-Lys Ser-Glu-Asn-Leu-Ile-Arg Thr-Tyr

 $NH_3^+$ 

 $NH_2$ 

 $C = NH_{2}$ 

**Peptide digestion with trypsin.** A) Trypsin is a proteolytic enzyme, or protease, that cuts specifically only peptide bonds where arginine or lysine provide the carbonyl group. B) The reaction products are a mixture of peptide fragments with Arg and Lys as C-term amino acids and a single peptide deriving from the C-terminal of the polypeptide chain.





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Chimotrypsin digestion of a protein. Chimoptrypsin hydrolyses proteins where aromatic amino acids are

1.0

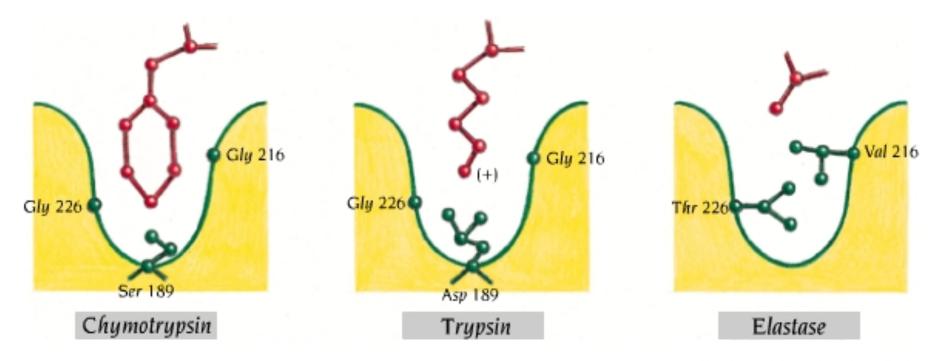
.

.

## The overlap of the sequences of fragments allows to determine the protein sequence

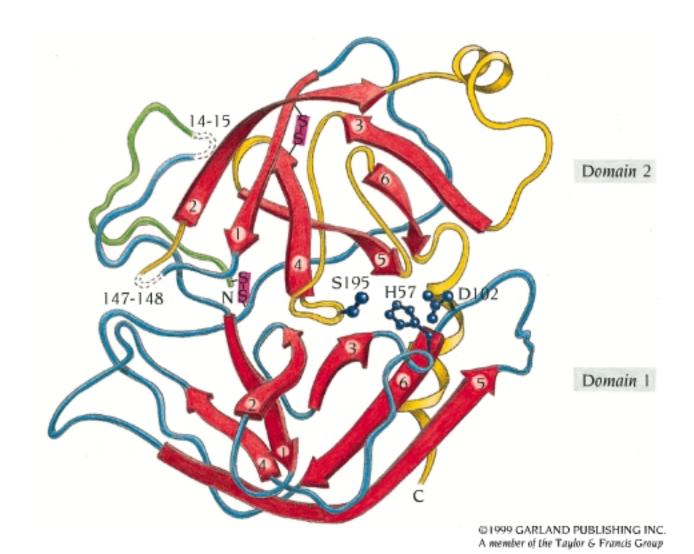
Chimotripsina	$H_3 \dot{N}$ — Leu — Asn — Asp — Phe
Bromuro di cianogeno	$H_3 N^+$ —Leu—Asn—Asp—Phe—His—Met
Chimotripsina	His—Met—Thr—Met—Ala—Trp
Bromuro di cianogeno	Thr — Met
Bromuro di cianogeno	Ala—Trp—Val—Lys—COO <sup>-</sup>
Chimotripsina	Val—Lys—COO <sup>-</sup>
Sequenza complessiva	$H_3$ <sup>+</sup> - Leu - Asn - Asp - Phe - His - Met - Thr - Met - Ala - Trp - Val - Lys - COO <sup>-</sup>

#### **Protease active sites**

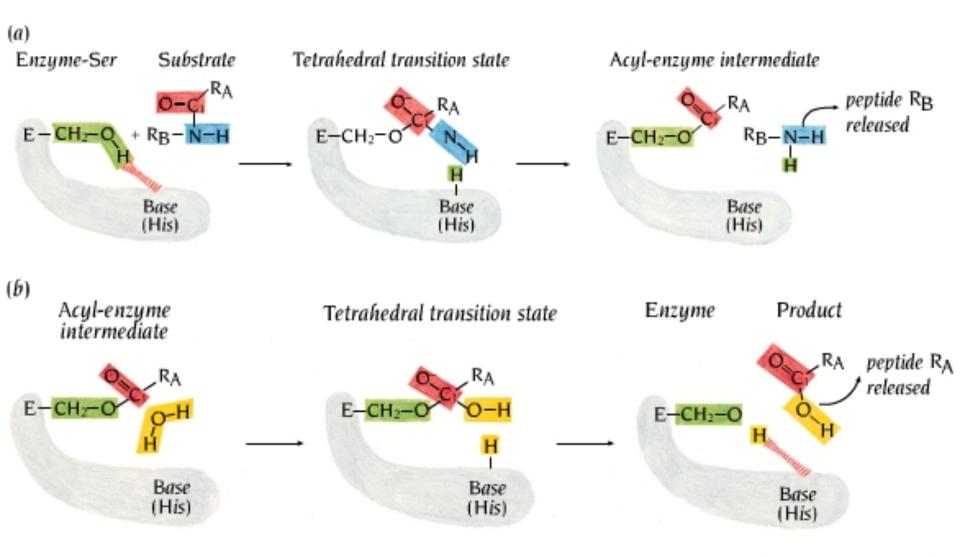


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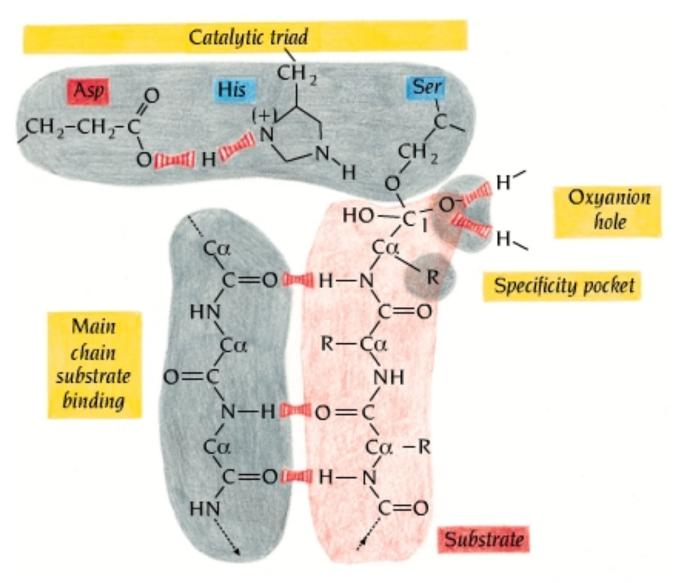
## Mechanism of protease activity



## Mechanism of protease activity

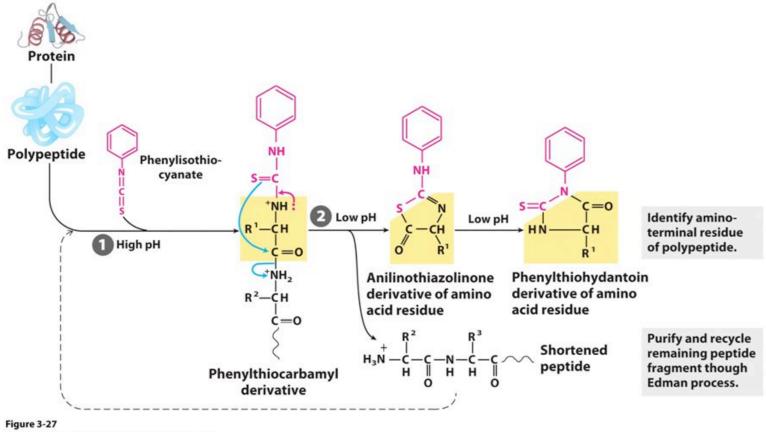


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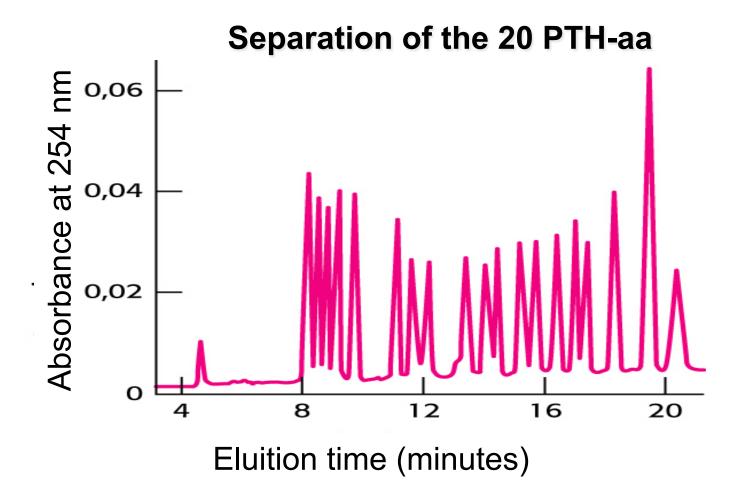
#### Edman degradation



Lehninger Principles of Biochemistry, Sixth Edition © 2013 W. H. Freeman and Company

**Edman degradation.** 1) In moderate alkaline conditions, phenylthioisocianate combines with the N-terminal of the peptide to form a phenyl thiocarbamoyl (PTC) derivative. 2) After treatment with trifluoracetic acid (TFA), a cyclic compound is formed and the first N-term amino acid is released as a thiazoline derivative, whereas the other peptide binds are not hydrolized. 3) After organic extraction and treatment with with an aqueous and acidic solution, a phenylthiohydantoin derivative (PTH) of the N-term amino acid is formed. The process is repeated several times to determine the amino acid at each step until the sequence of the peptide is complete.

#### Identification of theN-termPTH-aa N-terminale through chromatography



### Peptide analysers

- The two major direct methods of protein sequencing are the <u>Edman degradation</u> reaction and <u>mass spectrometry</u>.
- Proteomics

