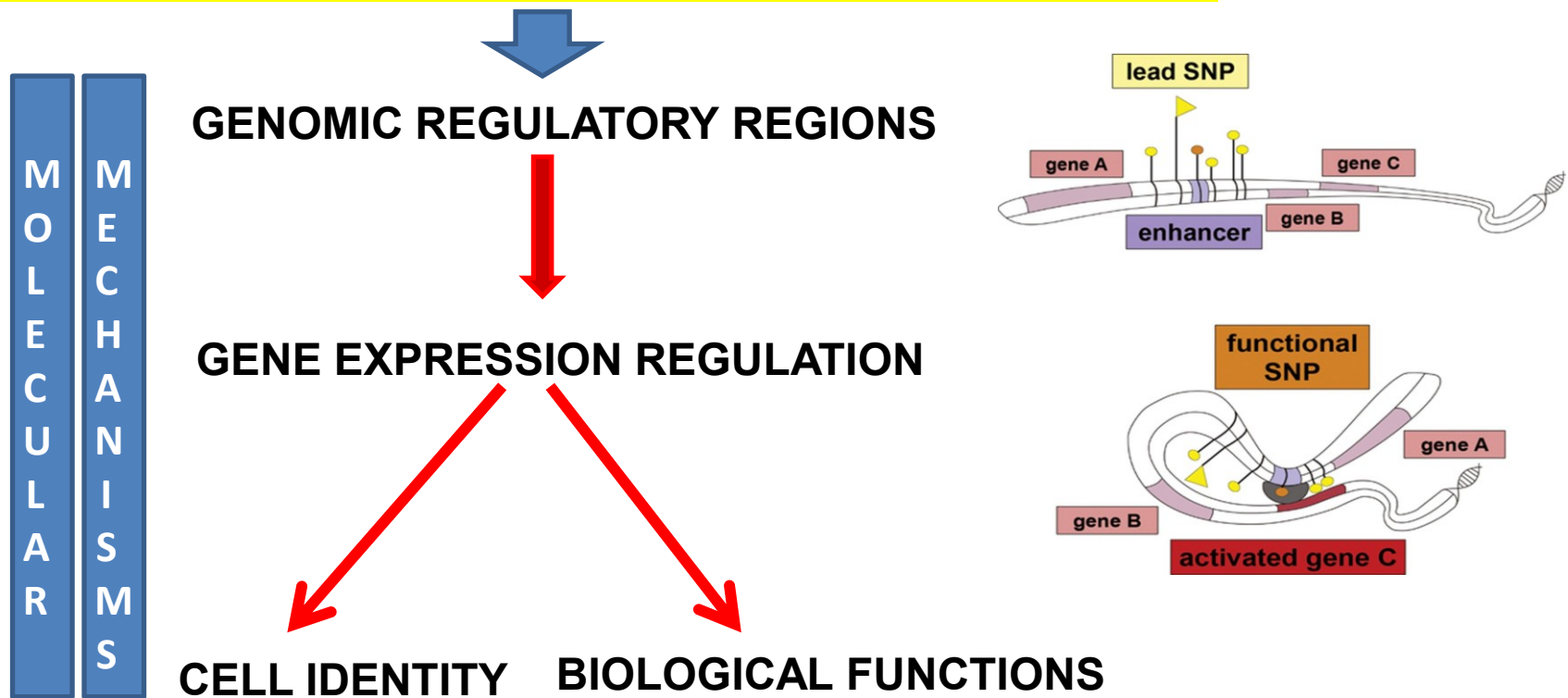


# In Lesson 1

- **Functional genomics is a field of molecular biology based on genome-wide sequencing data.**
- **Genome-wide sequencing data describe genomic regulatory regions that control gene expression**
- **Gene expression dysregulation may be linked to the disease**
- **Understanding molecular mechanisms of disease outcome opens the way to discovery drug and identify biomarkers**

Genomic regulatory regions are **cell regulatory pattern** that defines cell identity and biological functions. Variants in the genomic regulatory regions may change the molecular mechanisms to control gene expression and may be linked to disease occurrence and development.

## IDENTIFICATION AND CHARACTERIZATION



**TO UNDERSTAND DISEASES**

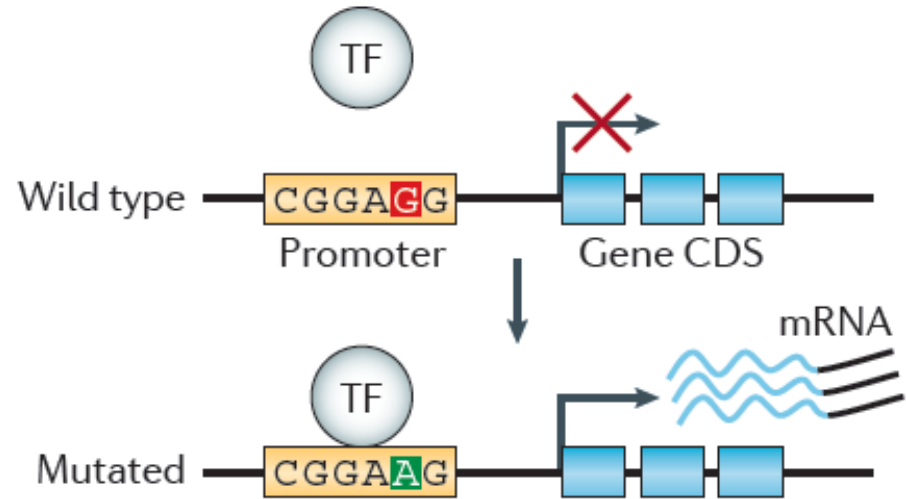
# How SNPs play a FUNCTIONAL role in disease:

## Impact on transcription

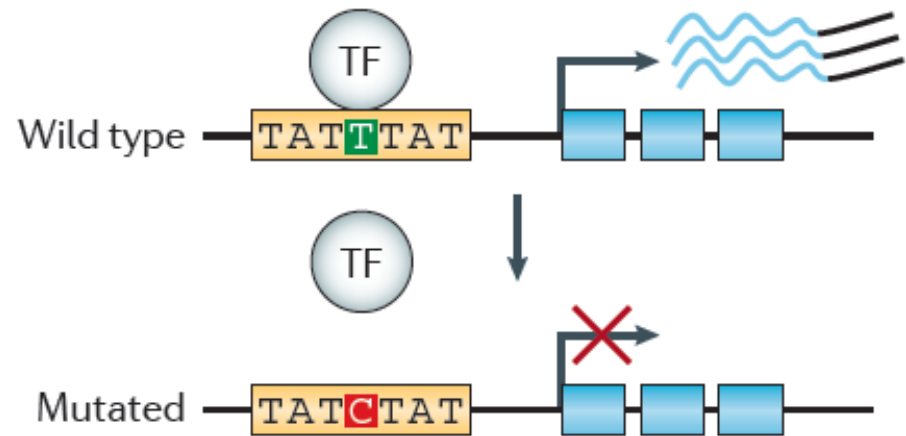
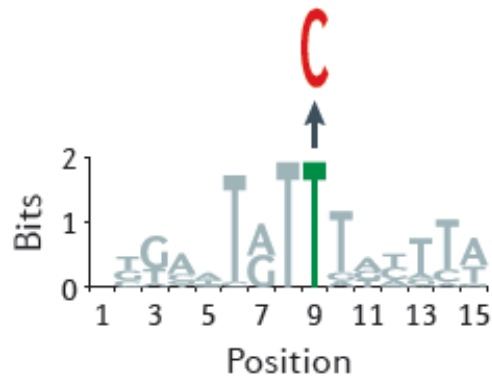
- Changing consensus sequences for transcription factors binding sites
- Changing interaction between for transcription factors
- Changing epigenetic profiling of specific genomic regions
- Changing long range interaction between two genomic regions

# SNPs types functions:

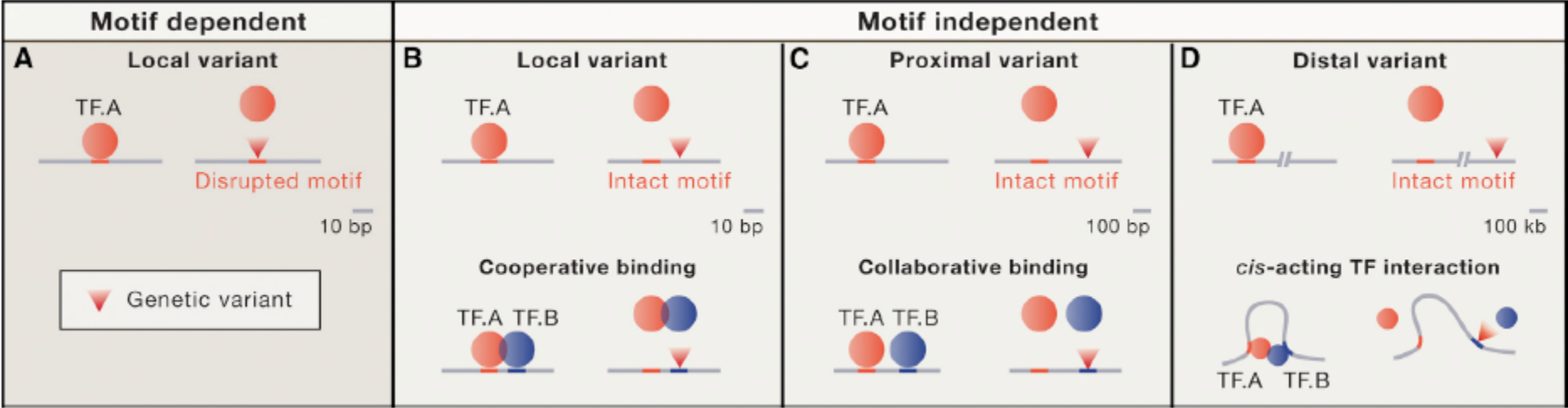
## Ba Gain of motif



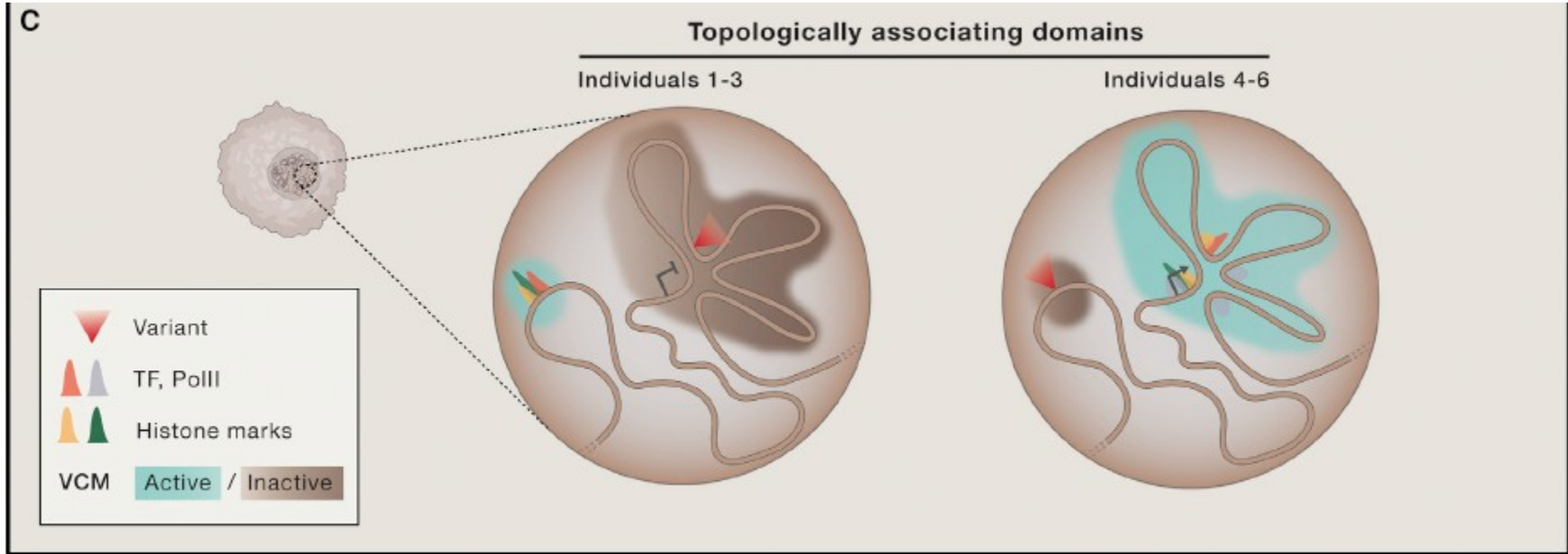
## Bb Loss of motif



# SNPs mechanisms for alteration of regulatory transcription factors complexes



# SNPs may change long range interactions



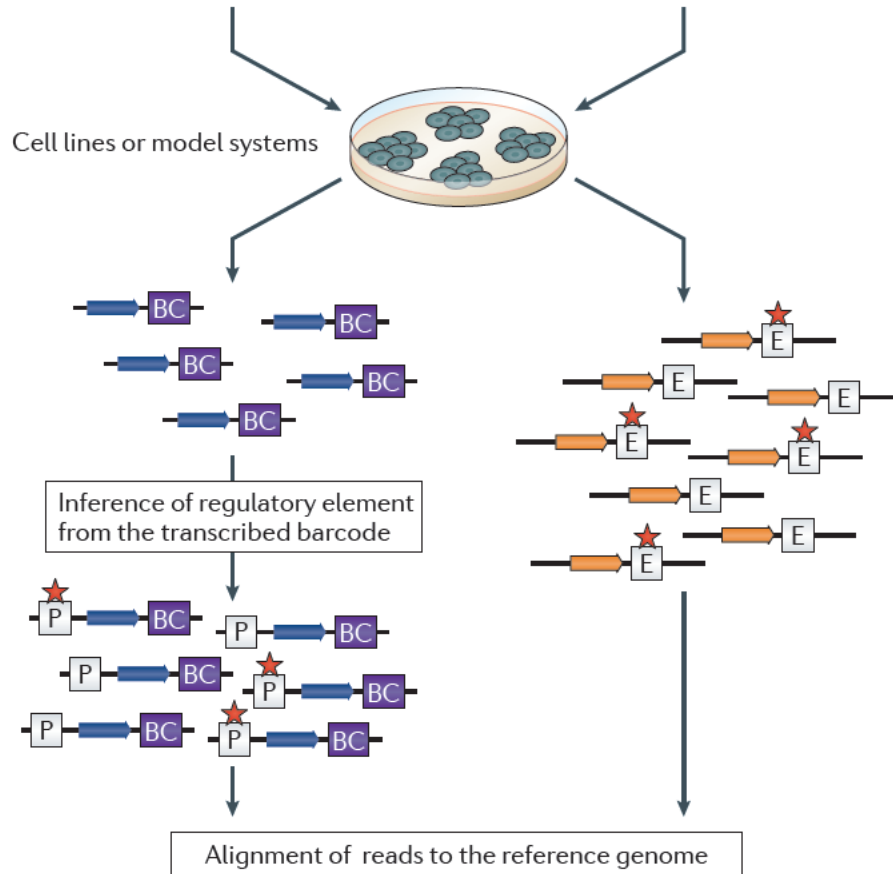
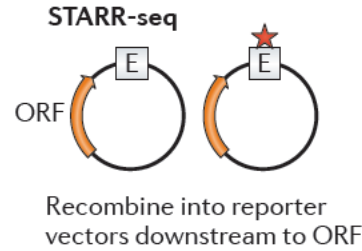
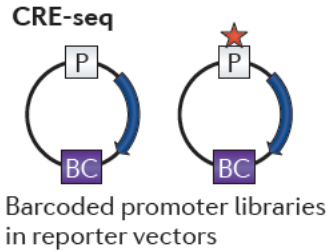
**How SNPs play a FUNCTIONAL role in disease:**

**Alteration of cell identity  
and  
biological functions**

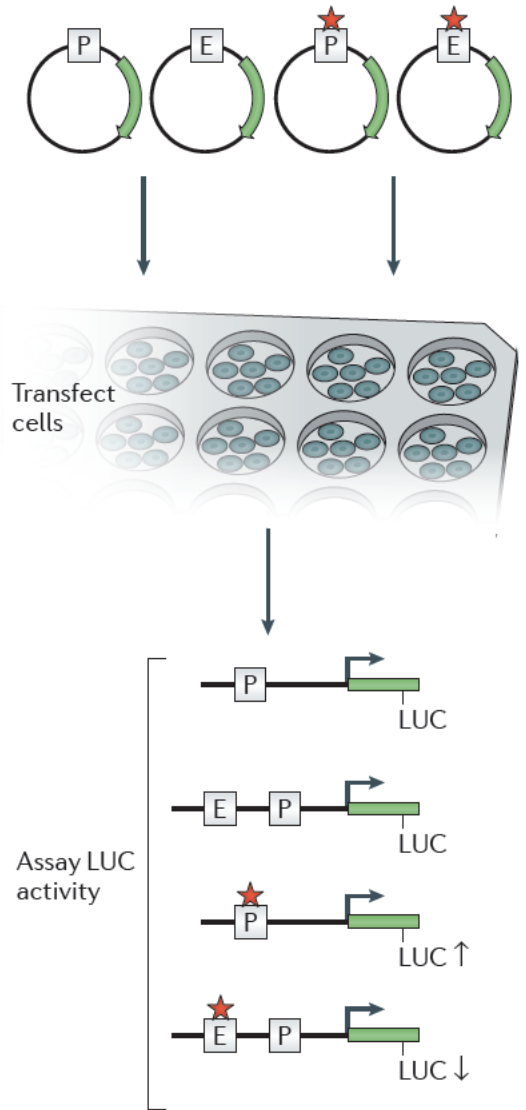
# MOLECULAR FUNCTIONAL EFFECTS

## b Test molecular functional effects on target gene

### Combined analysis and validation using high-throughput sequencing



### LUC reporter activity

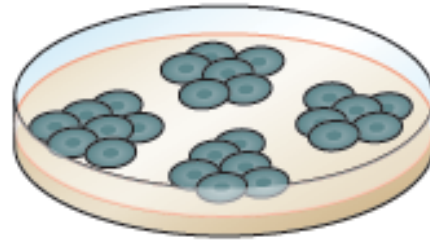




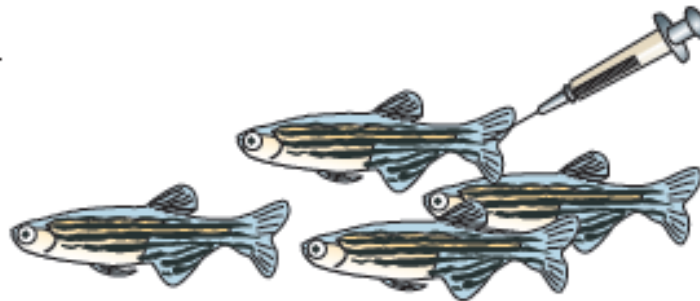
# BIOLOGICAL FUNCTION TESTS

## c Test effects on oncogenesis

- Proliferation
- Invasion
- Migration



Cell lines



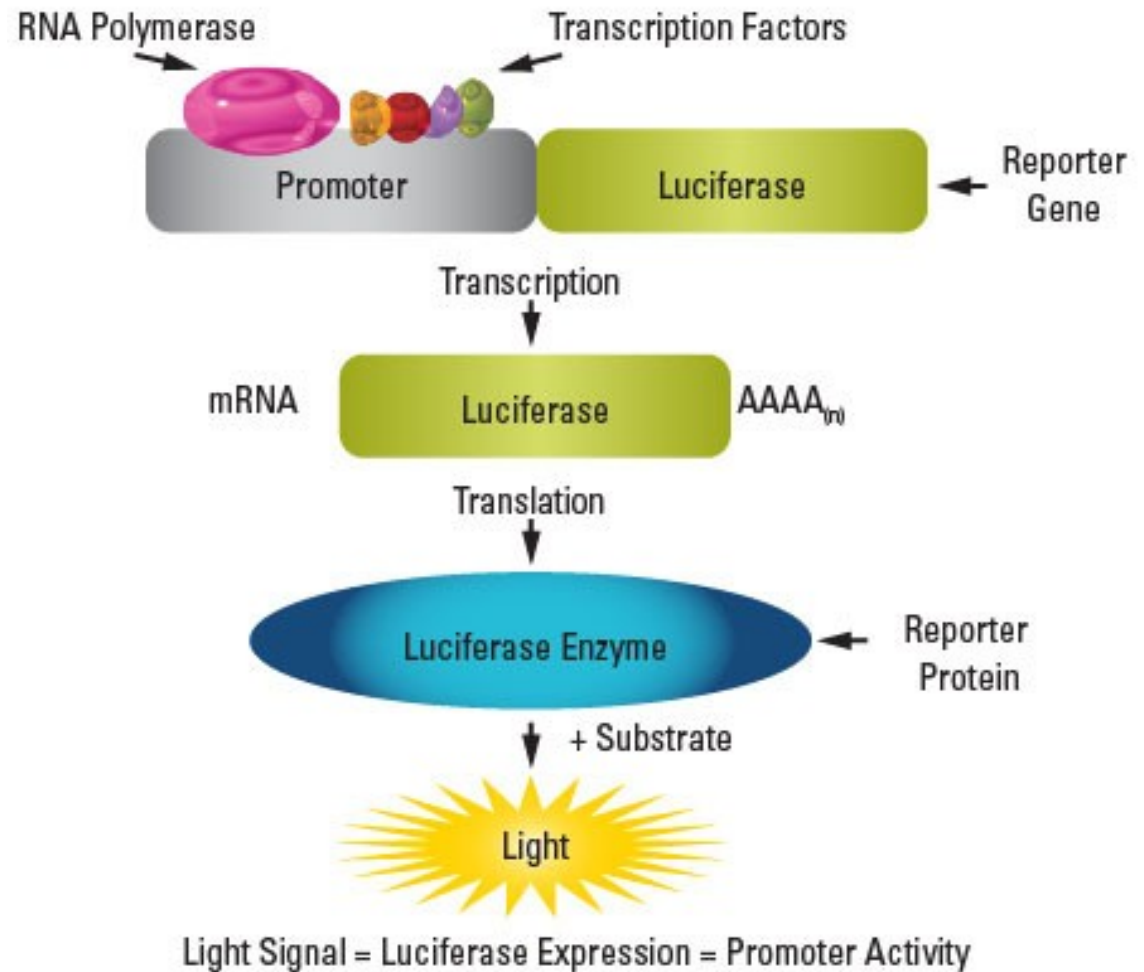
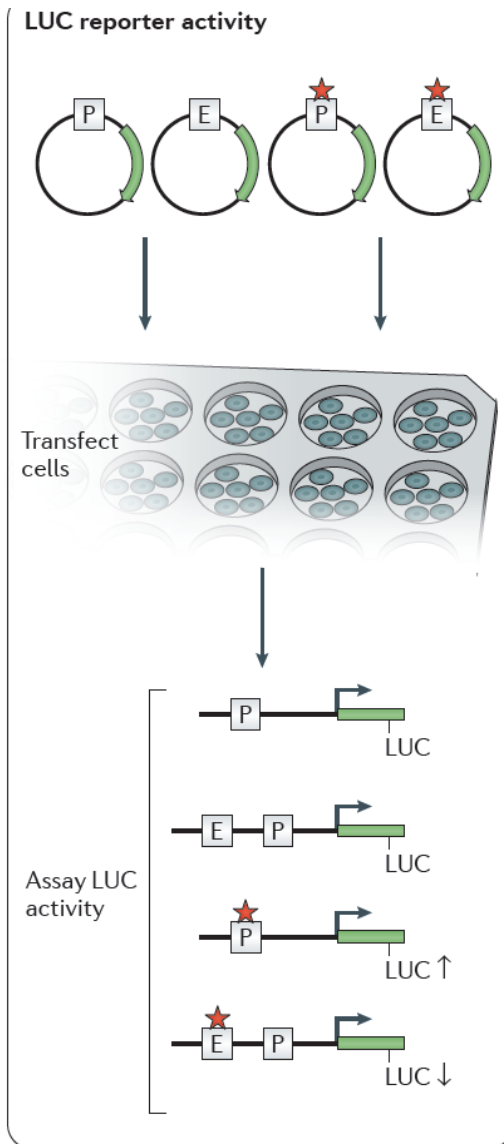
Zebrafish

Tumorigenesis

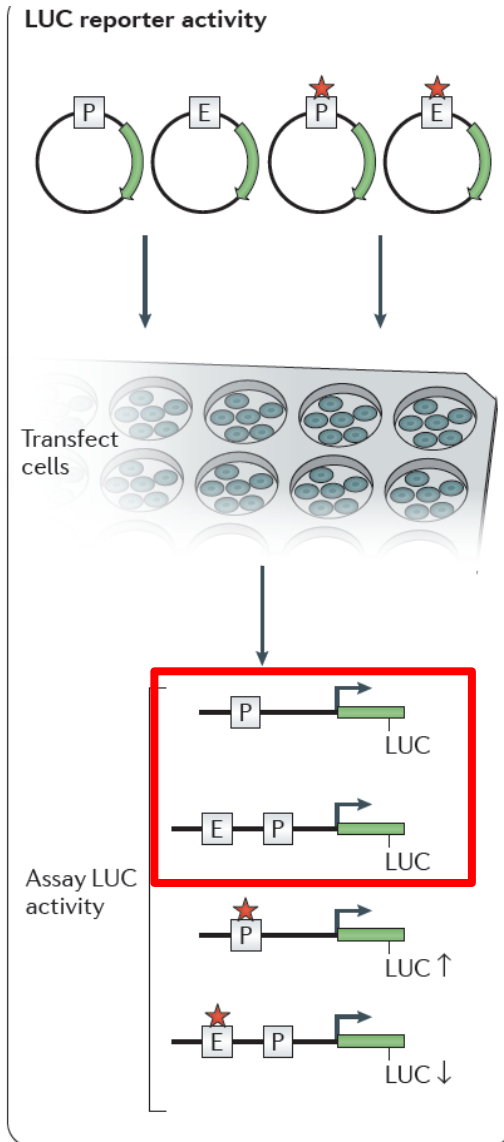


Mouse

In order to test if SNP has a role in the transcription rate by alteration of TFBS, luciferase assay (REPORTER GENE ASSAY) can be used



In order to test if SNP has a role in the transcription rate by alteration of TFBS, luciferase assay can be used



To test enhancer and promoter with SNPs:

Question:

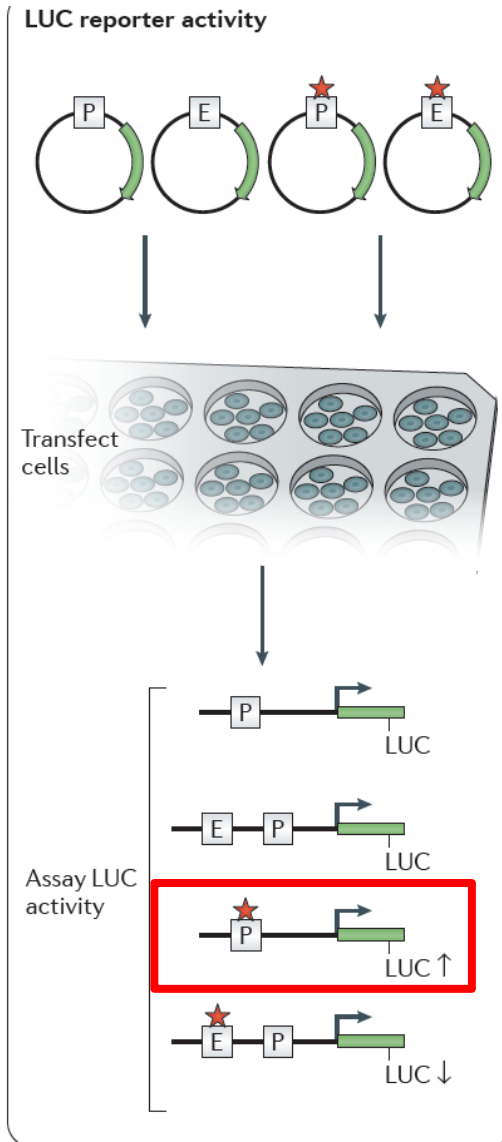
Is the **SNP in the promoter** or **in enhancer** able to change transcription activation?

How

Does Transcription Increase or Decrease?

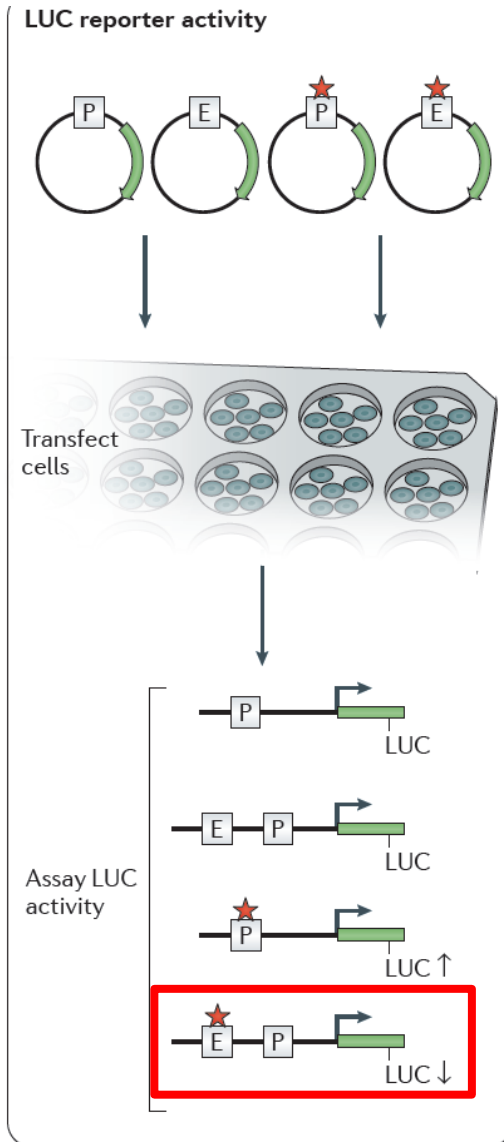
Promoter and Enhancer wild type do not induce transcription

In order to test if SNP has a role in the transcription rate by alteration of TFBS, luciferase assay can be used



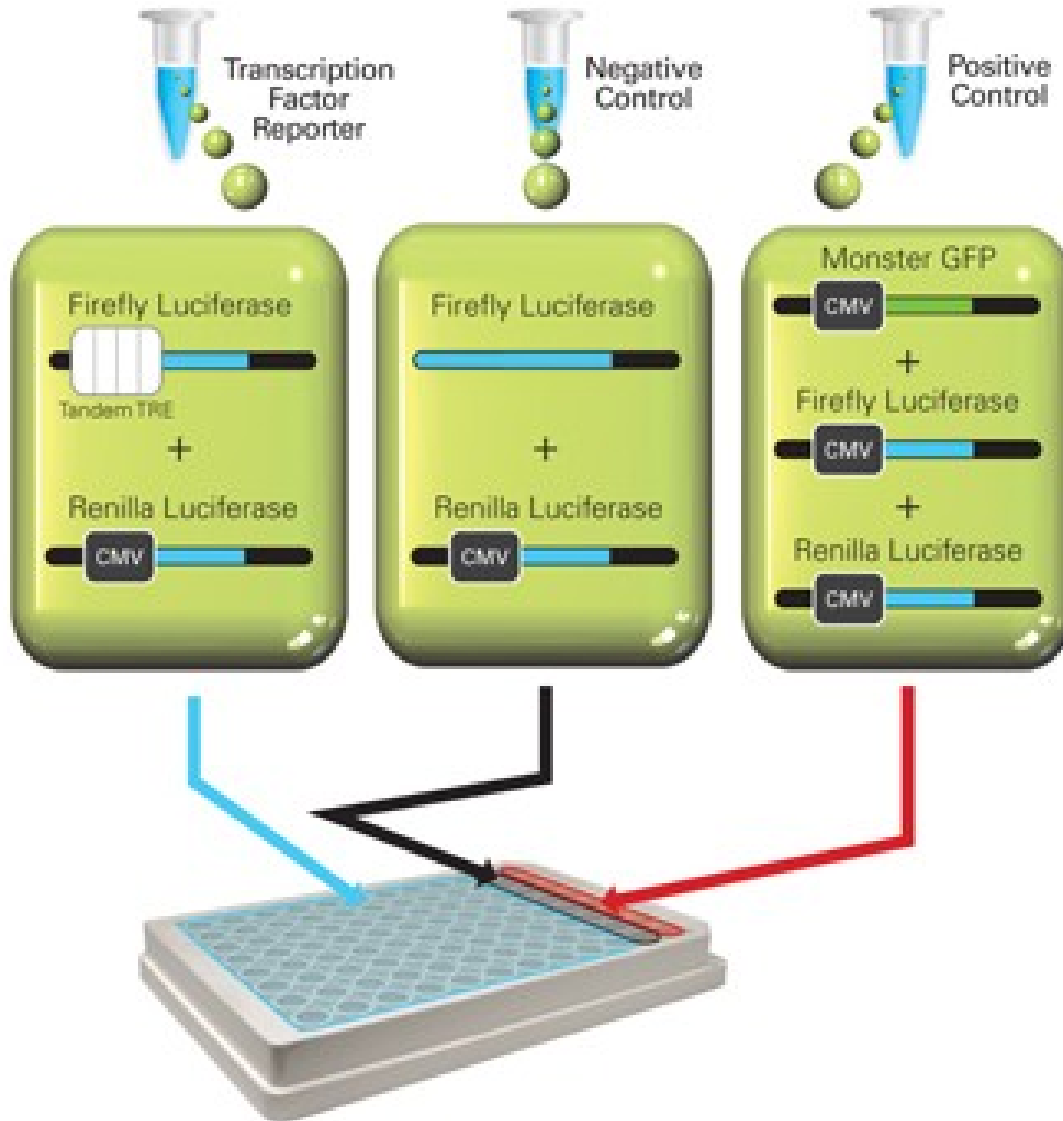
SNP in the Promoter increases transcription

In order to test if SNP has a role in the transcription rate by alteration of TFBS, luciferase assay can be used

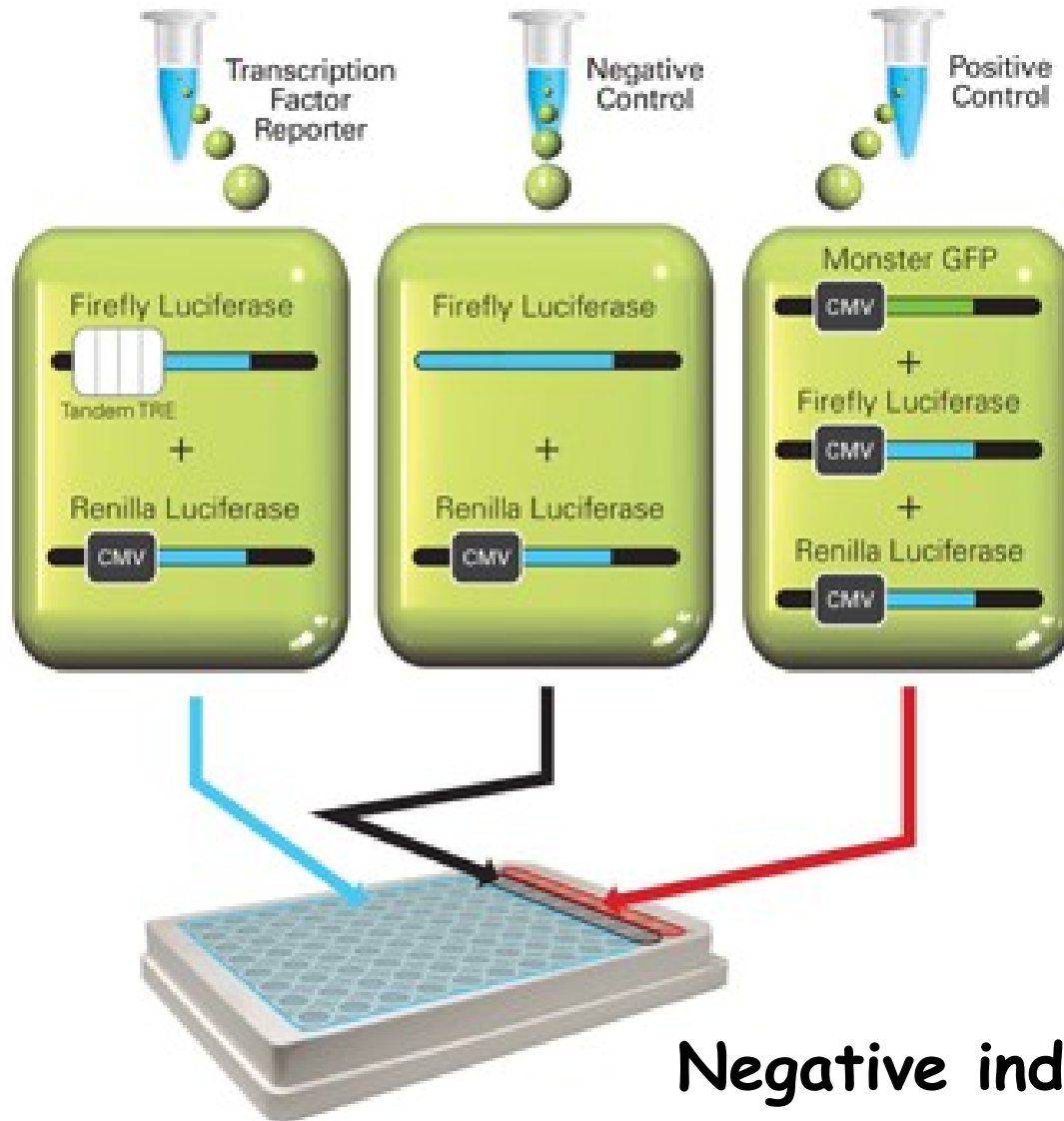


SNP in the Enhancer decrease the transcription

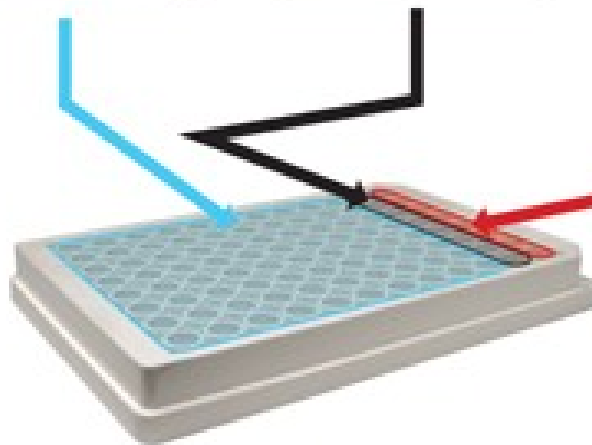
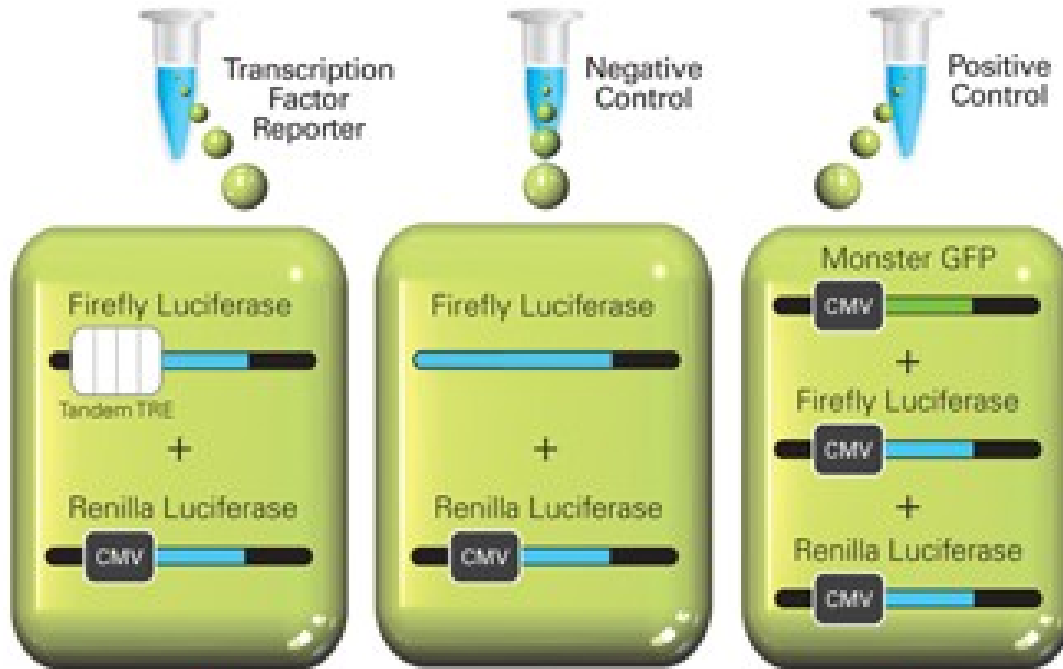
# What are positive and negative controls?



**Negative does not have a “regulatory sequence” that you want to test**



Positive control has a “constitutive active sequence” that induce transcription



Positive control indicates that the assay works well: reagents in the kit are good and are not degraded



# In this Lesson

- **Enhancer Overview**
- **Genomic regulatory network to define cell identity**
- **Genetic variations meaning in cell identity**

# GENOMIC REGULATORY REGIONS ARE PROMOTER, in proximity of gene target, and ENHANCER, distant from gene target

## THE TOPIC IN BRIEF

- Epigenomics is the study of the key functional elements that regulate gene expression in a cell.
- Epigenomes provide information about the patterns in which structures such as methyl groups tag DNA and histones (the proteins around which DNA is packaged to form chromatin), and about interactions between distant sections of chromatin.
- They also contain information about regulatory elements in DNA itself: both those that lie in the promoter region immediately upstream of where a gene's transcription begins, and those in distant enhancer sequences.

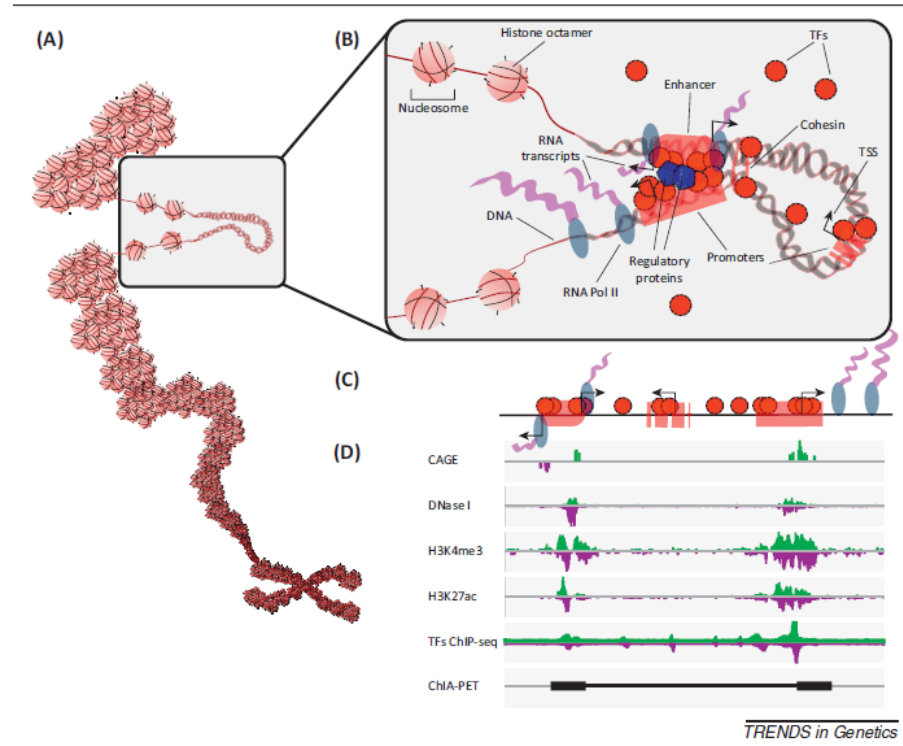
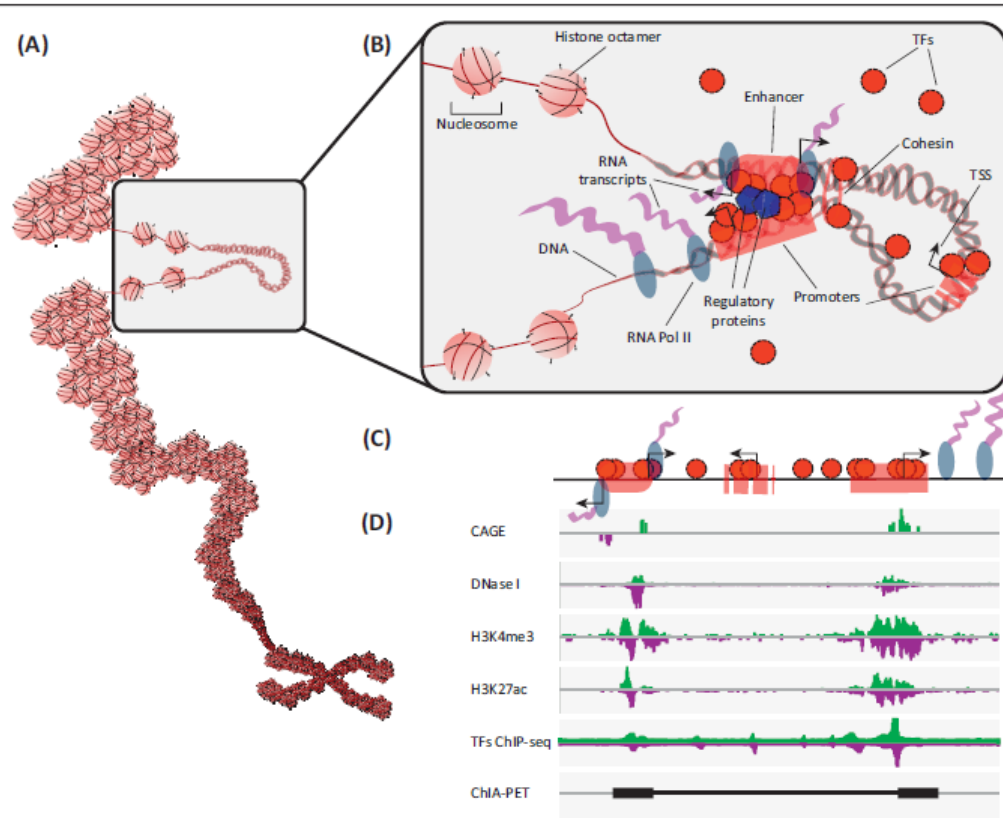


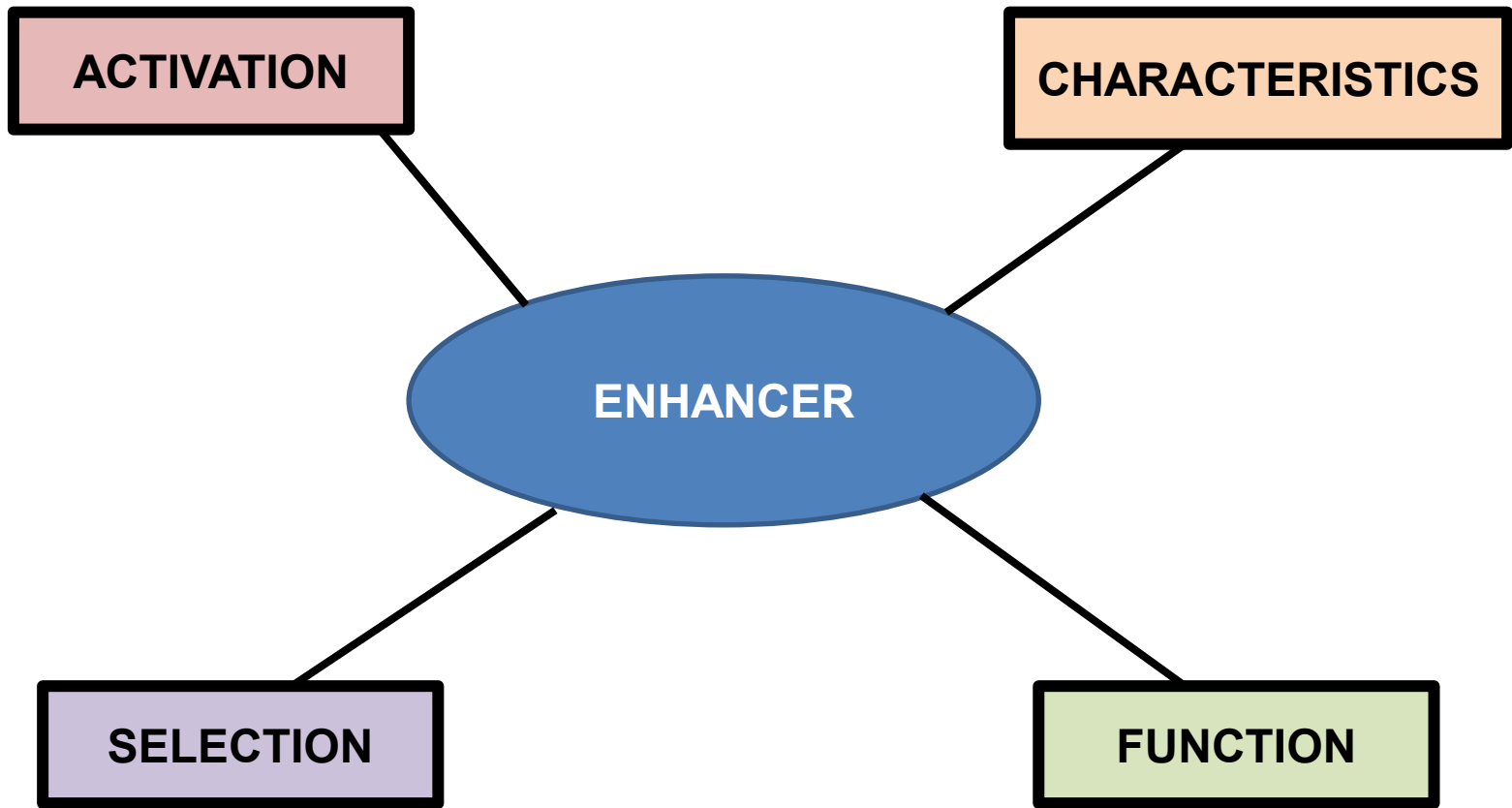
Figure B shows a region of the DNA which is transcribed and in particular it point out the importance of both trans-activating and cis-activating regulatory elements. In fact, we can see many transcripts and proteins (Regulatory Factors and Transcription factors), interacting with the DNA and also the formation of a loop with enable two distant regions to be found closer.



# The selection and function of cell type-specific enhancers

*Sven Heinz<sup>1</sup>, Casey E. Romanoski<sup>2</sup>, Christopher Benner<sup>1</sup> and Christopher K. Glass<sup>2,3</sup>*

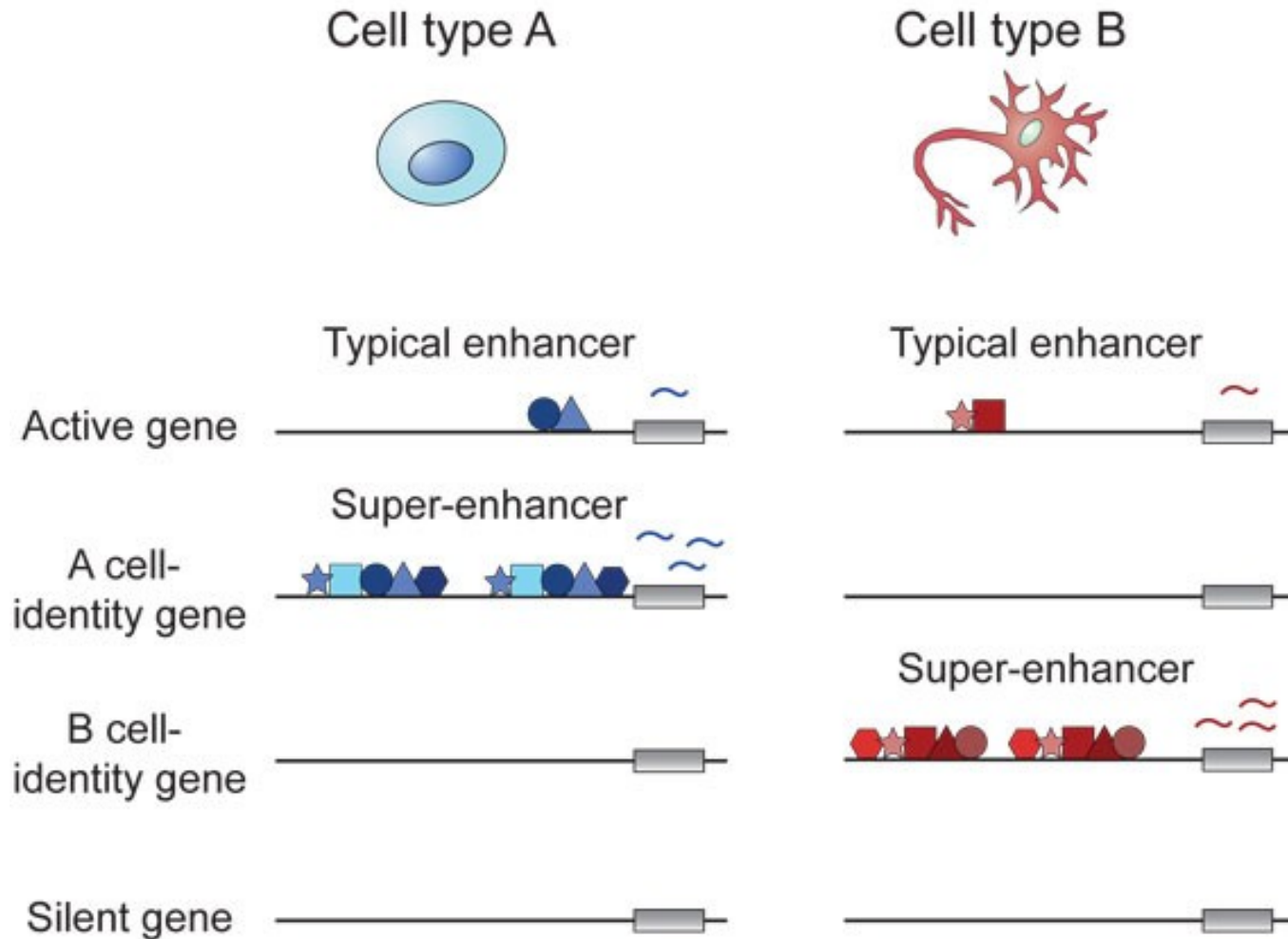
Abstract | The human body contains several hundred cell types, all of which share the same genome. In metazoans, much of the regulatory code that drives cell type-specific gene expression is located in distal elements called enhancers. Although mammalian genomes contain millions of potential enhancers, only a small subset of them is active in a given cell type. Cell type-specific enhancer selection involves the binding of lineage-determining transcription factors that prime enhancers. Signal-dependent transcription factors bind to primed enhancers, which enables these broadly expressed factors to regulate gene expression in a cell type-specific manner. The expression of genes that specify cell type identity and function is associated with densely spaced clusters of active enhancers known as super-enhancers. The functions of enhancers and super-enhancers are influenced by, and affect, higher-order genomic organization.



# Enhancer Characteristics

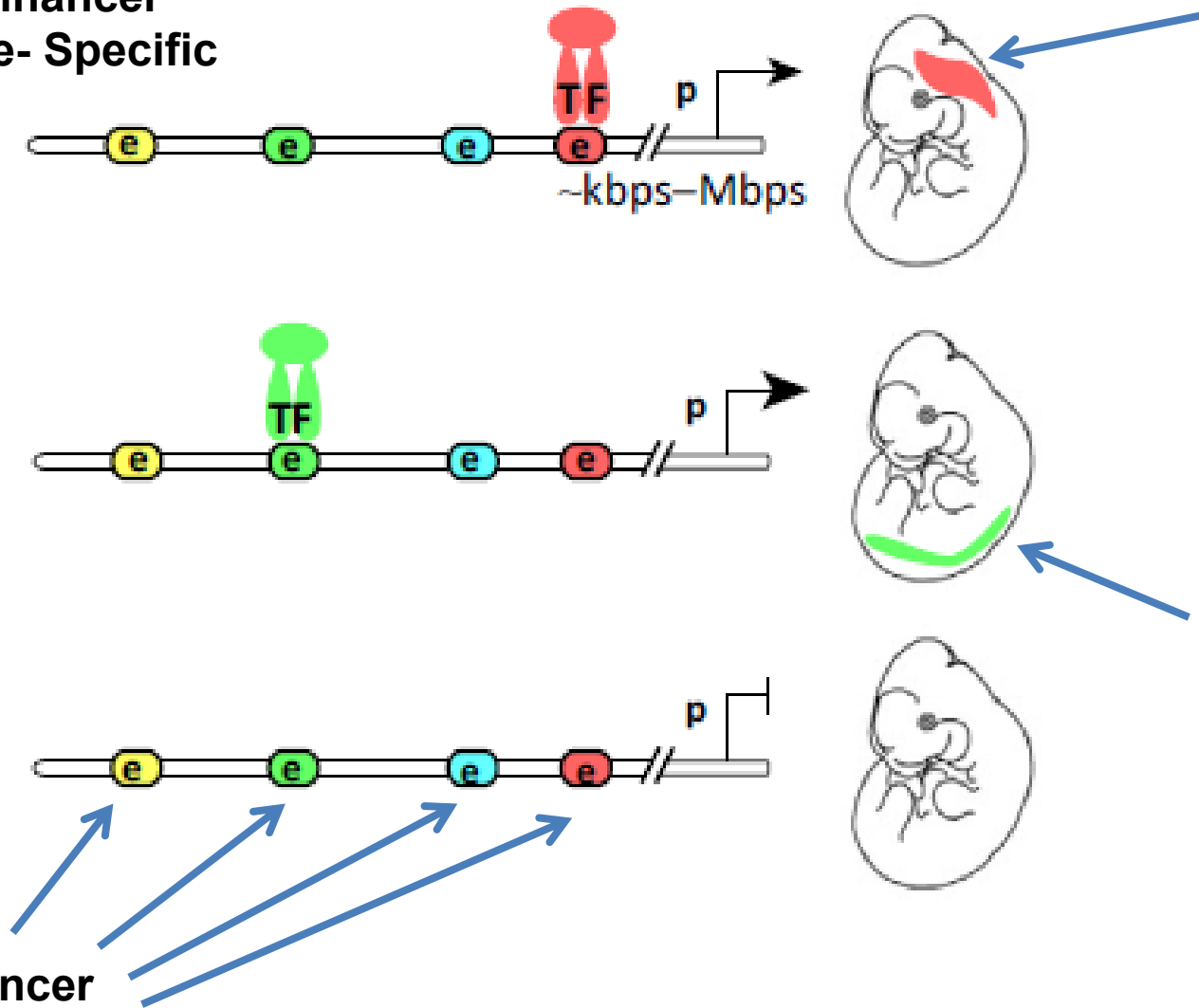
- **Enhancers are cis-regulatory elements in proximity of genes**
- **Each cell has a set of enhancers**
- **Enhancers have motifs for sequence-specific transcription factors**
- **Enhancers are marked with epigenetic modifications**
- **Enhancers are in different states of activation**

# MUCH OF THE **REGULATORY CODE** THAT DRIVES CELL-TYPE-SPECIFIC GENE EXPRESSION IS LOCATED IN DISTAL ELEMENTS CALLED **ENHANCERS**



# CELL TYPE USE A SMALL SUBSET OF MILLIONS OF POTENTIAL ENHANCERS

Active Enhancer  
in Cell-Type- Specific



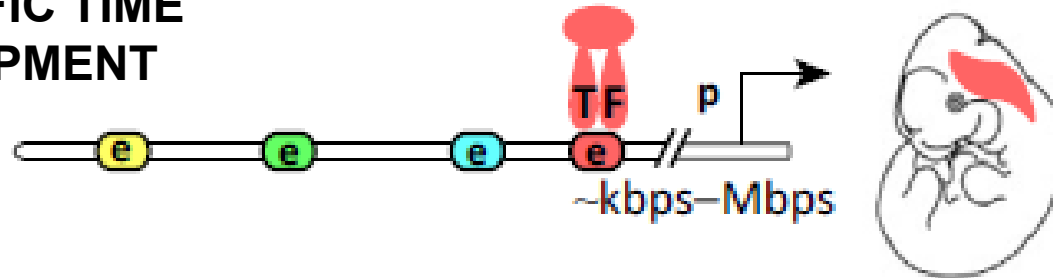
Potential enhancer

Enhancers in tissue/cell-specific gene expression

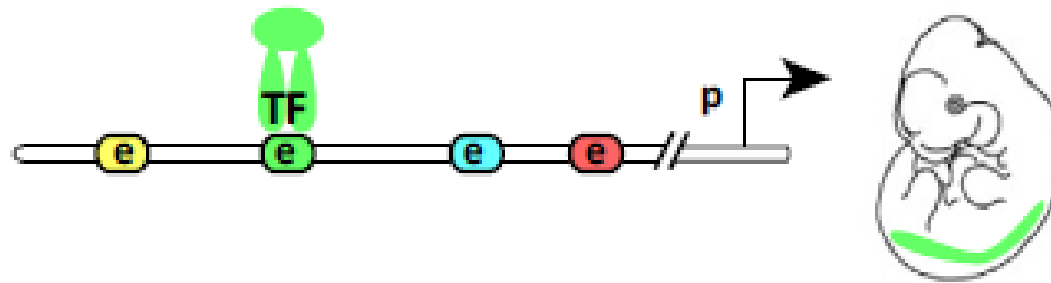


# CELL TYPE USE A SMALL SUBSET OF MILLIONS OF POTENTIAL ENHANCERS

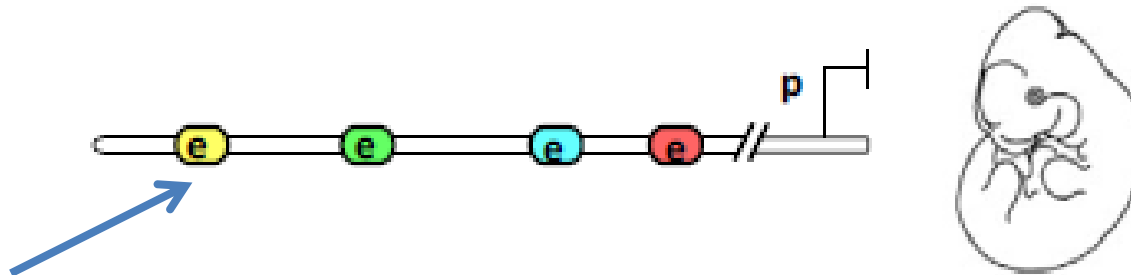
Active Enhancer  
during SPECIFIC TIME  
OF DEVELOPMENT



Stage 1



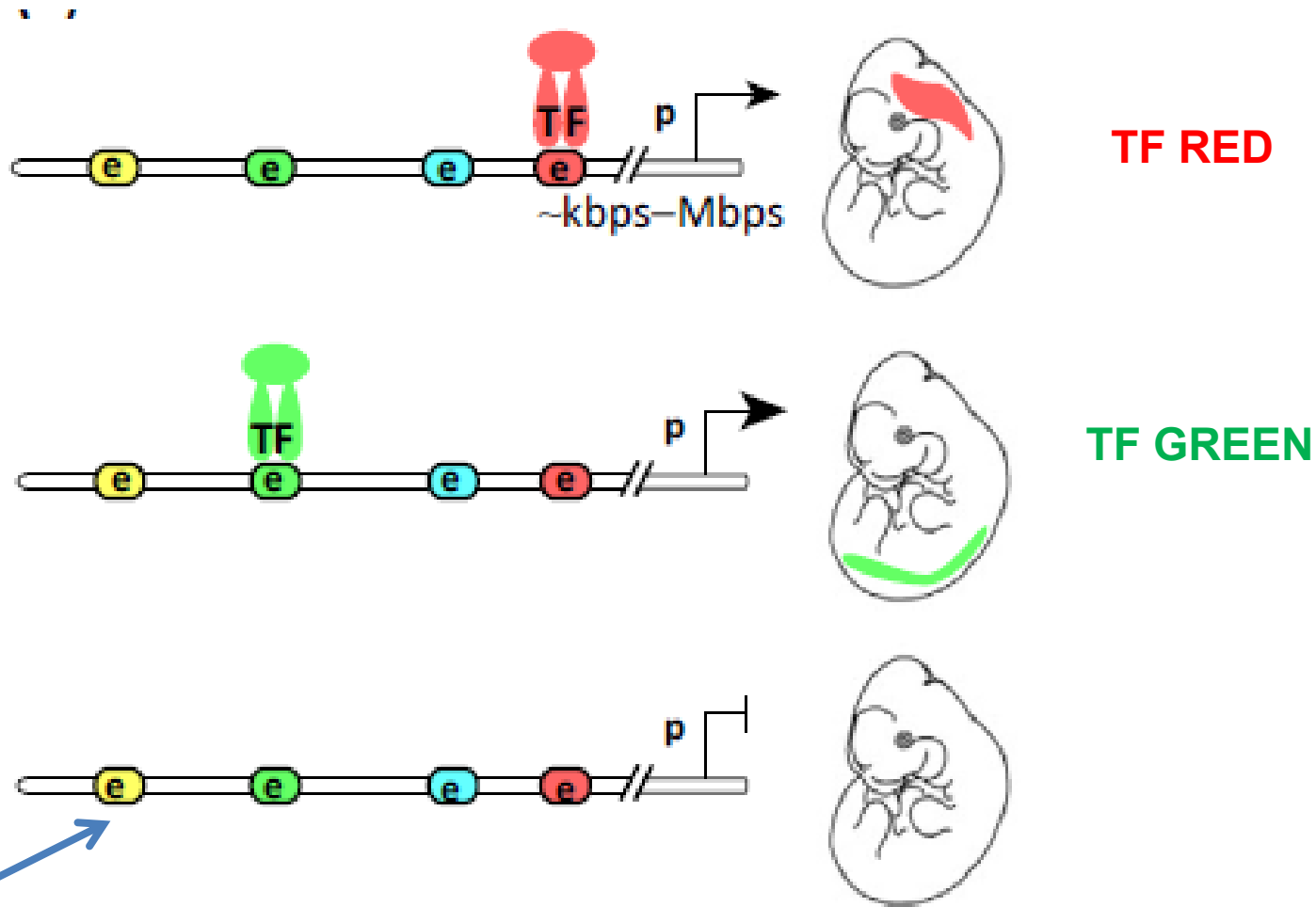
Stage 2



Potential enhancers

Enhancers in tissue/cell-specific gene expression

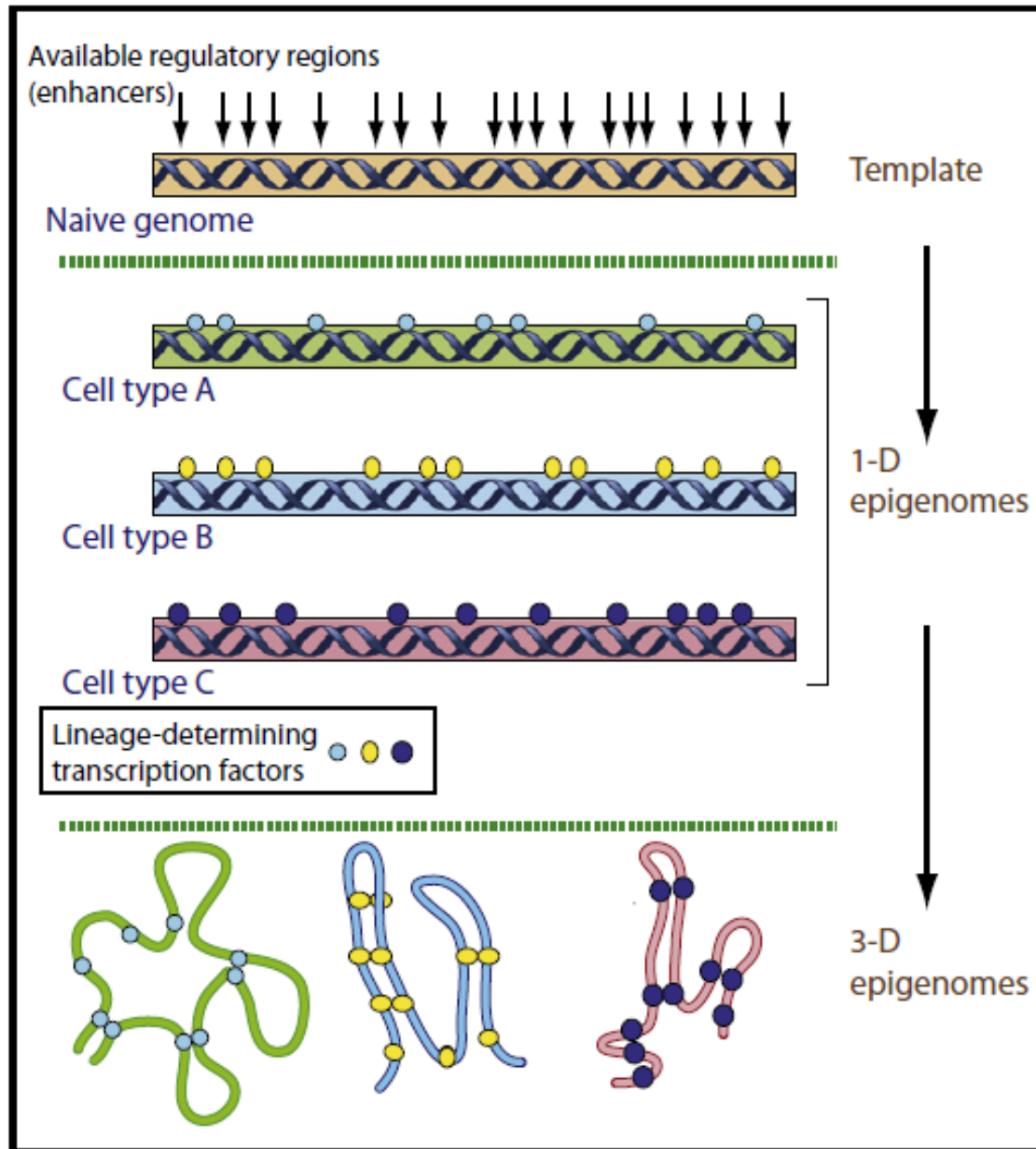
# LINEAGE-DETERMINING TRANSCRIPTION FACTORS BIND AT CELL-TYPE SPECIFIC ENHANCERS



Potential enhancers

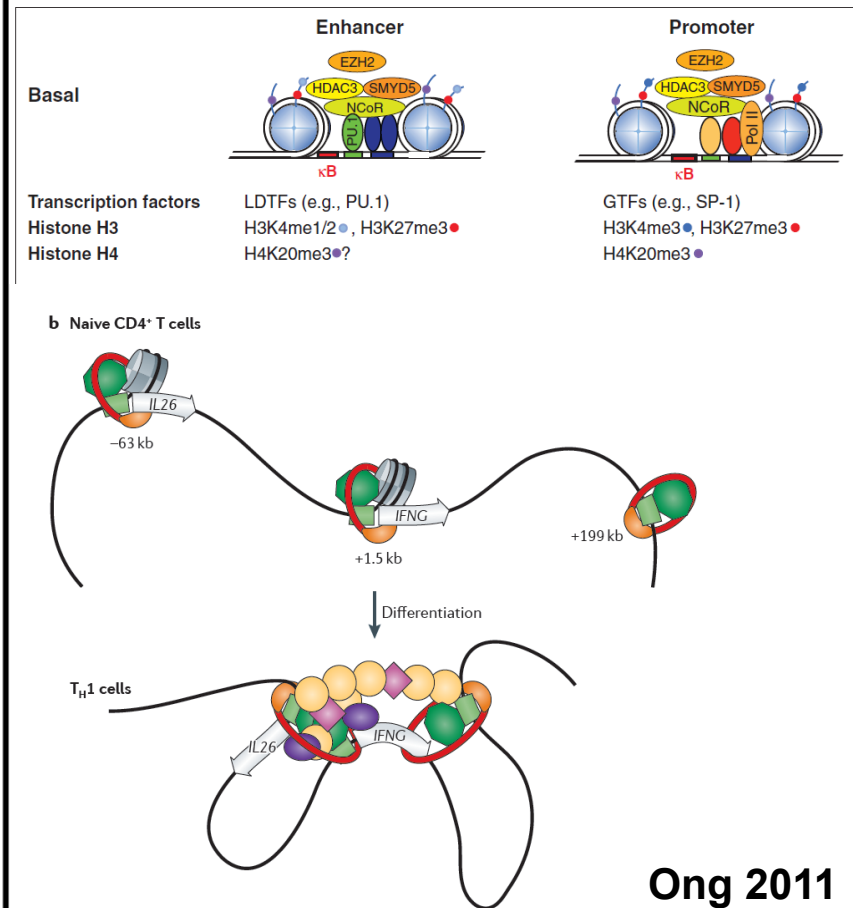
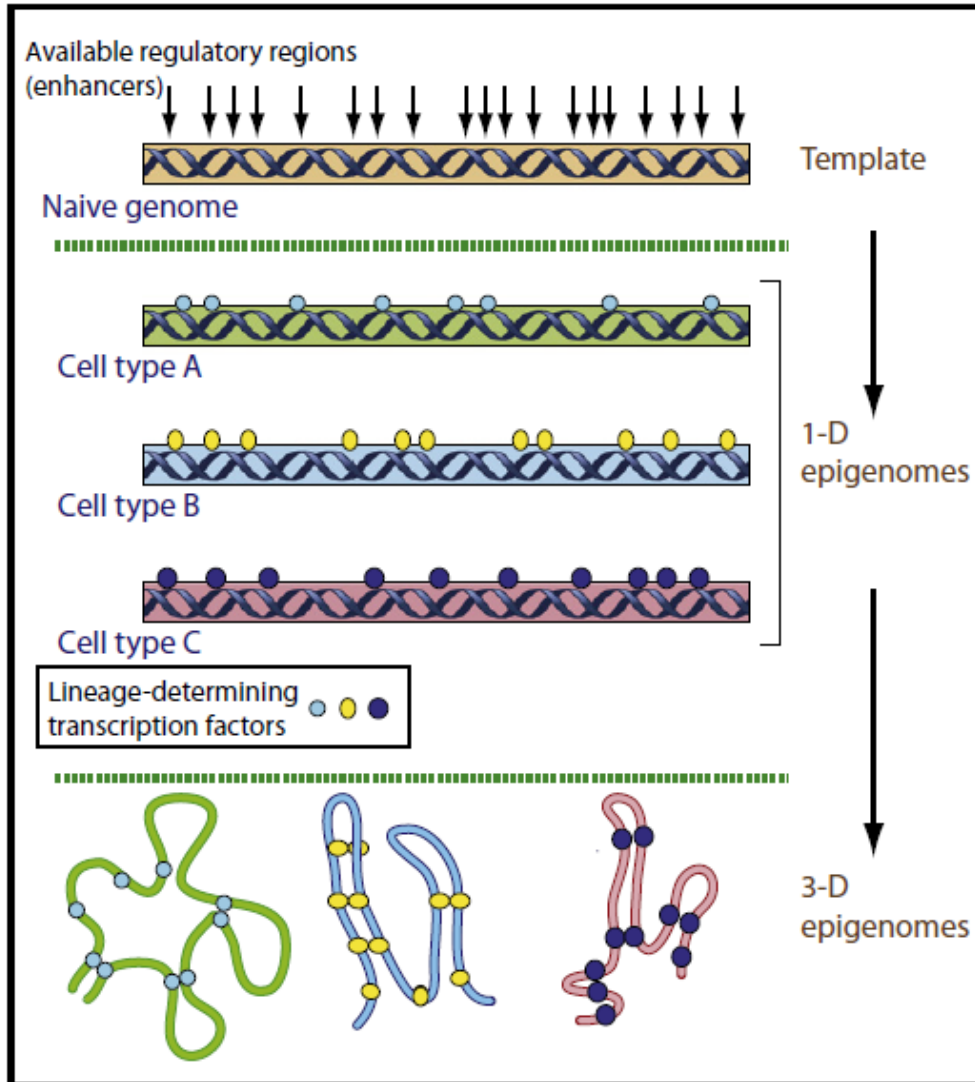
Enhancers in tissue/cell-specific gene expression

# EACH CELL HAS ACTIVE ENHANCERS



# Maintaining Cell Identity through Global Control of Genomic Organization

Gioacchino Natoli<sup>1,\*</sup>



Ong 2011

# TRANSCRIPTION FACTOR BINDS SPECIFIC CONSENSUS SEQUENCE IN ACTIVE ENHANCER

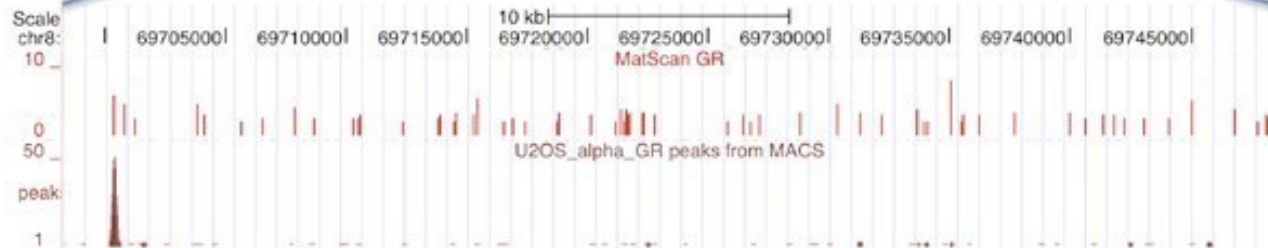


GR binding site motif found in approx. every 1000bp in genome



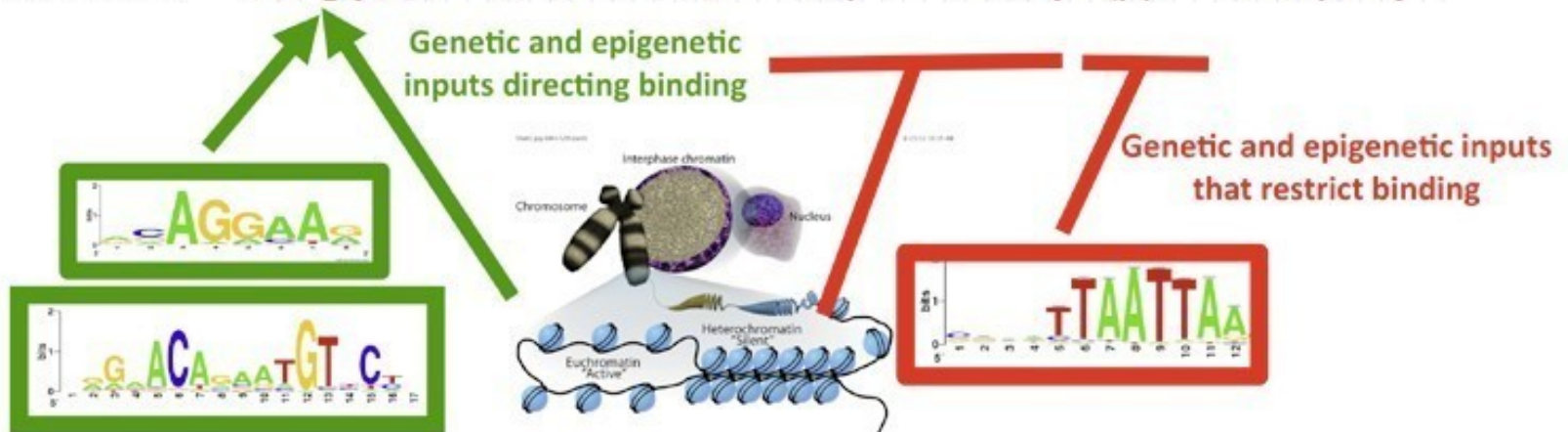
Red line marks  
GR binding motif:

Peak reflects actual  
GR Binding event:



Genetic and epigenetic  
inputs directing binding

Genetic and epigenetic inputs  
that restrict binding



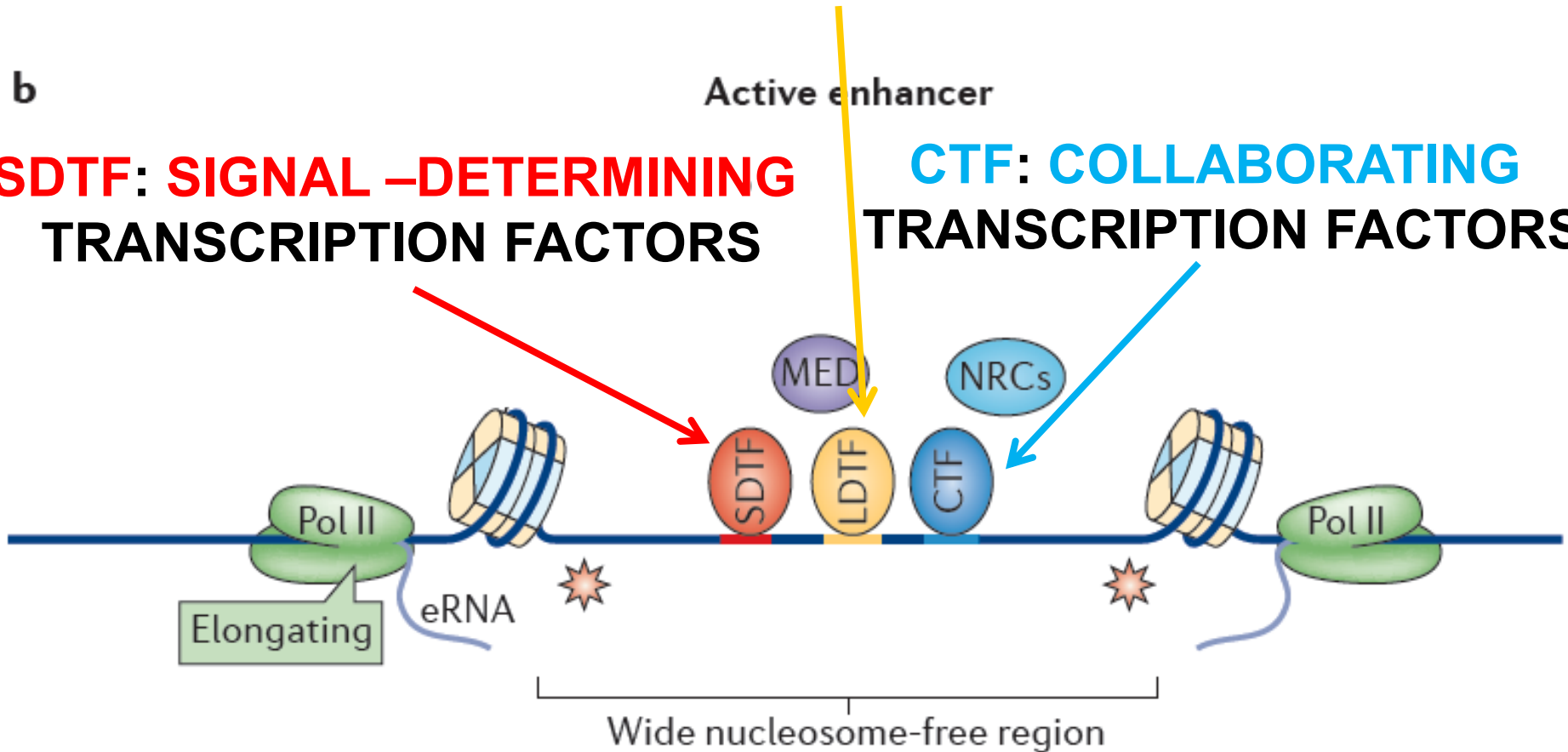
# TRANSCRIPTION FACTORS THAT BIND ENHANCERS

## LDTF: LINEAGE – DETERMINING TRANSCRIPTION FACTORS

b

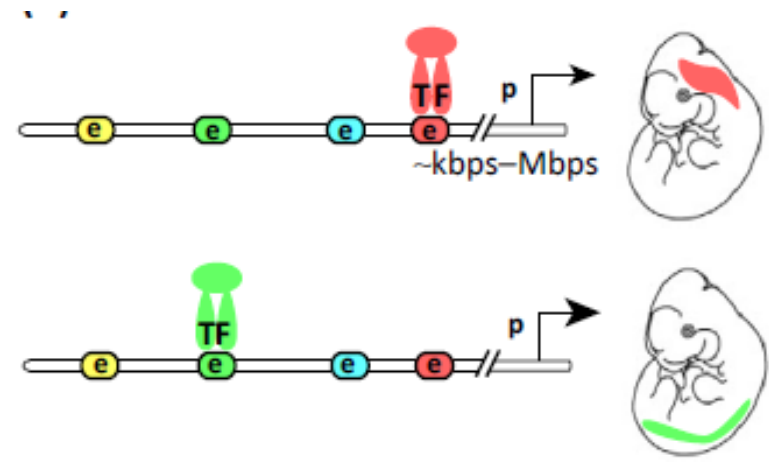
**SDTF: SIGNAL – DETERMINING  
TRANSCRIPTION FACTORS**

**CTF: COLLABORATING  
TRANSCRIPTION FACTORS**



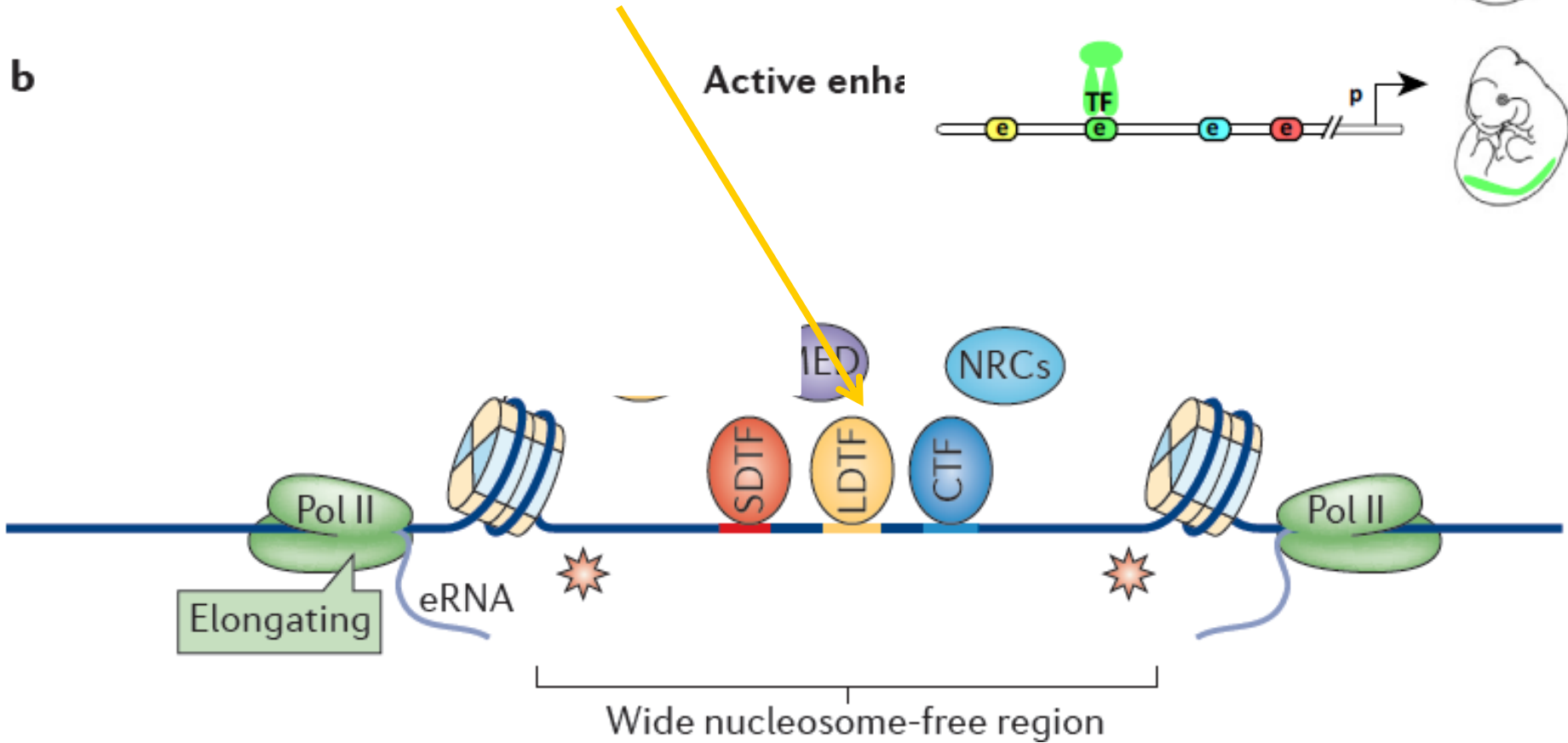
# TRANSCRIPTION FACTORS THAT BIND ENHANCERS

## LDTF: LINEAGE-DETERMINING TRANSCRIPTION FACTORS



b

Active enhancer

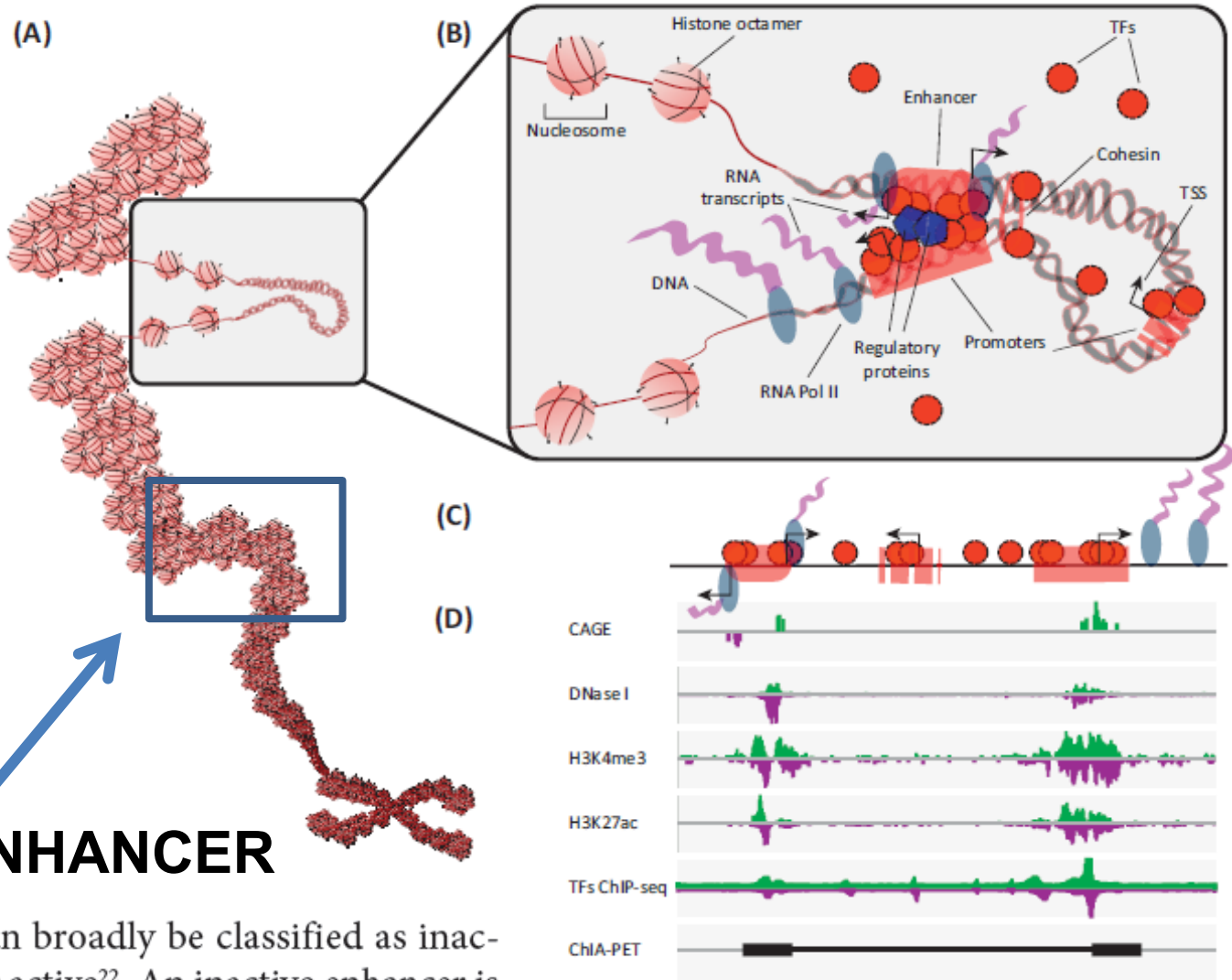


# Enhancer Activation

- **Transcription Factors bind specific genomic regions and allow access to other proteins remodelling chromatin**
- **Differentiation states and external stimuli induce enhancers activation**



# ACTIVE ENHANCER



# INACTIVE ENHANCER

Enhancer states can broadly be classified as inactive, primed, poised or active<sup>22</sup>. An inactive enhancer is essentially buried in compact chromatin and is devoid of transcription factor binding and histone modifications.

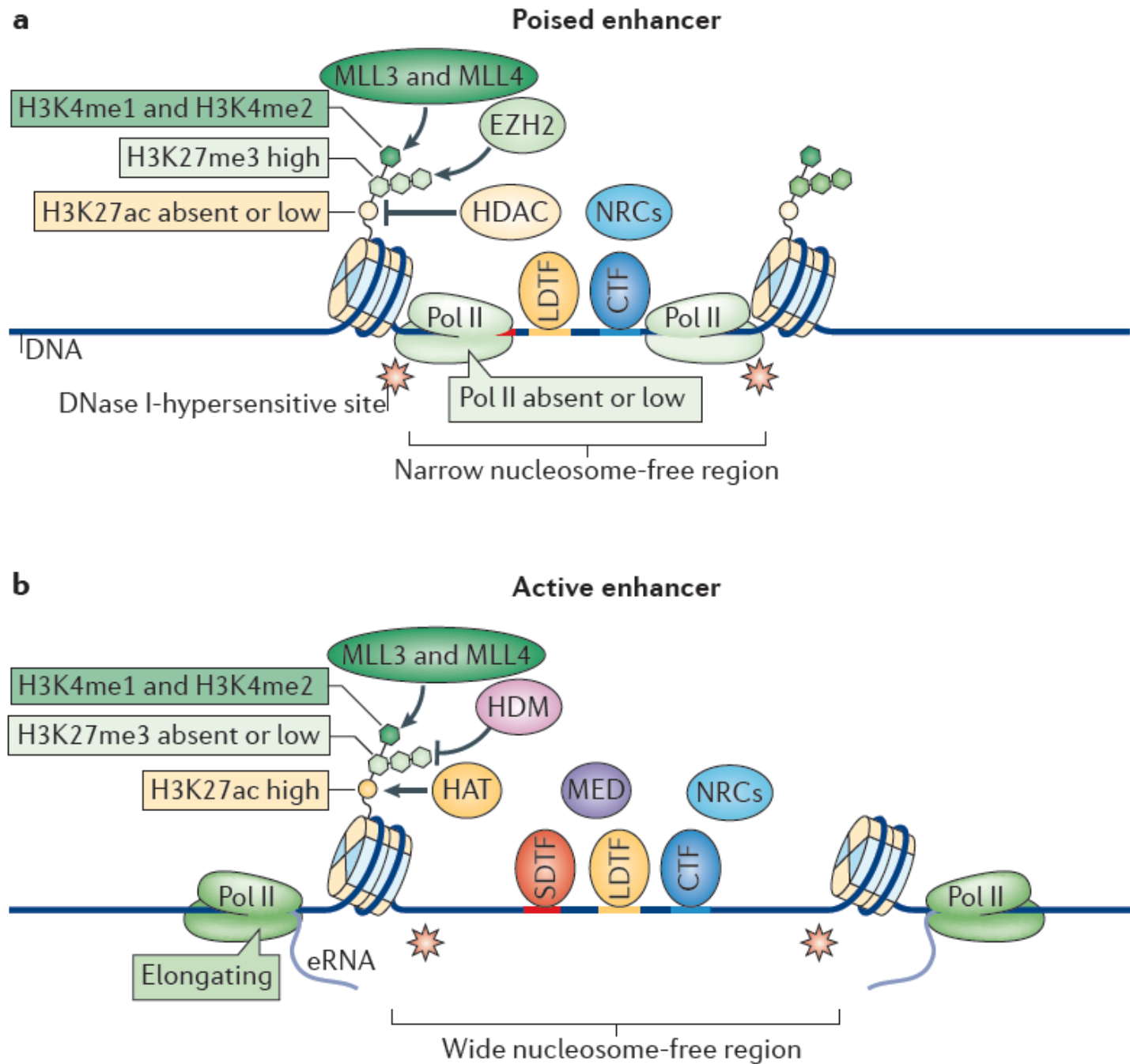
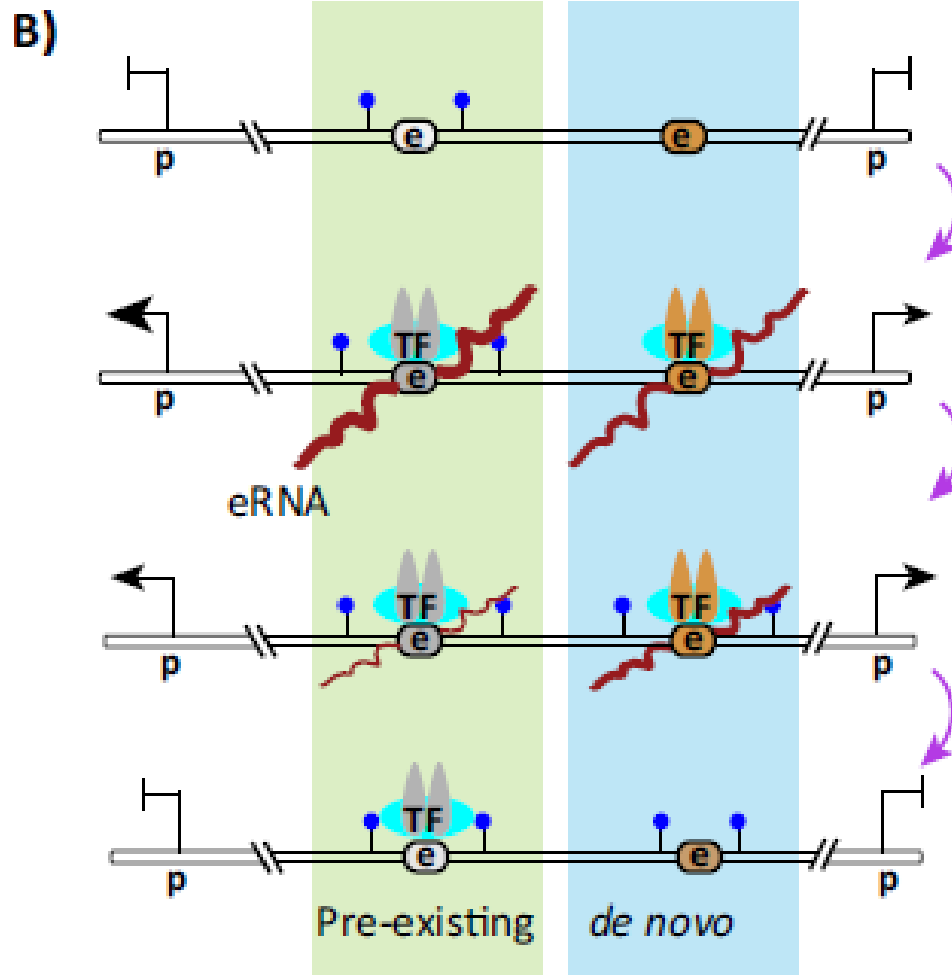


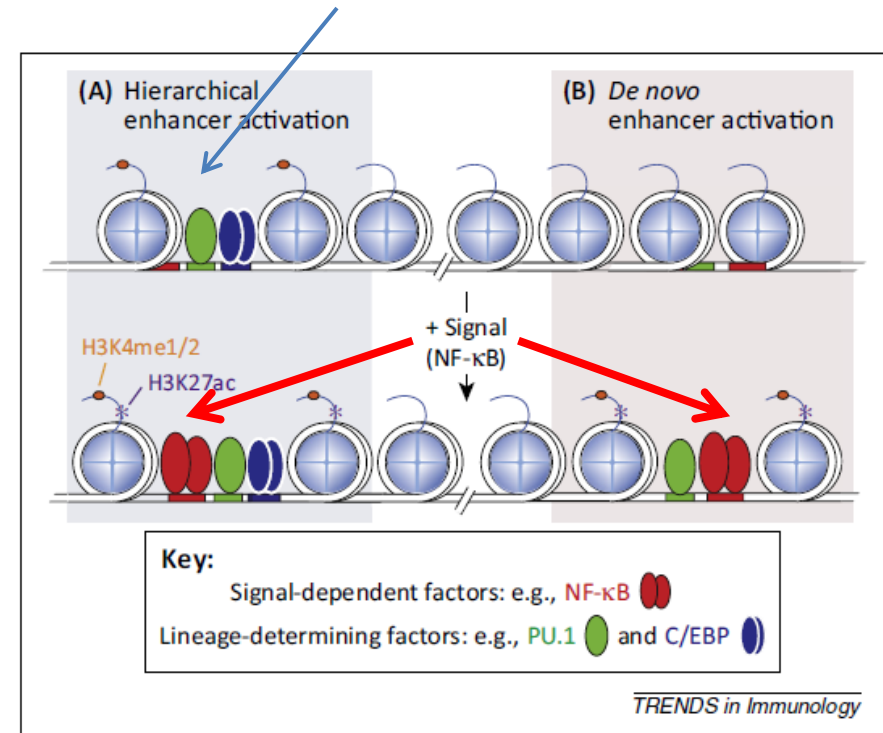
Figure 1 | **The anatomies of poised and active enhancers.** The characteristic features

# LDTF: LINEAGE – DETERMINING TRANSCRIPTION FACTORS



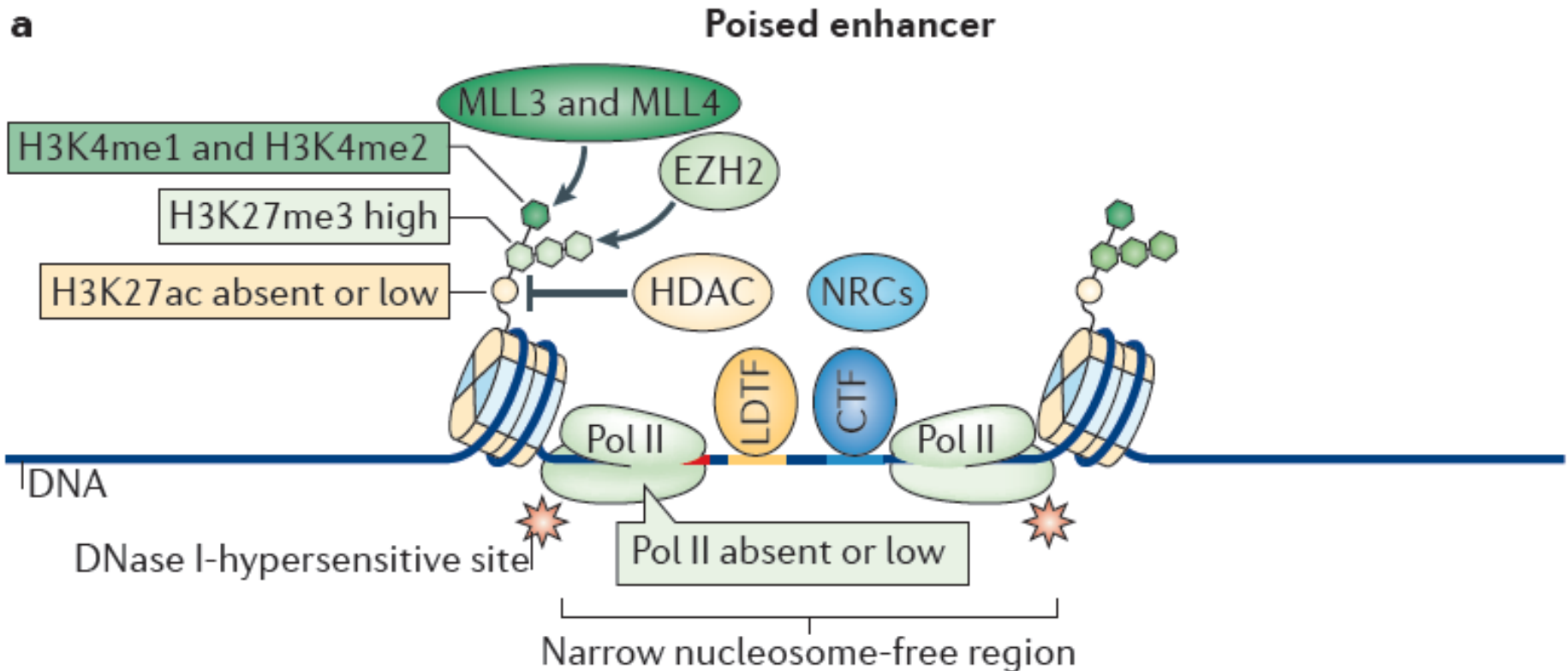
↑ Histone methylation ( $H3K4me^{1/2}$ )

Enhancers in stimulus-induced gene activation

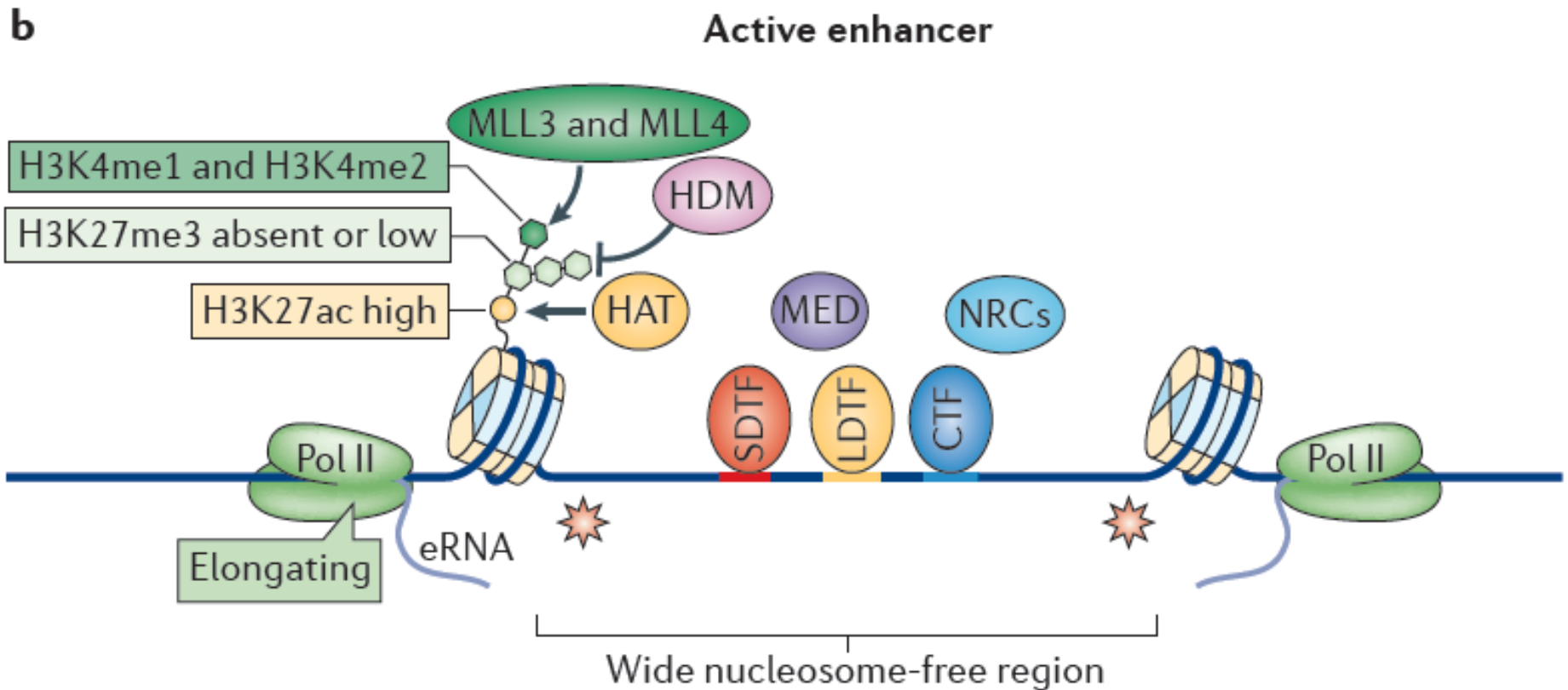


**Figure 3.** Chromatin transitions to active enhancers involve interactions between cell lineage-determining transcription factors and signal-dependent factors. **(A)** Enhancers primed by lineage-determining factors frequently require signal-dependent transcription factor binding to gain H3K27ac and become active. **(B)** Active enhancers can also be selected by interactions between signal-dependent factors and lineage-determining factors. Abbreviations: C/EBP, CCAAT/enhancer binding protein; NF- $\kappa$ B, nuclear factor- $\kappa$ B; PU.1, transcription factor originally named spleen focus forming virus (SFFV) proviral integration oncogene.

H3K4me1, H3K4me2, **lack histone acetylation and Pol II,**  
**high H3K27me3**  
mark **POISED ENHANCERS**

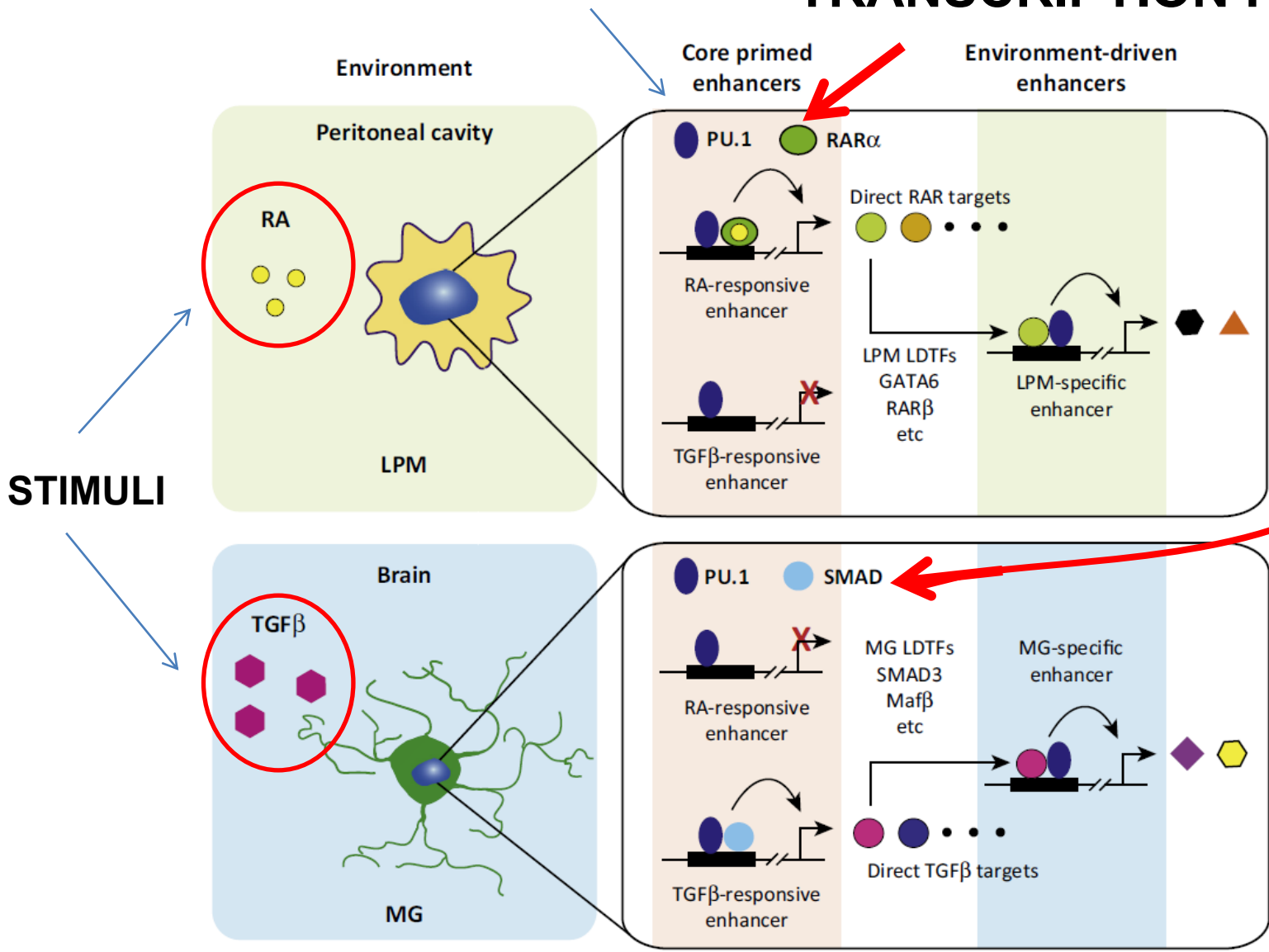


H3K4me1, H3K4me2, **high H3K27Ac**, lack H3K27me3,  
**presence of Pol II and RNA transcript**  
mark **ACTIVE ENHANCERS**



# LDTF: LINEAGE –DETERMINING TRANSCRIPTION FACTORS

# SDTF: SIGNAL –DETERMINING TRANSCRIPTION FACTORS



# Enhancer Selection

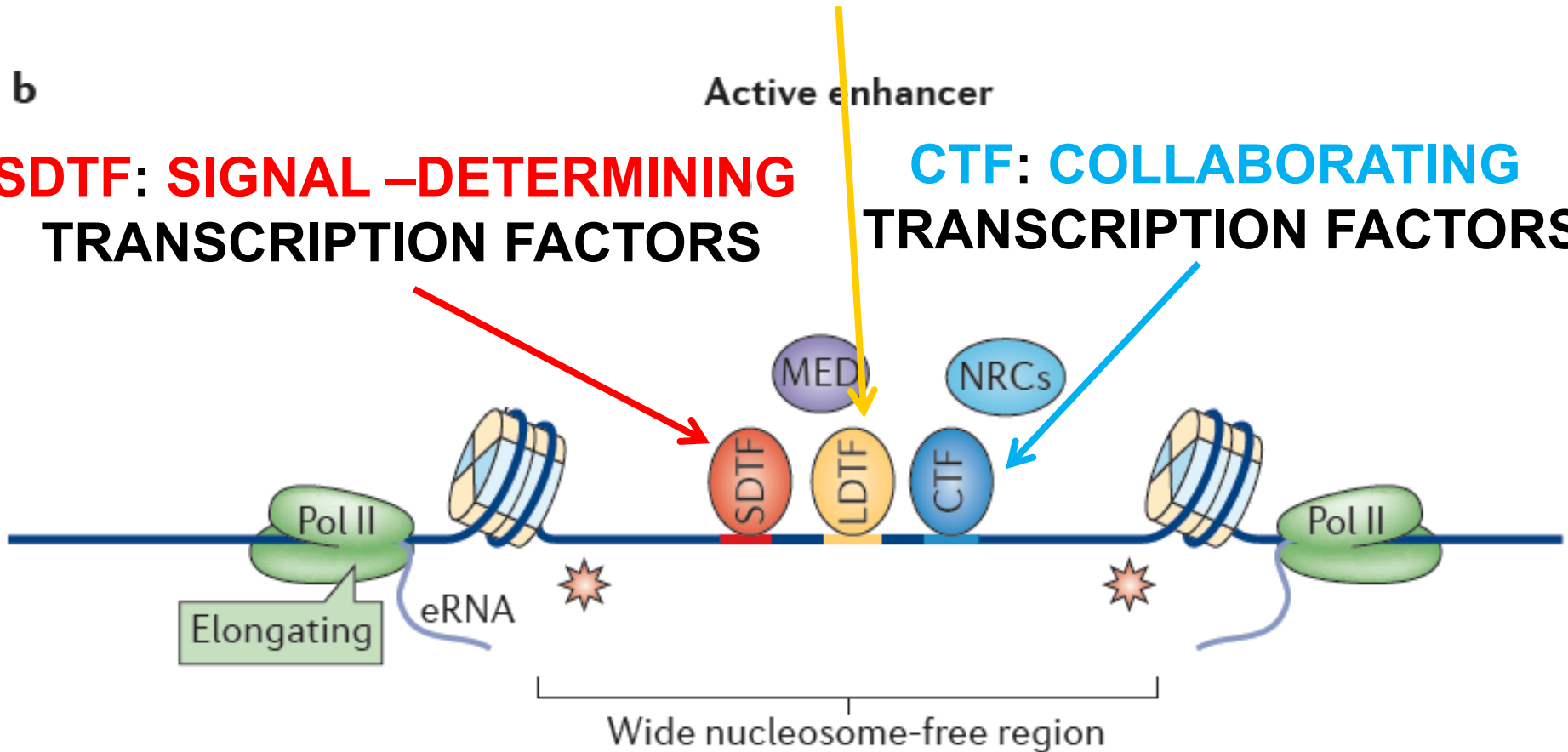
- **The role of lineage-determining transcription factors.**
- **The role of signal-dependent transcription factors.**

# LDTF: LINEAGE – DETERMINING TRANSCRIPTION FACTORS

b

**SDTF: SIGNAL – DETERMINING  
TRANSCRIPTION FACTORS**

**CTF: COLLABORATING  
TRANSCRIPTION FACTORS**

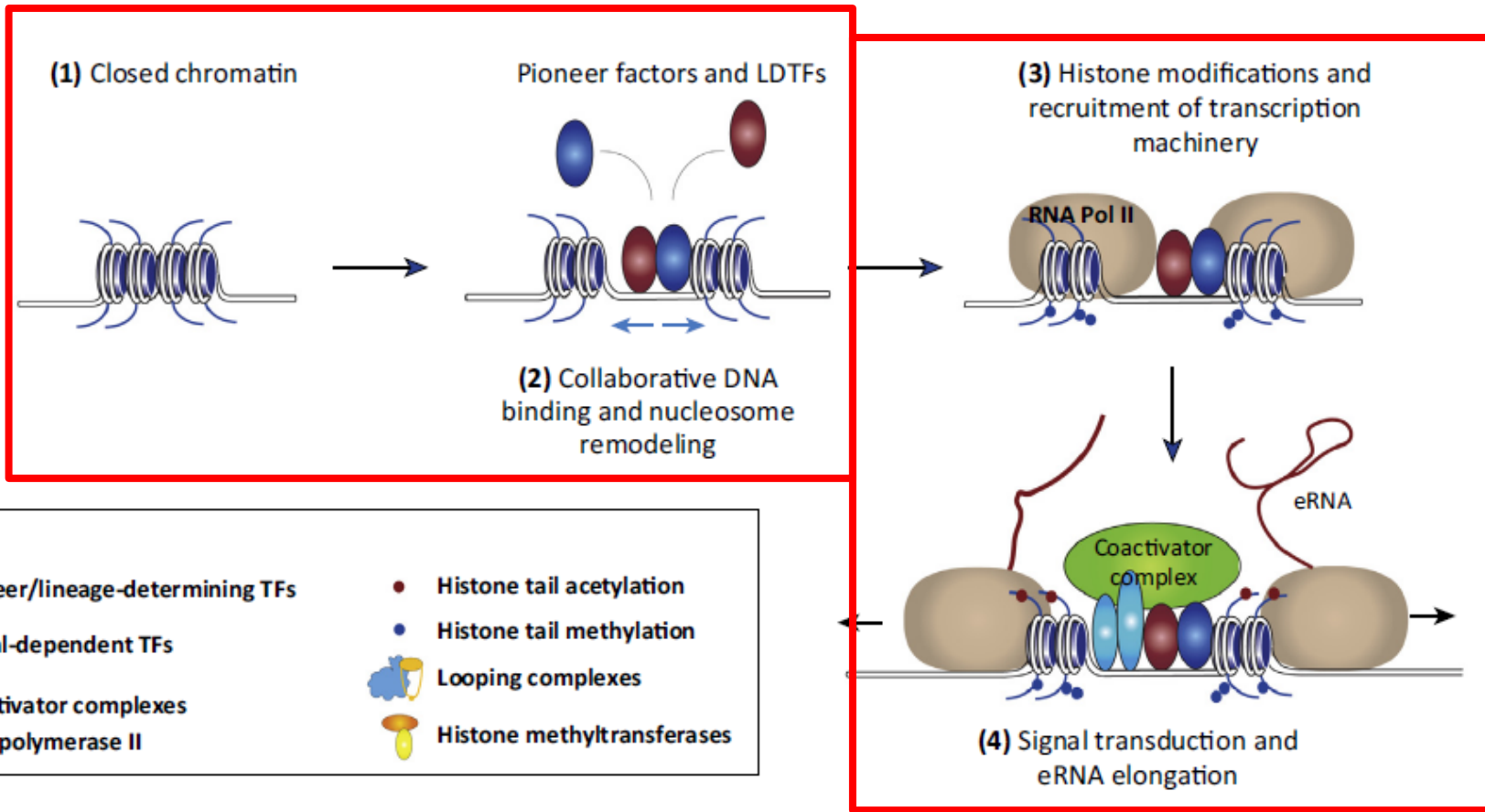




# Pioneer Factors and Lineage-determining Transcription Factors leads to nucleosome remodeling and increased chromatin accessibility



(A)



Pioneer Factors and Lineage-determining Transcription Factors leads to histone modifications and basal transcription machinery

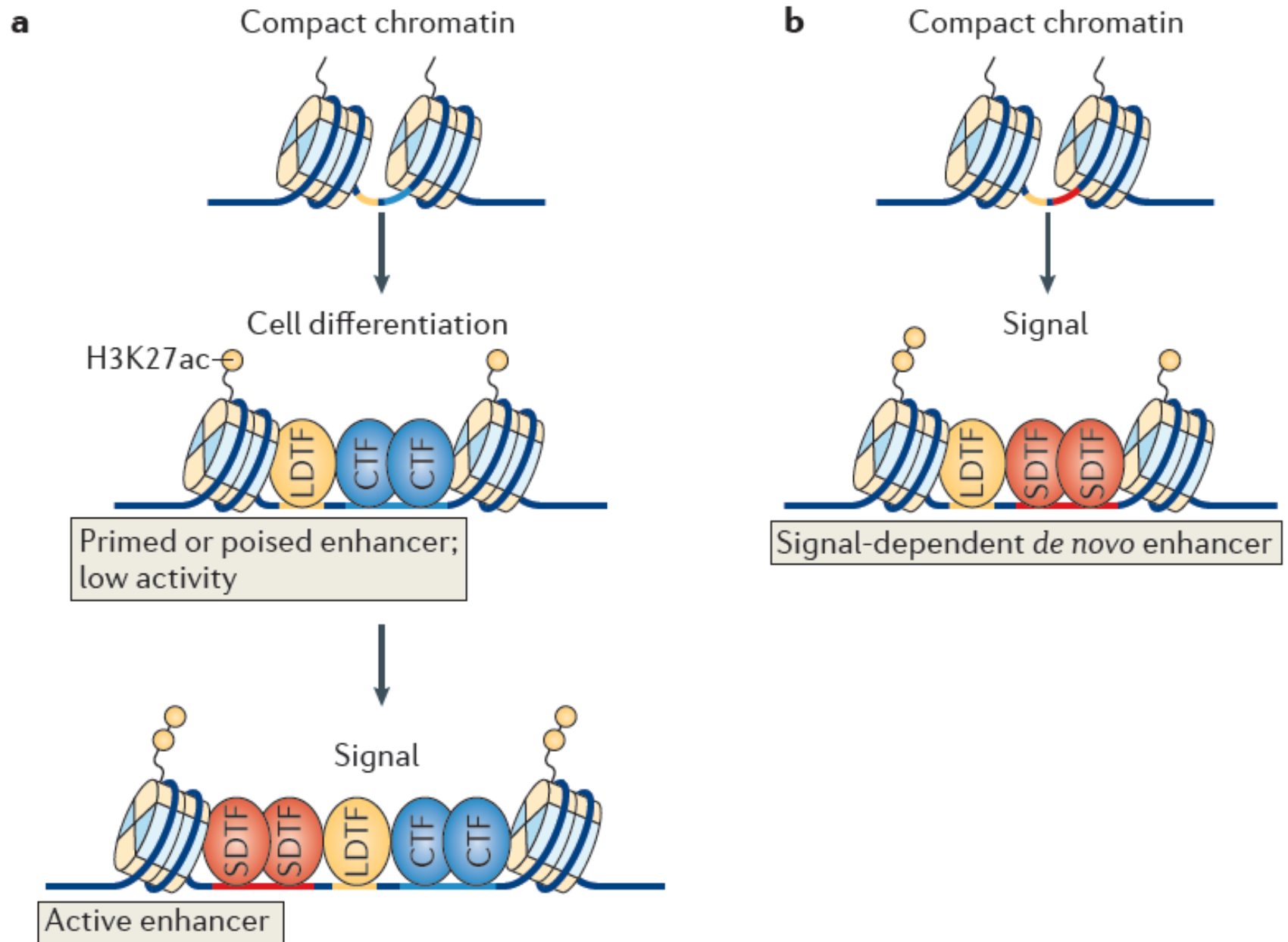
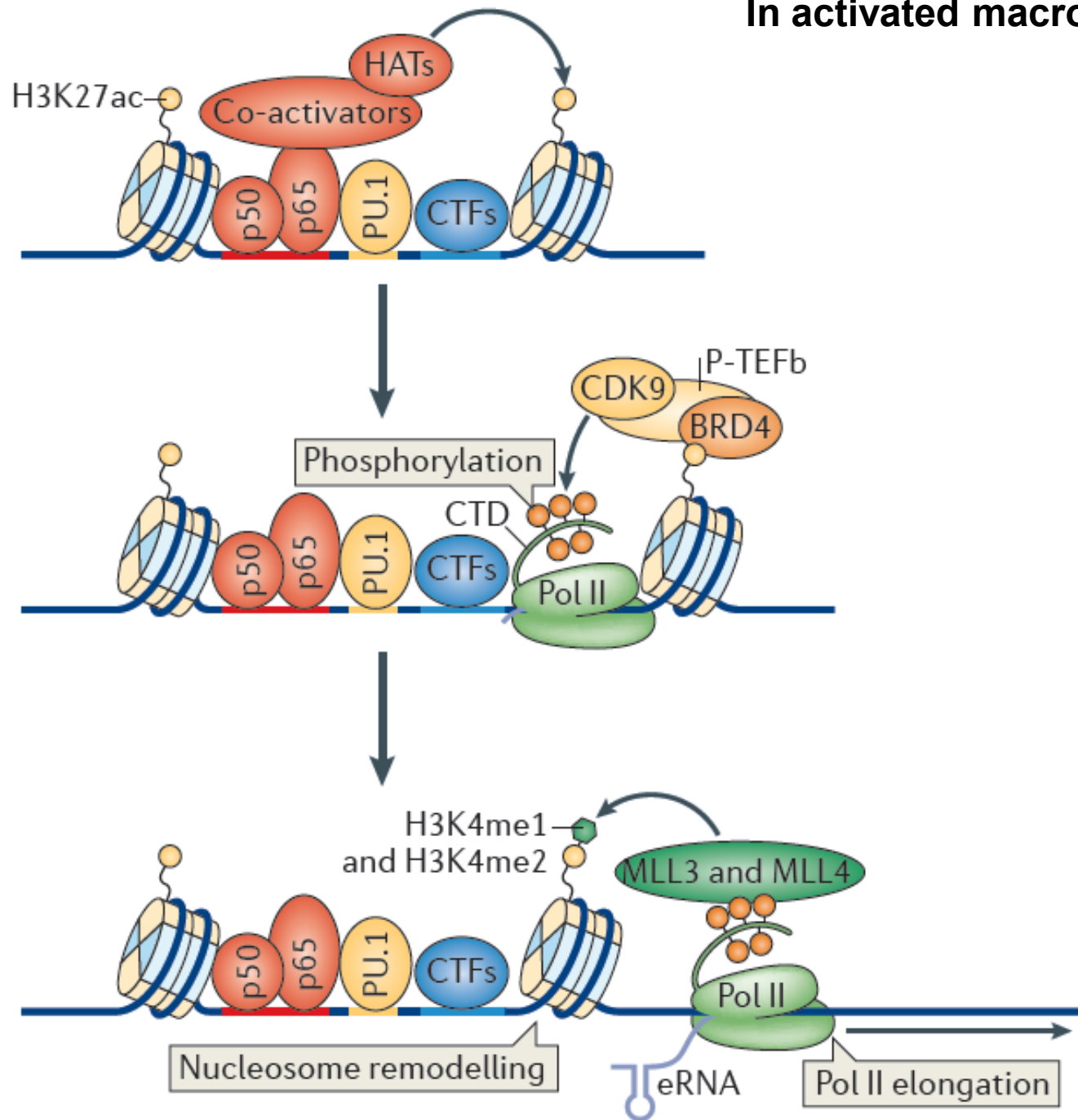


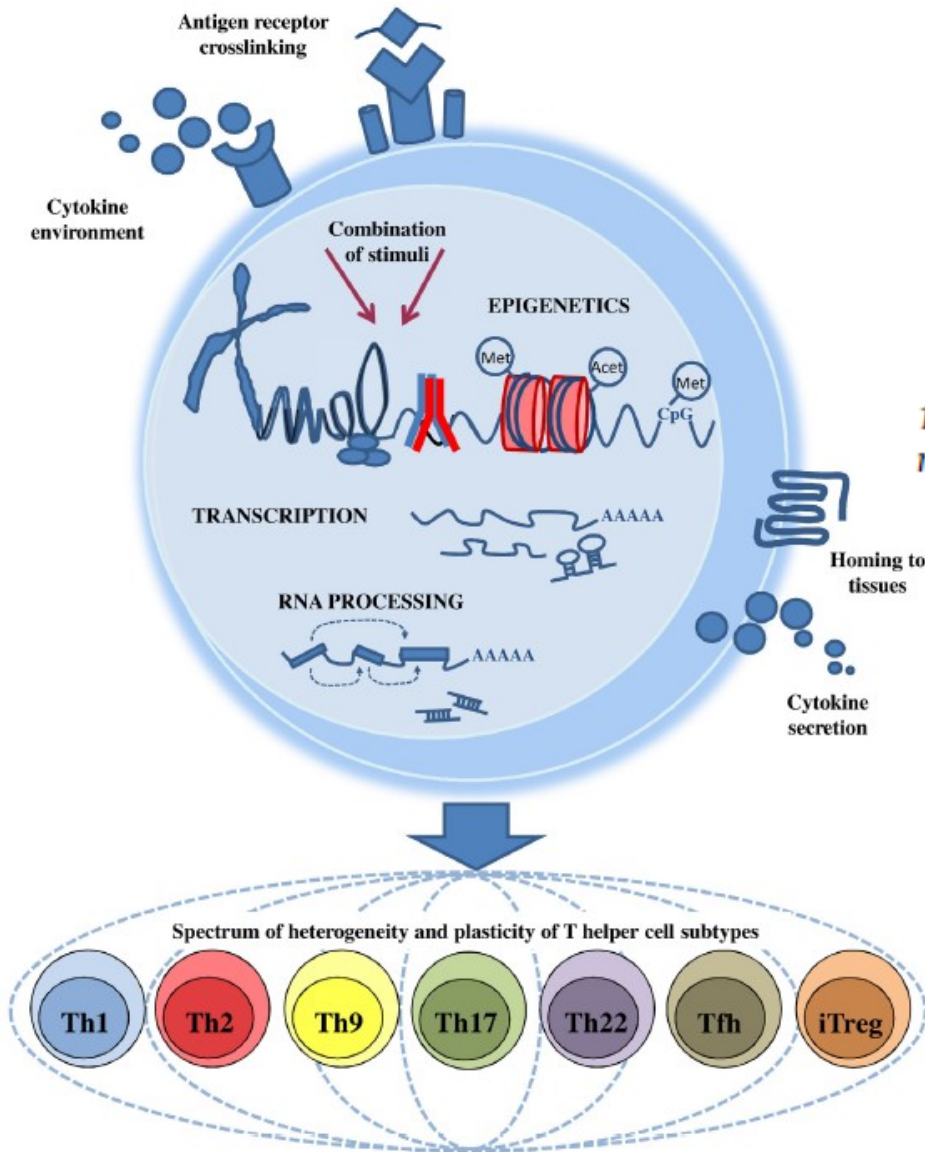
Figure 3 | **Cell type-specific enhancer selection and activation.** **a** | Collaborative

# In activated macrophages



# Early T helper cell programming of gene expression in human

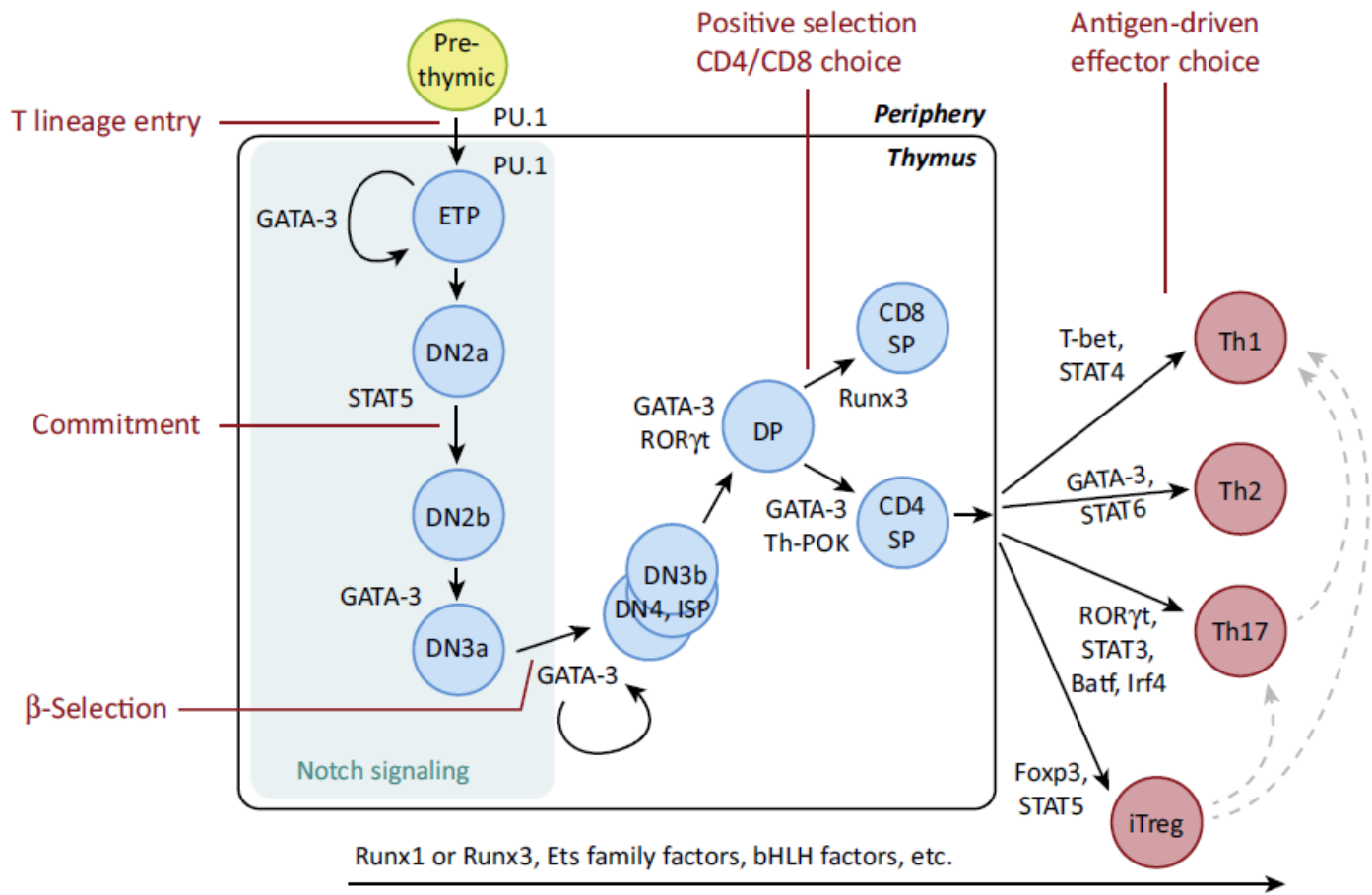
Soile Tuomela, Riitta Lahesmaa\*



1.1. *Transcriptional regulation of human Th cell priming*

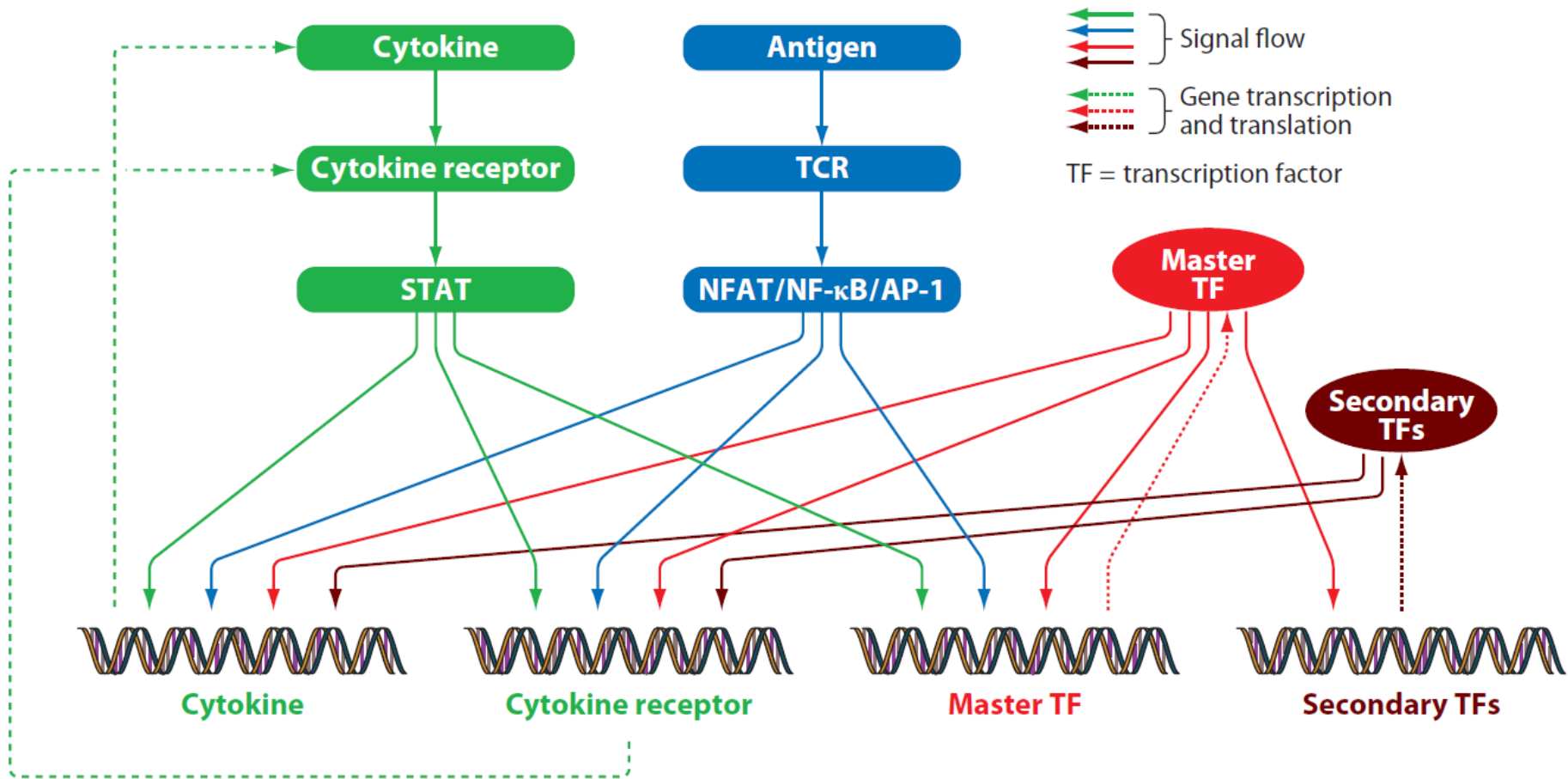
1.2. *Epigenetic regulation of Th cell priming in human*

1.3. *Regulation of Th cell differentiation by RNA processing and non-coding RNAs*



# Differentiation of Effector CD4 T Cell Populations\*

Jinfang Zhu, Hidehiro Yamane, and William E. Paul

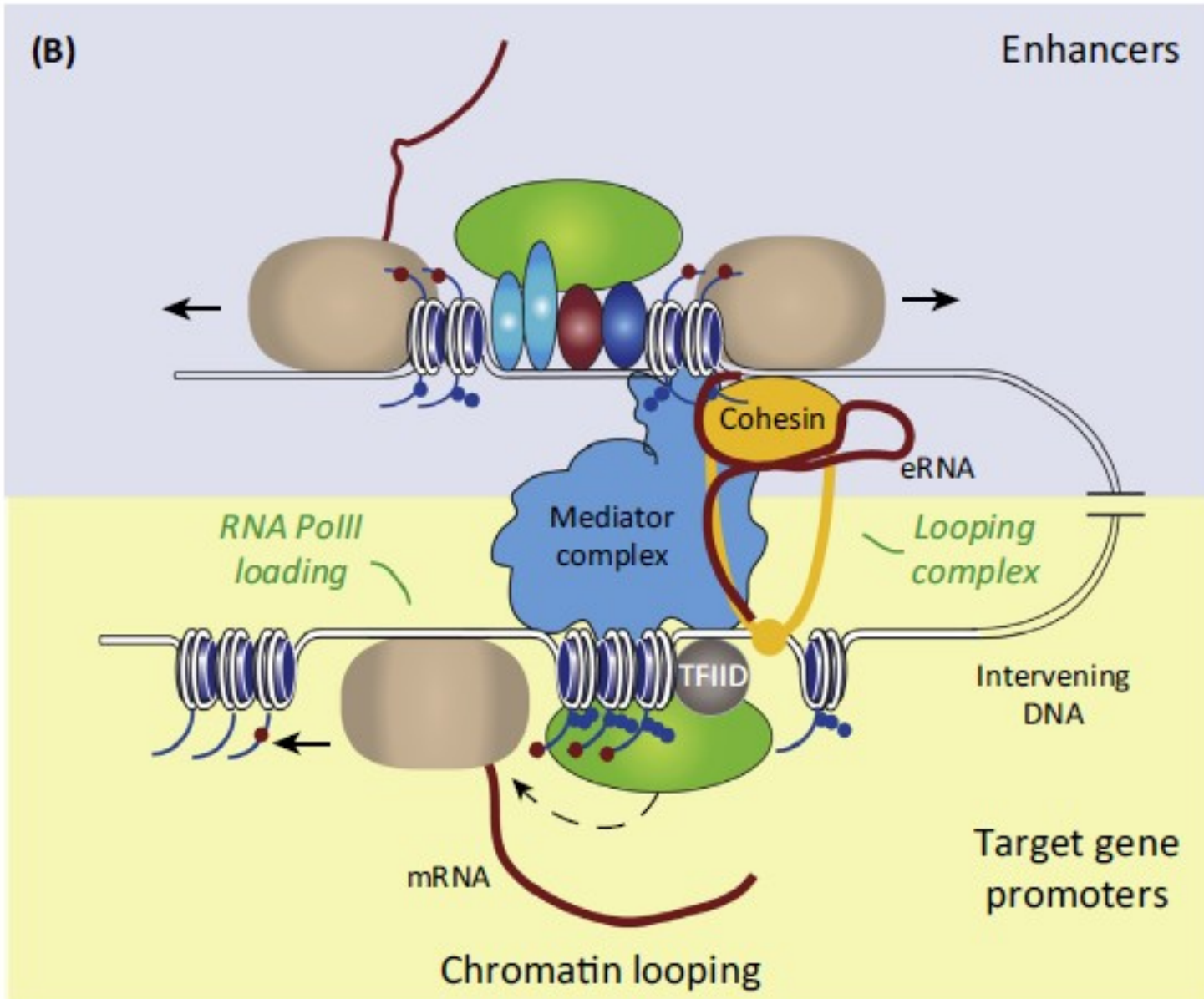


# Enhancer RNAs and regulated transcriptional programs

Michael T.Y. Lam<sup>1</sup>, Wenbo Li<sup>2</sup>, Michael G. Rosenfeld<sup>2</sup>, and Christopher K. Glass<sup>1,2</sup>

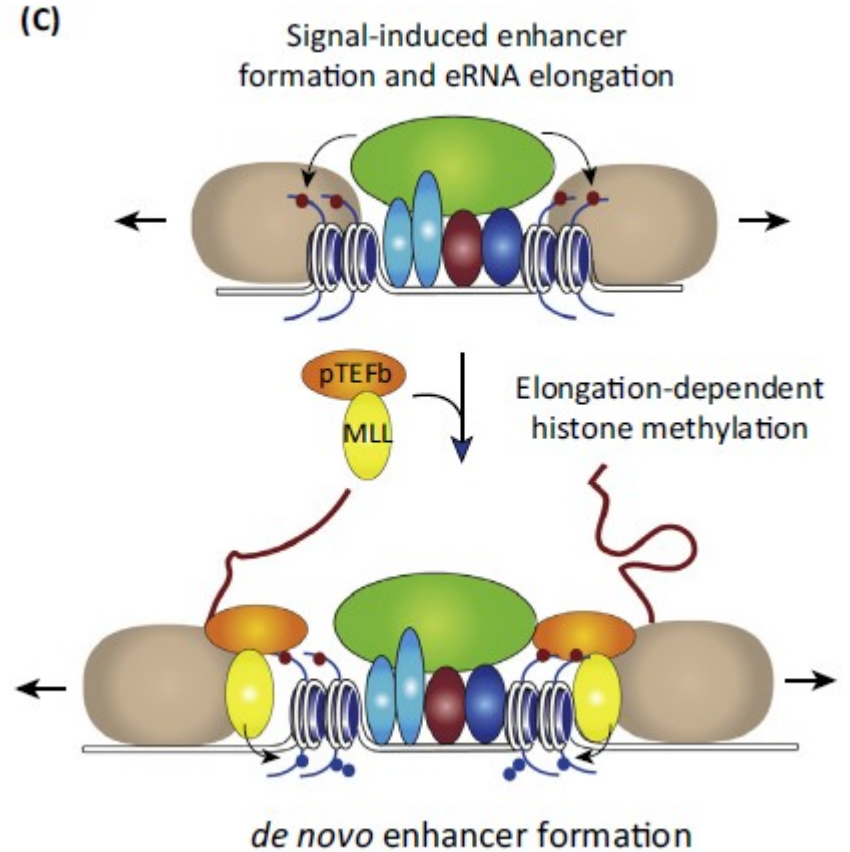
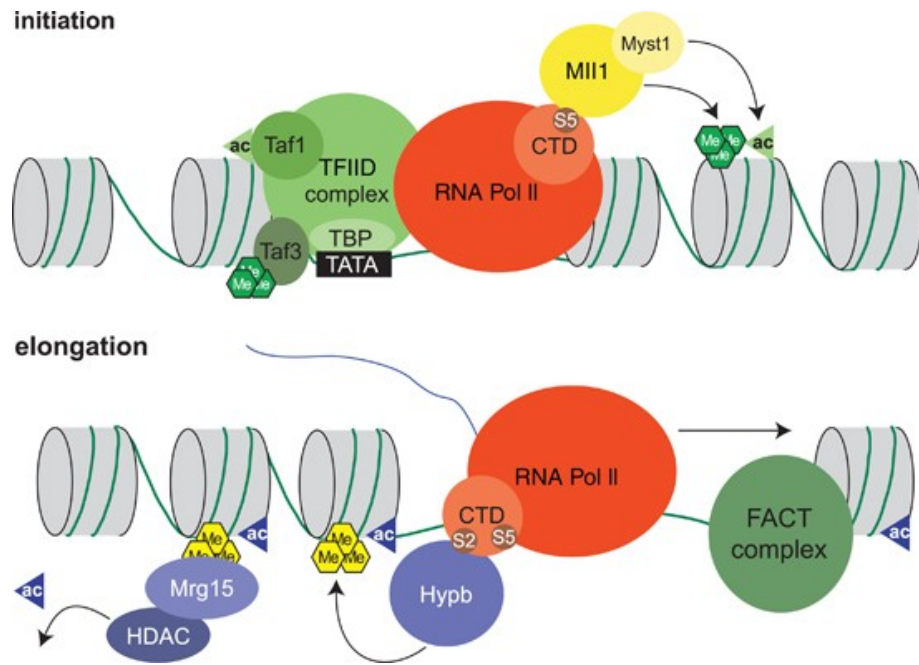
*Trends in Biochemical Sciences* April 2014, Vol. 39, No. 4

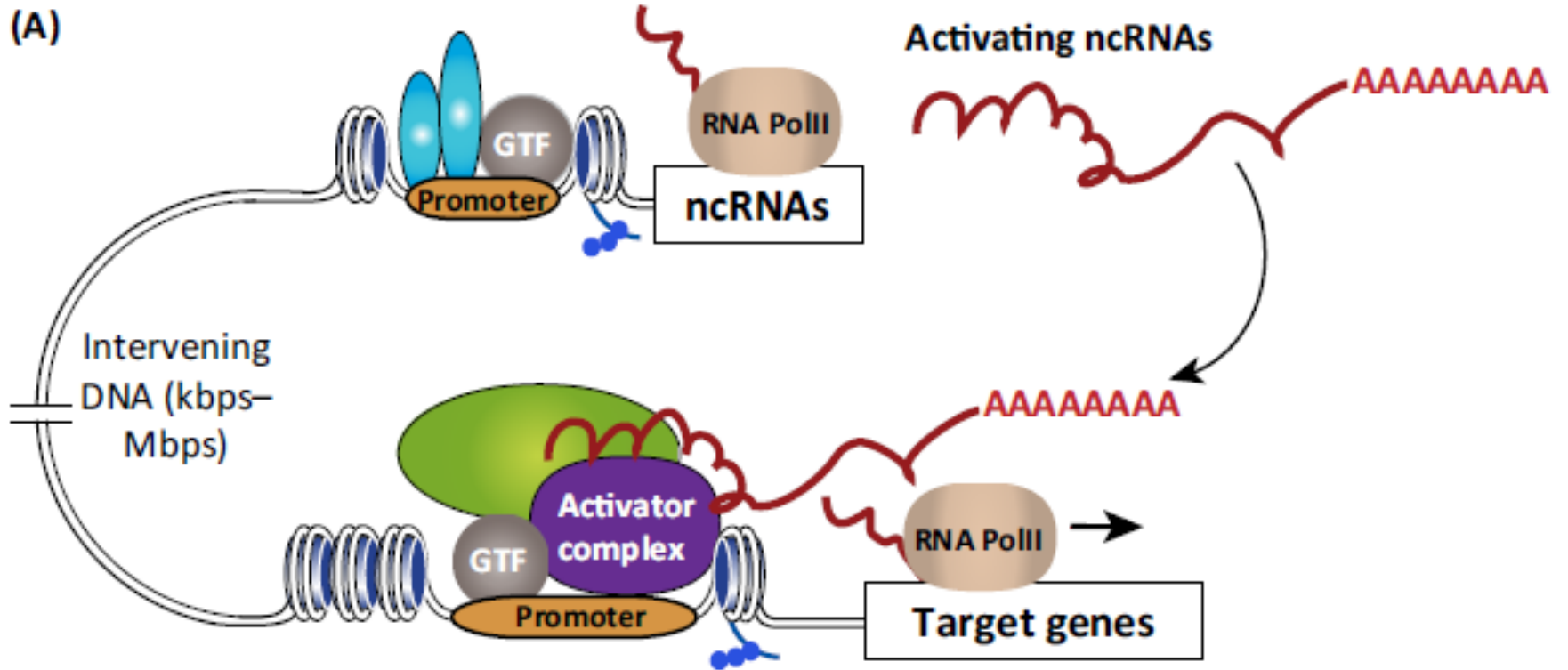
# eRNA mediates the long range interactions





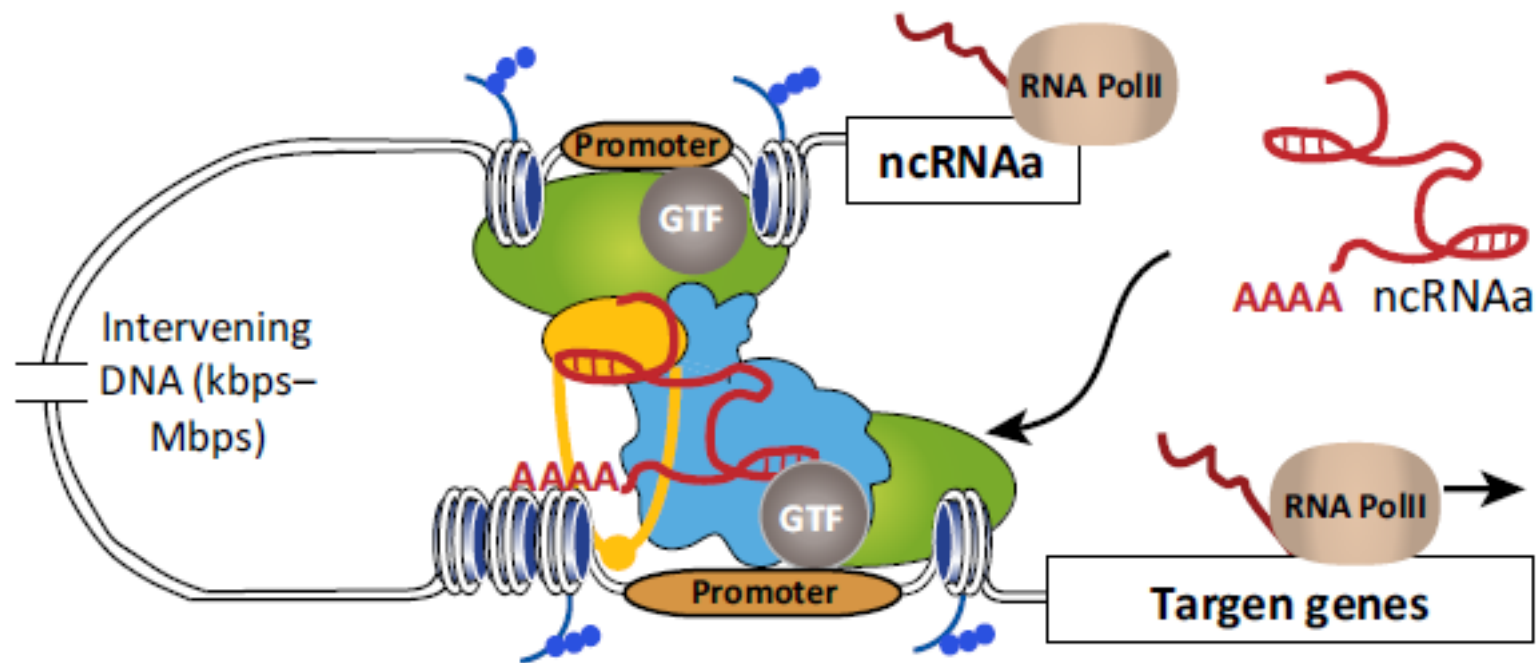
# Molecular mechanisms that underline enhancer activation





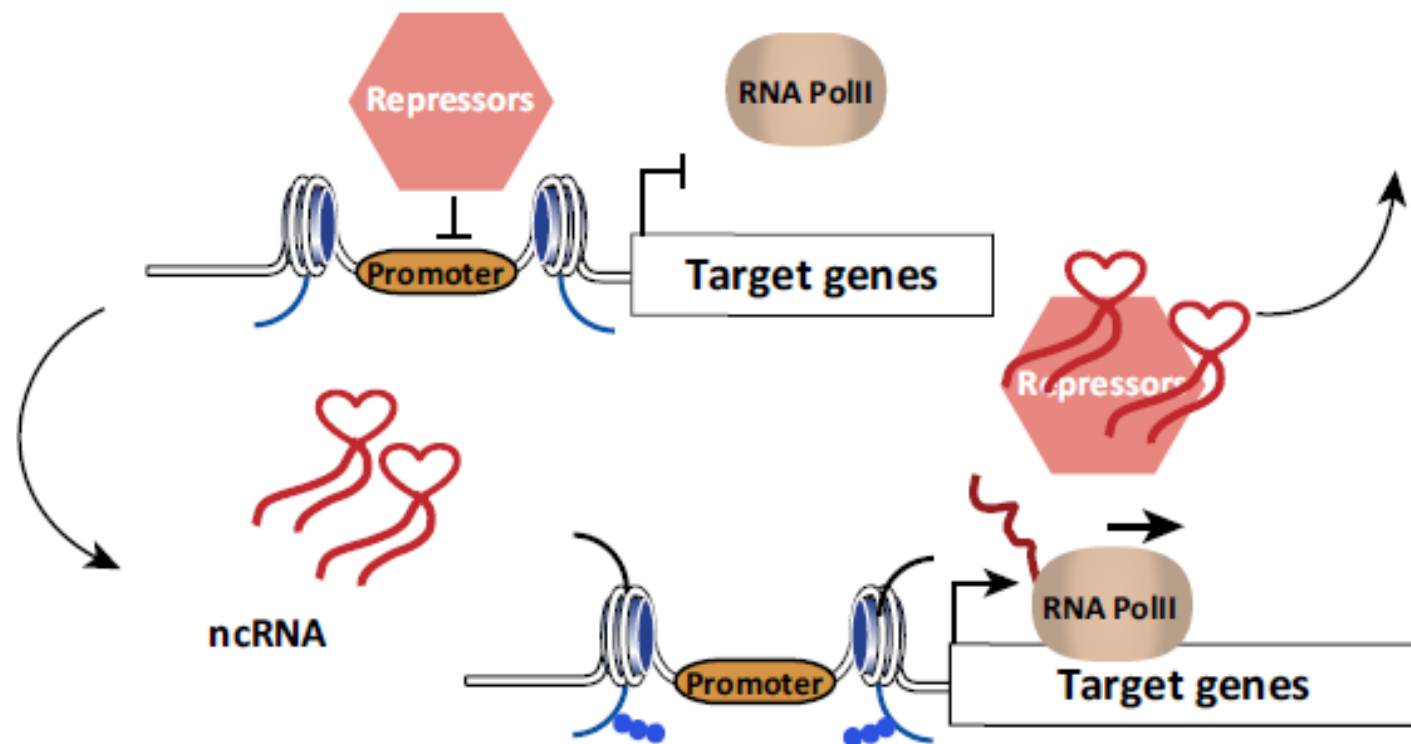
**Model 1: ncRNAs collaborate with transcriptional activators**

(B)



**Model 2: ncRNAs modulate chromatin loops**

(c)



### Model 3: ncRNAs evict transcriptional repressors

**Key:**

● Histone tail methylation



Signal dependent TFs



Looping complexes

● Coactivator complexes



General TFs



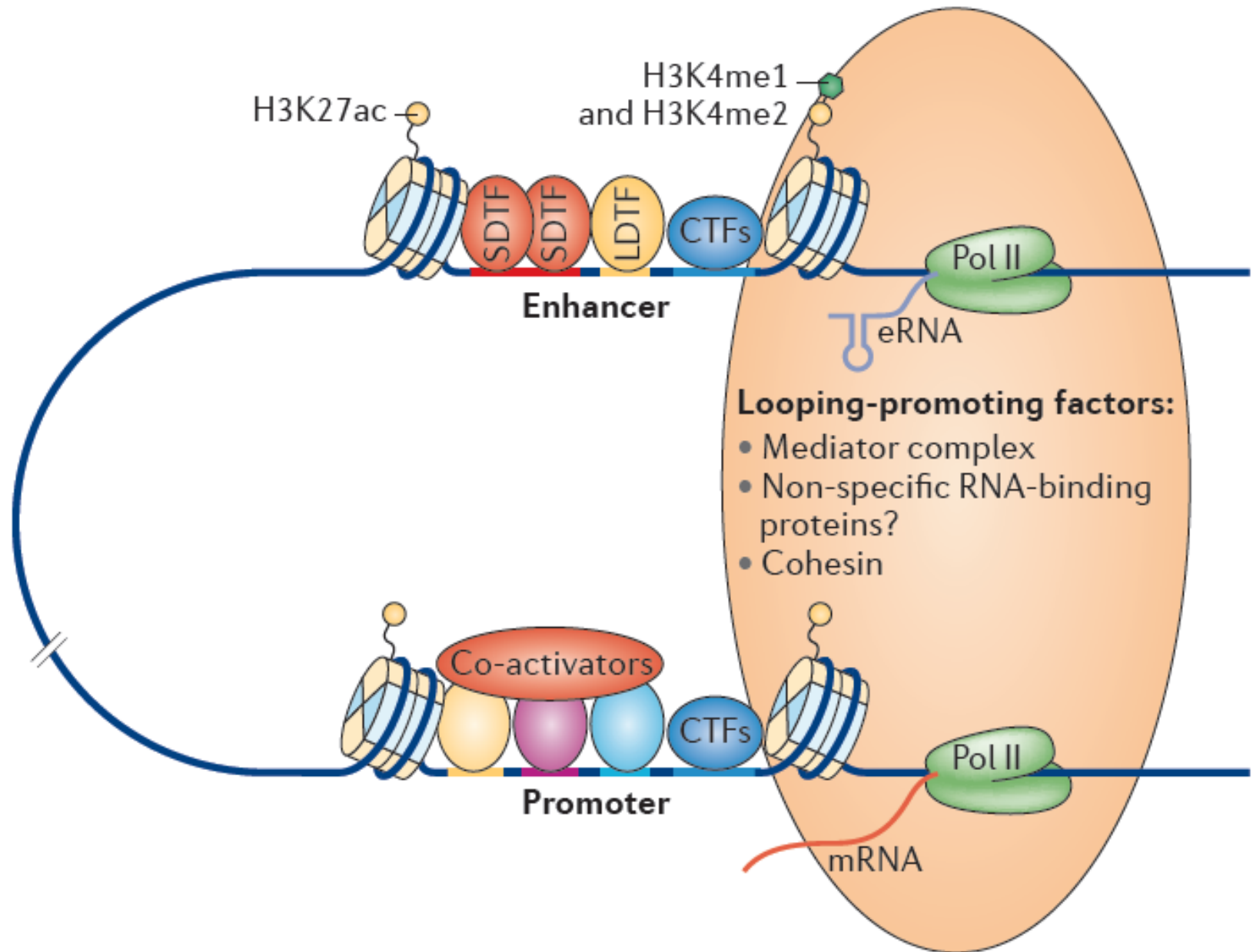
RNA polymerase II

● Transcriptional repressors

# Enhancer Function

- **Chromatin looping**
- **Super-enhancers, cluster of enhancers, key player in the cell identity and differentiation**

# CHROMATIN LOOPING



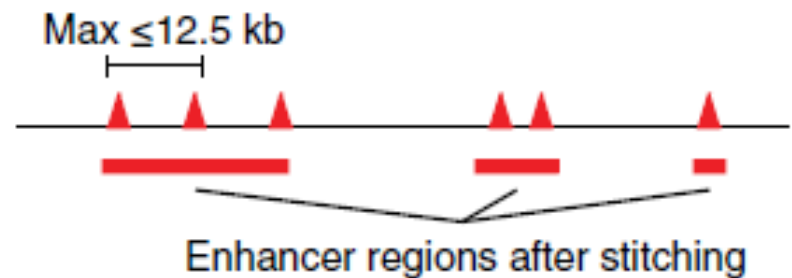
# Super-enhancers.

**a**

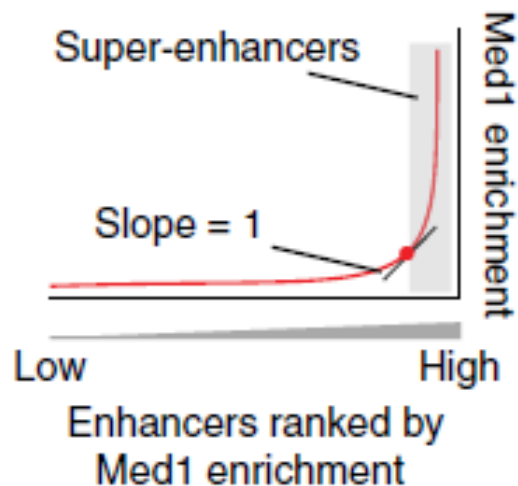
Step 1. Identification of enhancer locations



Step 2. Clustering of enhancers

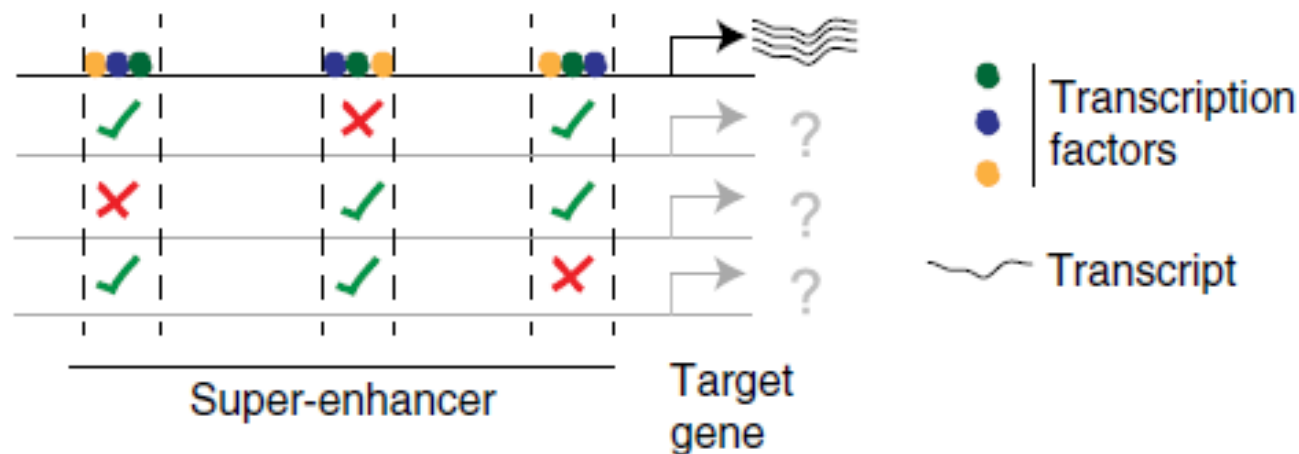


Step 3. Identify super-enhancers



**b**

Factor used for step 1	Factor used for step 3	Reference
Oct4 + Sox2 + Nanog, Pu.1	Med1	Whyte <i>et al.</i>
MyoD, T-bet, C/EBP $\alpha$	MyoD, T-bet, C/EBP $\alpha$	Whyte <i>et al.</i>
H3K27ac	H3K27ac	Hnisz <i>et al.</i>
Med1	Med1	Loven <i>et al.</i>



**Figure 2** Schematic of an experimental approach to characterizing super-enhancers. Use of genome editing tools, such as the CRISPR-Cas9 system, provides a methodology to create a minimal targeted deletion to test the activity of specific putative enhancers within super-enhancer loci by assessing the consequences of genetic deletions on gene activity.



# Making the case for chromatin profiling: a new tool to investigate the immune-regulatory landscape

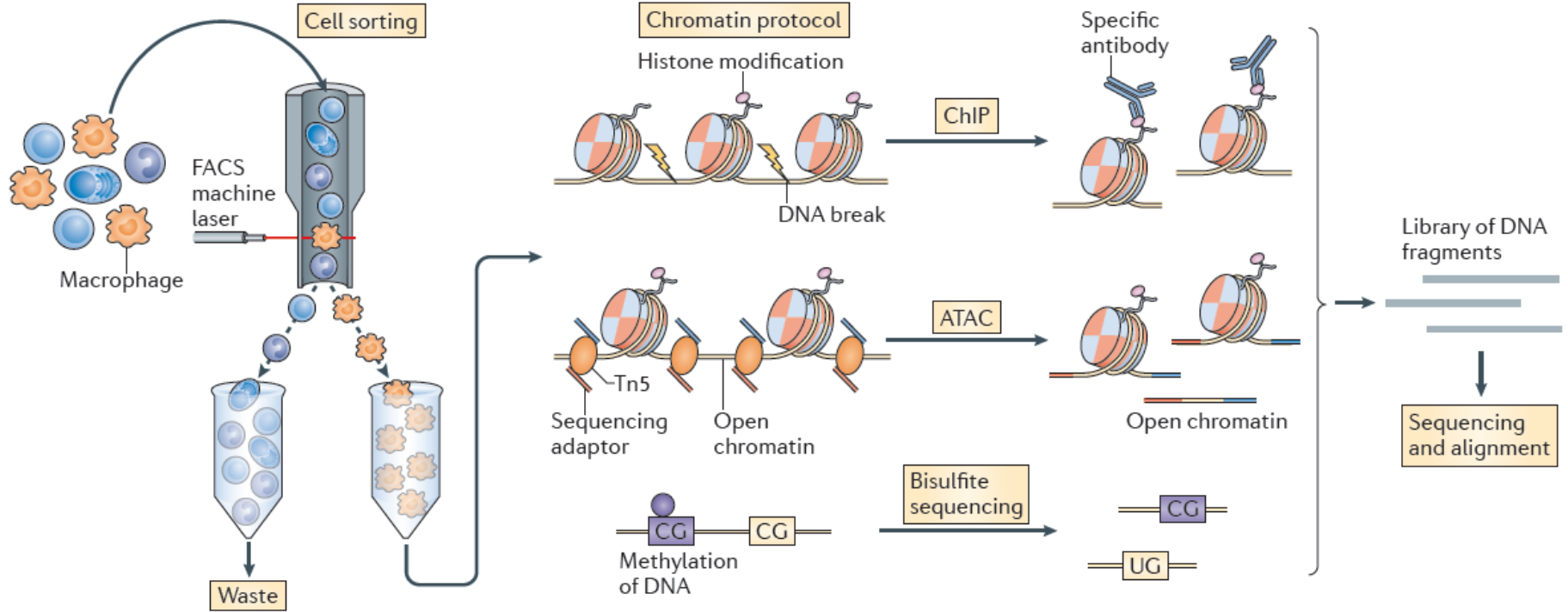
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*Deborah R. Winter, Steffen Jung and Ido Amit*

Abstract | Recent technological advances have enabled researchers to accurately and efficiently assay the chromatin dynamics of scarce cell populations. In this Opinion article, we advocate the application of these technologies to central questions in immunology. Unlike changes to other molecular structures in the cell, chromatin features can reveal the past (developmental history), present (current activity) and future (potential response to challenges) of a given immune cell type; chromatin profiling is therefore an important new tool for studying the immune-regulatory networks of health and disease.

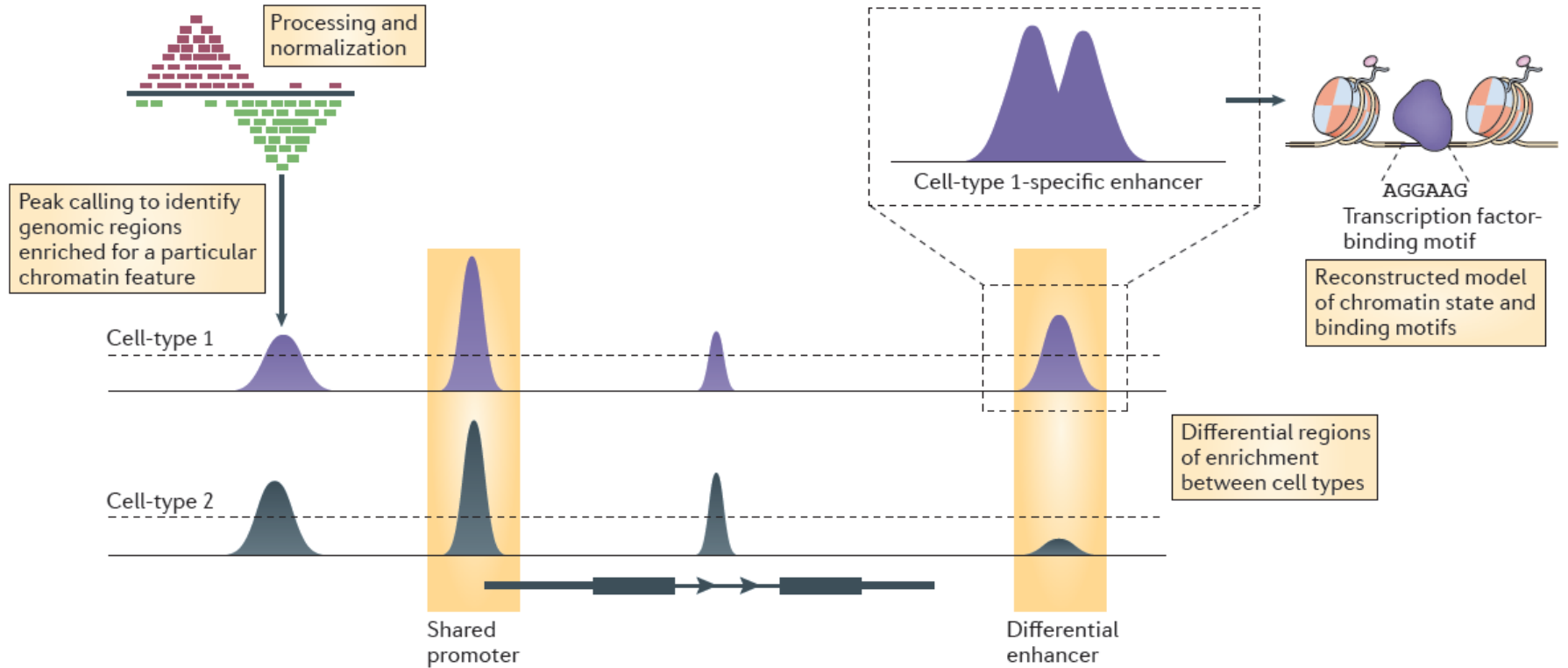
# Methods for identification of genomic regulatory regions

## a Experimental protocol



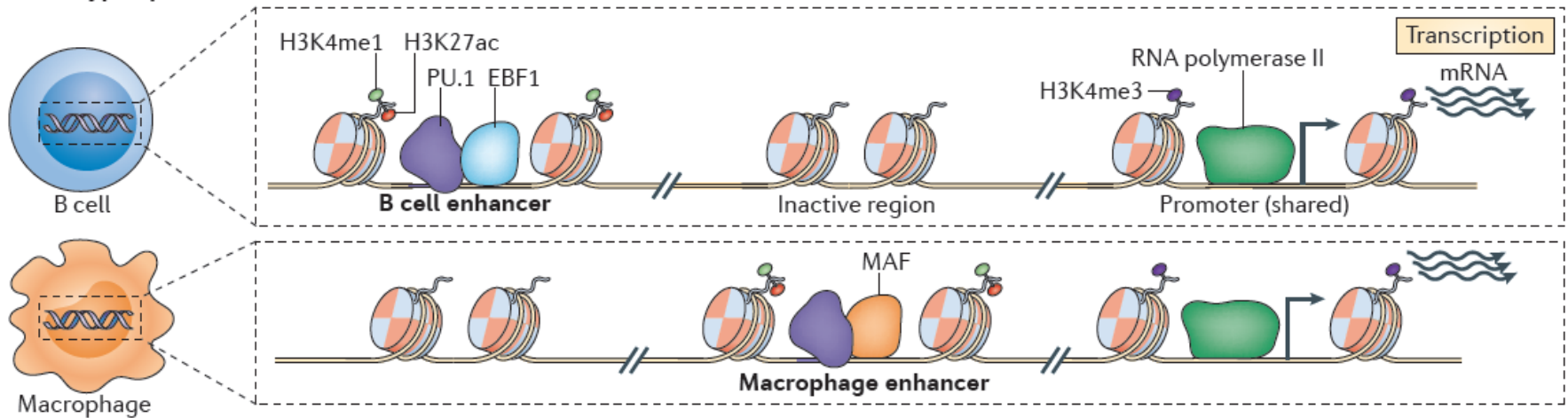
# From reads to DNA elements function

## b Data interpretation



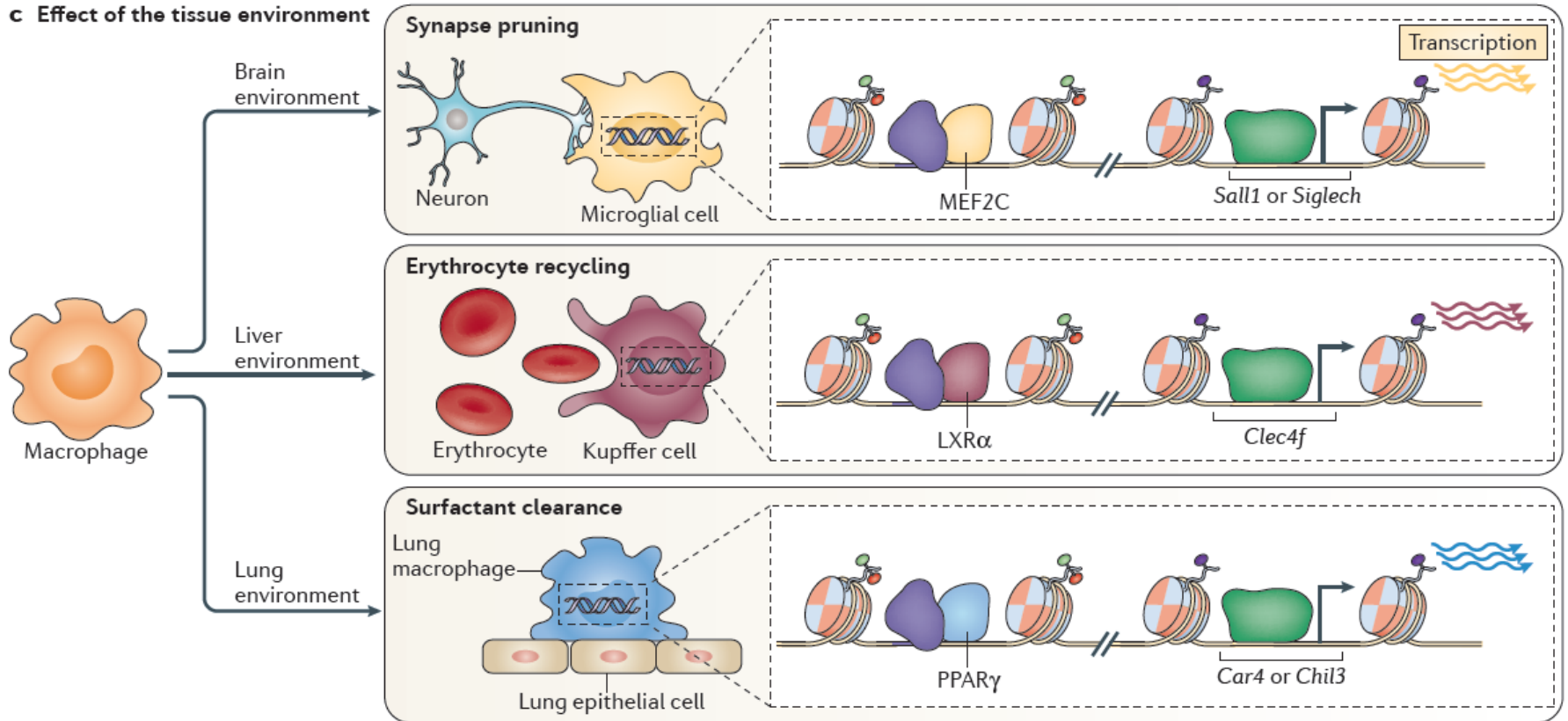
# Cell-type-specific enhancers to regulate same genes

## a Cell-type-specific enhancers



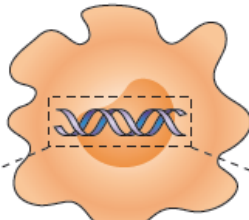
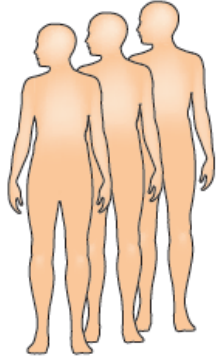
# Effect of the tissue environment

## c Effect of the tissue environment



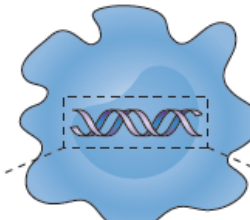
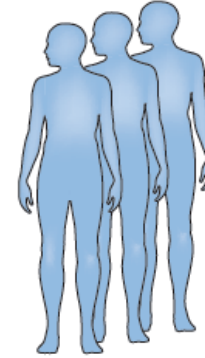
# Association of human chromatin data and susceptibility to immune disease

Healthy cohort

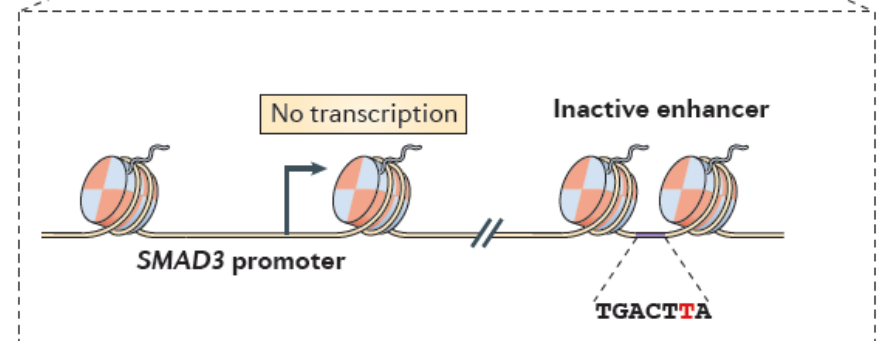
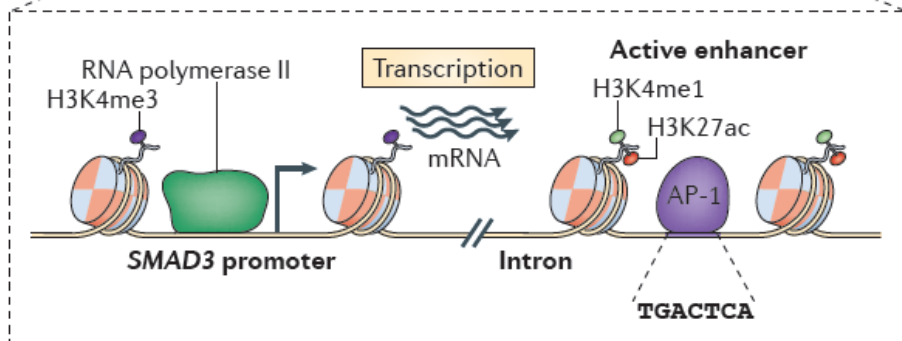


Monocyte

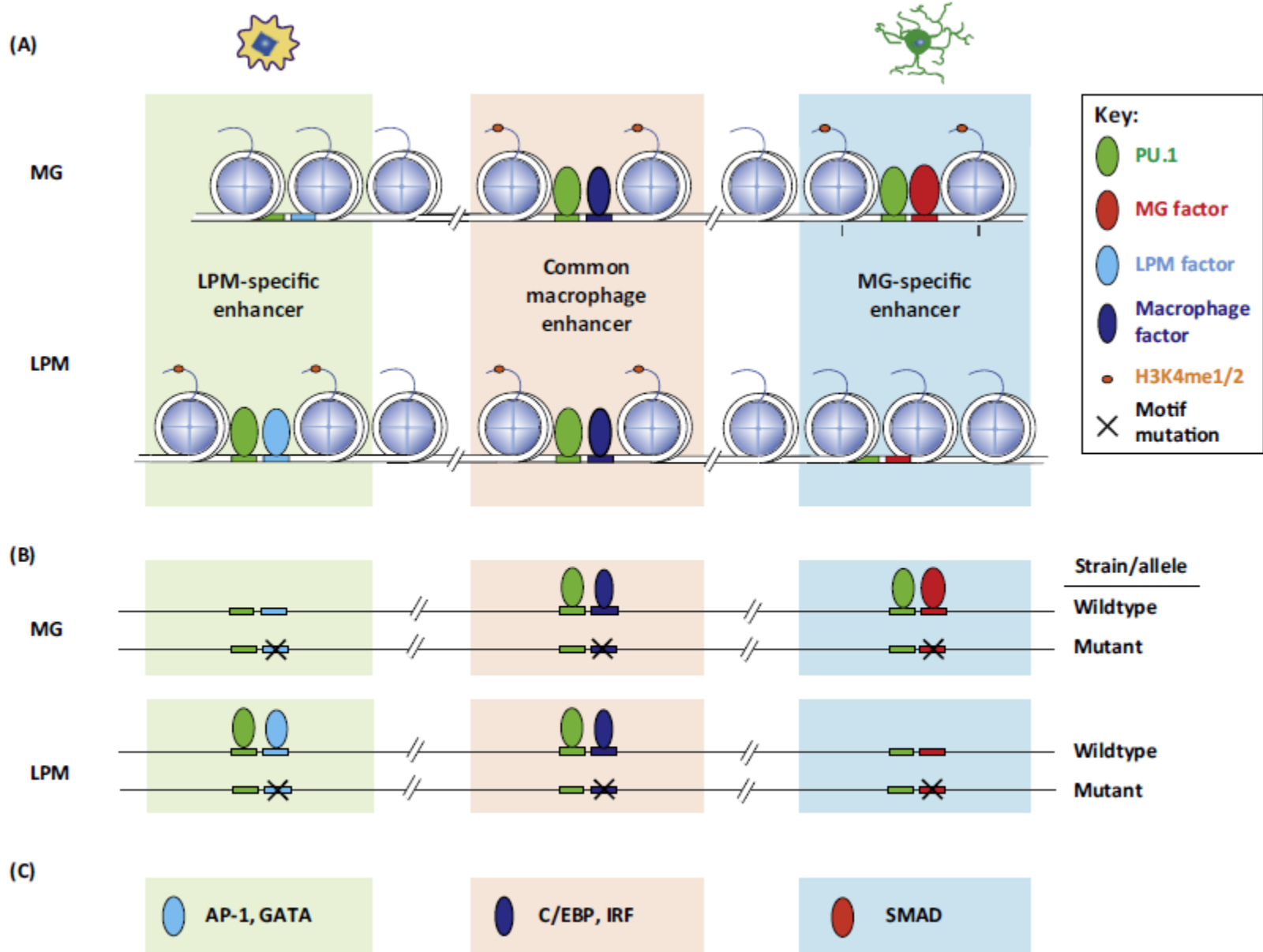
Disease cohort



Altered monocyte



# NATURAL GENETIC VARIATION IS ASSOCIATED WITH TF BINDING



# ACTIVATION

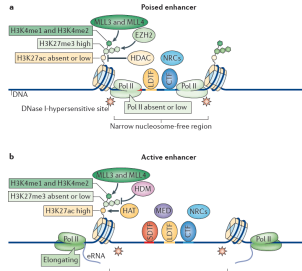
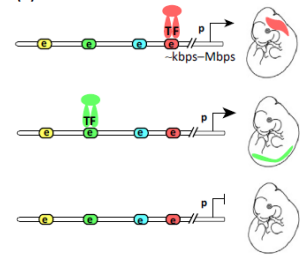


Figure 1 | The anatomies of poised and active enhancers. The characteristic features

# CHARACTERISTICS



Enhancers in tissue/cell-specific gene expression

# ENHANCER

# SELECTION

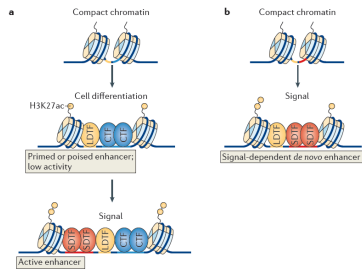
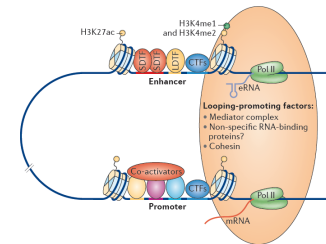


Figure 3 | Cell type-specific enhancer selection and activation. a) Collaborative

# FUNCTION

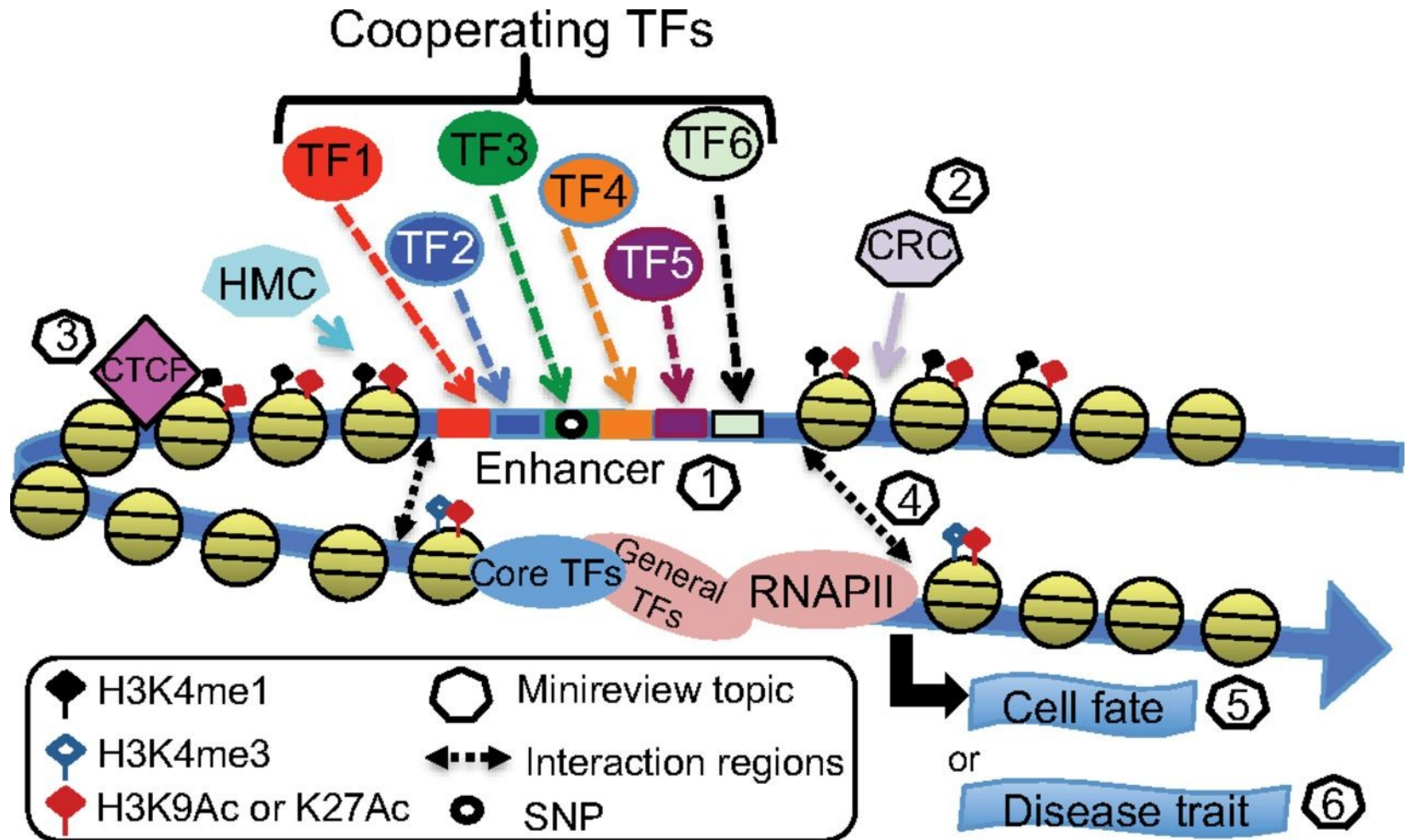




## **SNPs in the genomic regulatory regions may affect:**

- **Enhancer Activation: loss of TFs interaction or TFs recruitment.**
- **Enhancer Selection: loss or association of LTDF**
- **Alteration of timing or specific tissues activation**
- **Long range interaction between genomic regulatory regions**

## Genome-wide characterizations of regulatory regions.



Peggy J. Farnham *J. Biol. Chem.* 2012;287:30885-30887