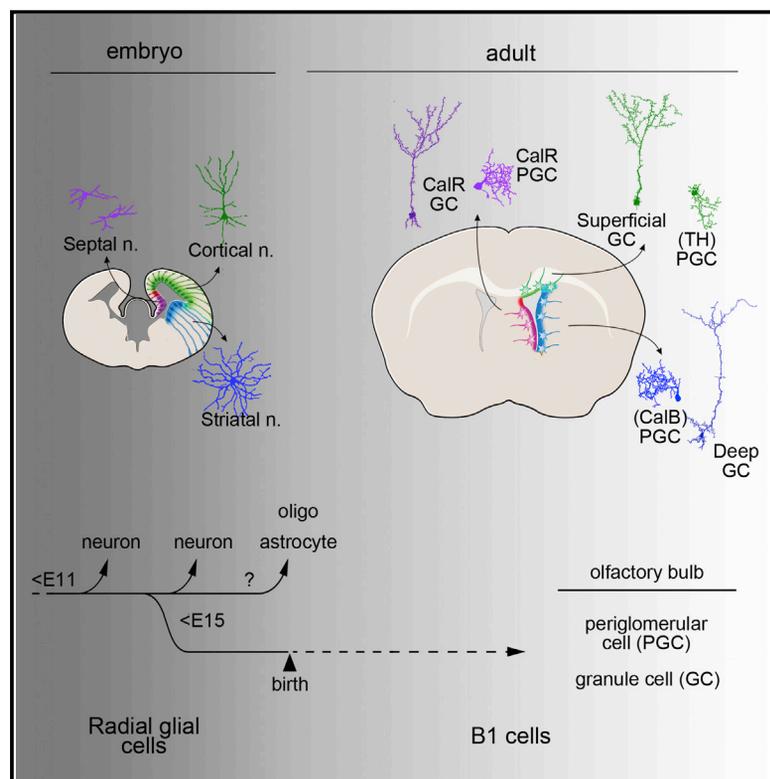


# Embryonic Origin of Postnatal Neural Stem Cells

## Graphical Abstract



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## In Brief

Postnatal neural stem cells become regionally specified early in embryonic development and remain largely quiescent until reactivation.

## Highlights

- Postnatal neural stem cells (NSCs) become regionally specified early in development
- Postnatal and fetal forebrain NSCs share common progenitors in the early embryo
- Lineages for postnatal and embryonic NSCs diverge during mid-fetal development
- After generated in the embryo, postnatal NSCs remain largely quiescent



# Embryonic Origin of Postnatal Neural Stem Cells

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## SUMMARY

Adult neural stem/progenitor (B1) cells within the walls of the lateral ventricles generate different types of neurons for the olfactory bulb (OB). The location of B1 cells determines the types of OB neurons they generate. Here we show that the majority of mouse B1 cell precursors are produced between embryonic days (E) 13.5 and 15.5 and remain largely quiescent until they become reactivated postnatally. Using a retroviral library carrying over 100,000 genetic tags, we found that B1 cells share a common progenitor with embryonic cells of the cortex, striatum, and septum, but this lineage relationship is lost before E15.5. The regional specification of B1 cells is evident as early as E11.5 and is spatially linked to the production of neurons that populate different areas of the forebrain. This study reveals an early embryonic regional specification of postnatal neural stem cells and the lineage relationship between them and embryonic progenitor cells.

## INTRODUCTION

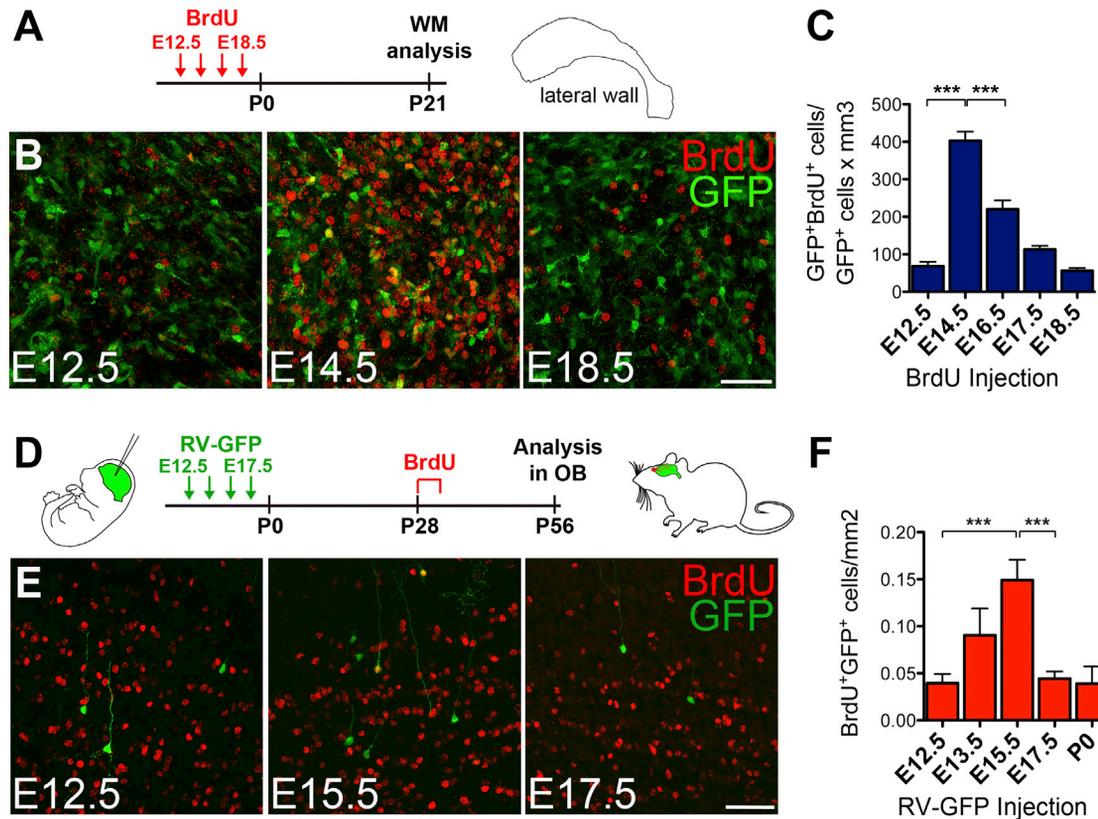
Somatic stem cells are retained throughout life in germinal niches where they maintain some of the cellular and molecular characteristics of their embryonic counterparts. Although the origin of adult stem cells is unclear, these similarities have prompted the hypothesis that postnatal somatic stem cells could correspond to embryonic progenitors that persist into postnatal and adult life (Alvarez-Buylla et al., 2001; Eckfeldt et al., 2005; Benitah and Frye, 2012; Costa et al., 2012). An understanding of the origin of adult stem cells may shed light on how they have retained or acquired their potential.

Neural stem cells (NSCs) are retained into adulthood in the ventricular-subventricular zone (V-SVZ) (Doetsch et al., 1999; Zhao et al., 2008; Ming and Song, 2011). These adult NSCs, known as B1 cells, have been best studied in rodents and lie within the walls of the lateral ventricles, next to the cortex, hippocampus, striatum, and septum (Cx, Hp, St, and Sp). B1 cells have many features of astrocytes (Doetsch et al., 1999) and retain expression of Nestin, BLBP, GLAST, and Sox2 (Lagace et al., 2007; Giachino et al., 2014), which are also expressed in radial glia cells (RGs), the NSCs in the developing brain. Indeed,

B1 cells are derived from RGs (Merkle et al., 2004) and display epithelial apico-basal organization reminiscent of RG morphology (Mirzadeh et al., 2008). These observations have suggested a linear NSC lineage from neuroepithelial cells to RGs to adult B1 cells (Alvarez-Buylla et al., 2001; Temple, 2001; Kriegstein and Alvarez-Buylla, 2009).

B1 cells give rise to neuroblasts that migrate a long distance to the olfactory bulb (OB) (Lois and Alvarez-Buylla, 1994), where they differentiate into multiple types of inhibitory interneurons (Carleton et al., 2003). Importantly, different types of OB interneurons are derived from different locations in the V-SVZ (Merkle et al., 2007; Ventura and Goldman, 2007). NSCs in the dorsal V-SVZ of the lateral wall generate mostly superficial granule cells (GCs) and dopaminergic periglomerular cells (PGCs), whereas ventral NSCs produce deep GCs and calbindin (CalB<sup>+</sup>) PGCs. In contrast, calretinin (CalR<sup>+</sup>) GCs and CalR<sup>+</sup> PGCs are derived from medial V-SVZ NSCs. The embryonic origin of this regional specification remains unknown, but it has been suggested that it is associated to the early subdivision of the embryonic forebrain into territories with the expression of a specific set of transcription factors (Alvarez-Buylla et al., 2008). The adult V-SVZ exhibits the expression of transcription factors present in different forebrain domains during development, such as Gsx1&2, Nkx6.2, Dbx1, Emx1, Pax6, SP8, and Zic1/2/3 (Hack et al., 2005; Waclaw et al., 2006; Kohwi et al., 2007; Young et al., 2007; López-Juárez et al., 2013; Merkle et al., 2014). Mice null for some of these transcription factors are deficient in the production of specific subtypes of OB interneurons in adult mice (Alvarez-Buylla et al., 2008). This raises the interesting question of whether adult B1 cells share a lineage with and inherit regional specification from RGs that earlier in development produced the different types of forebrain neurons, e.g., cortical pyramidal cells, striatal medium spiny neurons, or septal neurons.

In this study, we investigated the origin of B1 cells from dividing embryonic progenitors and their clonal relationship to neurons and glial cells in Cx, Hp, St, and Sp. Our results indicate that the embryonic progenitors of B1 cells (pre-B1 cells) were produced during mid-fetal development (E13.5–E15.5) and remained relatively quiescent until they were reactivated in postnatal life. We found that the regional specification of B1 cells took place as early as E11.5. Interestingly, some of these adult progenitors were related to RGs that generated neurons in Cx, St, and Sp, but this relationship was lost before E15.5. This work indicates that (1) adult NSCs were allocated and specified early in embryonic development, and (2) adult and embryonic NSC cell lineages diverge during mid-embryonic development.



**Figure 1. The Majority of Pre-B1 cells Are Derived at E13.5–E15.5**

(A) Timed-pregnant *hGFAP::GFP* female mice received two injections of BrdU at E12.5, E14.5, E16.5, E17.5, or E18.5, and whole-mounts of the V-SVZ of the offspring were analyzed 21 days after birth.

(B) The number of GFP<sup>+</sup>/BrdU<sup>+</sup> cells was significantly higher in mice injected at E14.5 compared to other ages. Many of the ependymal cells were also BrdU-labeled at E14.5, as previously shown (Spassky et al., 2005).

(C) Quantification of V-SVZ GFP<sup>+</sup>/BrdU<sup>+</sup> cells from animals injected at different time points. Data are expressed as mean ± SEM, n = 6/stage, three independent experiments each (\*\*\*) p < 0.0001).

(D) NIT-GFP retroviruses were intraventricularly delivered into CD1 embryos at E12.5, E13.5, E15.5, E17.5, or P0. Injected animals received BrdU for 1 week at P28, and their OBs were analyzed 3 weeks later.

(E) The majority of GFP<sup>+</sup>/BrdU<sup>+</sup> OB neurons were derived from retroviral injections at E15.5.

(F) Quantification of GFP<sup>+</sup>/BrdU<sup>+</sup> cells in the OBs of animals injected at different stages. Data are expressed as mean ± SEM, n = 12 (E12.5), n = 6 (E13.5), n = 16 (E15.5), n = 11 (E17.5), n = 4 (P0), three independent experiments each stage (\*\*\*) p < 0.0001).

Scale bars represent 50 μm in (B) and (E). See also Figure S1.

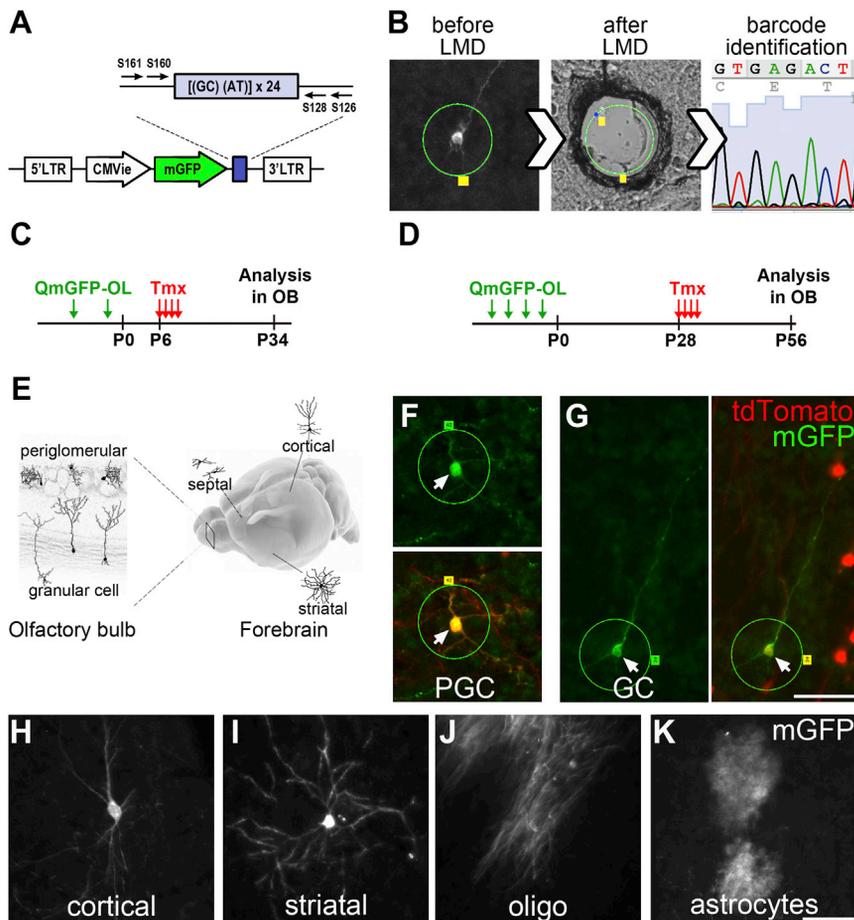
## RESULTS

### The Majority of Adult NSCs Are Generated at E13.5–E15.5

The neuroepithelial-RG-B1 cell lineage has been suggested to contain the central NSC continuum from which differentiated neurons and glia are derived (Kriegstein and Alvarez-Buylla, 2009). However, whether B1 cells are the end product of this lineage or whether they become separated from other RG during development has not been investigated. To address the contribution of dividing RGs at different stages of development to the postnatal day 21 (P21) B1 cell population, we injected timed pregnant *hGFAP::GFP* mice at different embryonic stages with the thymidine analog bromodeoxyuridine (BrdU) (Figures 1A–1C, S1A, and S1B). We found that the majority of double-labeled BrdU<sup>+</sup>/GFP<sup>+</sup> B1 cells in the P21 V-SVZ were derived from ani-

mals injected with BrdU at E14.5 and E16.5. This suggests that the majority of B1 cells were derived from RGs that divided during this period, with smaller contributions at E12.5 and E17.5–E18.5. Surprisingly, many of these B1 cells were brightly labeled, suggesting that they did not divide repeatedly in the intervening period between the time of BrdU injection in the embryo and P21.

We could not rule out the possibility that dilution of BrdU, by intervening cell cycles, could account for the lower numbers of BrdU<sup>+</sup>/GFP<sup>+</sup> cells observed at E12.5 or E17.5–E18.5. Moreover, although B1 cells can be distinguished from parenchymal astrocytes based on their morphology (Ponti et al., 2013), it is not clear whether all GFP<sup>+</sup> cells in the V-SVZ correspond to B1 cells that actually function as postnatal neuronal progenitors. We therefore used GFP-expressing retroviruses to transduce dividing embryonic VZ progenitors at different ages to permanently label their progeny in the postnatal brain (Cepko et al., 1998). We then



**Figure 2. Lineage Tracing of Individual Embryonic Progenitors with QmGFP-OL**

(A) The QmGFP-OL retroviral library: Each retrovirus expresses mGFP and contains a unique 24 bp sequence.

(B) M-GFP<sup>+</sup> cell tagged for laser microdissection (LMD, green circle), laser-cut and captured, followed by barcode identification.

(C) QmGFP-OL retroviruses were injected intraventricularly into *Nestin::CreER;Ai14* embryos (at E12.5 or E15.5); 6 days after birth, injected mice received Tmx for 4–5 days; neurons that were mGFP<sup>+</sup>/TdT<sup>+</sup> in OB and mGFP<sup>+</sup> Cx, Hp, St, and Sp cells were laser-captured at P34.

(D) QmGFP-OL retroviruses were injected into *Nestin::CreER;Ai14* embryos (at E11.5, E12.5, E13.5, or E15.5); 28 days after birth, injected mice received Tmx; mGFP<sup>+</sup>/TdT<sup>+</sup> OB neurons and mGFP<sup>+</sup> Cx, Hp, St, and Sp cells were laser-captured at P56.

(E) Schematics showing regions analyzed and examples of dissected cells in the OB and in Cx, Hp, St, and Sp.

(F and G) Examples of a mGFP<sup>+</sup>/TdT<sup>+</sup> PGC (F) and a GC (G) before LMD.

(H–K) Examples of mGFP<sup>+</sup> cells found in the telencephalon: neurons (H, cortex; I, striatum), an oligodendrocyte (J, cortex), and an astrocyte (K, cortex).

Scale bars represent 50  $\mu$ m in (G) and (K). See also Figures S2A–S2G.

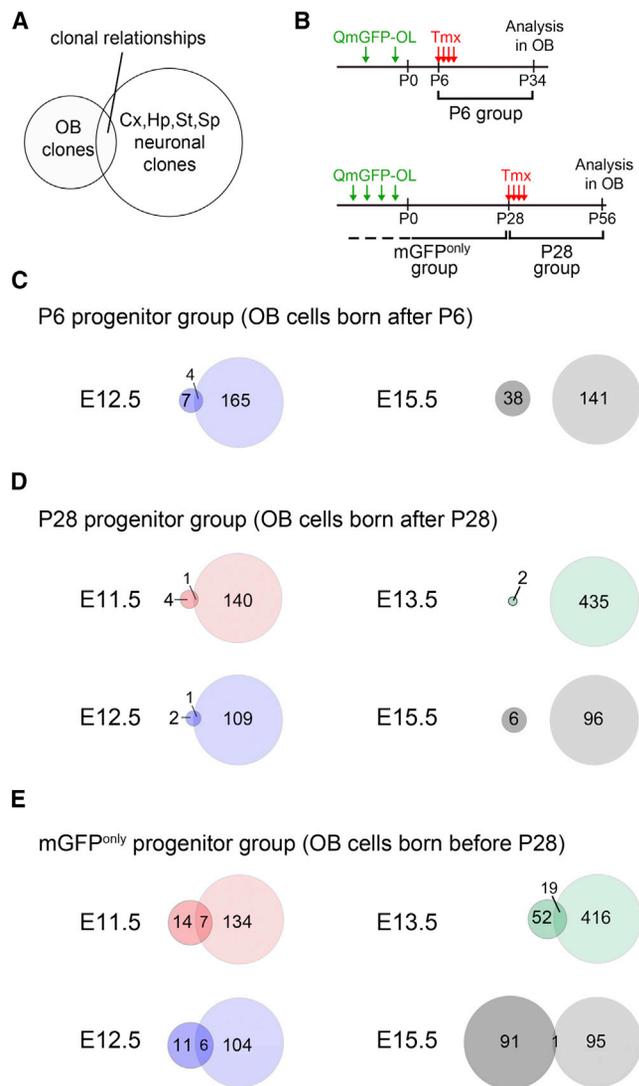
labeled a cohort of postnatal (P28) dividing V-SVZ cells with BrdU for 7 days and quantified the number of GFP<sup>+</sup>/BrdU<sup>+</sup> interneurons in the OB 3 weeks later (Figures 1D–1F). The majority of GFP<sup>+</sup>/BrdU<sup>+</sup> OB interneurons were derived from progenitor cells that divided between E13.5–E15.5. A higher production of pre-B1 cells at E14.5 compared to E11.5 was also observed in *Nestin::CreER;Ai14* mice. Injection of tamoxifen (Tmx) at E14.5 resulted in a higher number of OB interneurons generated at P28 compared to mice that receive the Tmx at E11.5 (Figures S1C–S1F).

The above experiments suggest that postnatal B1 cells are derived from embryonic cells that divide during mid-fetal development and then remain relatively quiescent until they become reactivated to generate progeny in the postnatal brain (see below). Interestingly, the period when pre-B1 cells became quiescent coincided with increased numbers of cells expressing VCAM-1, a recently described apical marker for quiescent B1 cells (Figures S1G–S1K) (Kokovay et al., 2012; Codega et al., 2014).

### Postnatal B1 Cells Are Clonally Related to Early NSCs that Generate Forebrain Cells in the Embryo

B1 cells are found in the dorsal, lateral, and medial walls of the postnatal lateral ventricles—far from the OB and next to Cx, Hp, St, and Sp. It is unclear whether adult NSCs are derived from multipotent RGs, which earlier in development generate different

subpopulations of forebrain neurons and glial cells. An alternative possibility is that a reserved subpopulation of RGs, separate from those producing neurons during embryogenesis, retain NSC properties for later use in postnatal V-SVZ neurogenesis (see Table S1 for representations of these two alternatives). We asked whether early embryonic progenitor cells, before turning into pre-B1 cells, also generate neurons and glia for Cx, Hp, St, and Sp. Because OB neurons migrate far from their progenitor cells in the walls of the lateral ventricles, a lineage-tracing method that determines clonal relationships among cells that are widely distributed is required. A barcoded retroviral library containing a degenerate 24 bp oligonucleotide with a complexity of  $\sim 10^5$  (QmGFP-OL) (Walsh and Cepko, 1992; Golden et al., 1995) was modified to include membrane-bound GFP (mGFP). The retroviral library was injected intraventricularly at different embryonic stages, and individually labeled cells were identified as neurons, astrocytes, and oligodendrocytes by their morphology, revealed by mGFP, and by the expression of NeuN, GFAP, and Olig2, respectively (Figures 2 and S2A–S2G). Their locations were mapped, and individual labeled cells were collected using laser capture. Each barcode was then identified by PCR amplification and sequencing. In order to label different cohorts of OB neurons born during postnatal development, the QmGFP-OL retroviral library was injected into *Nestin::CreER;Ai14* embryos and Tmx was administered at P6 or P28 (hereafter identified as P6 or P28 group, respectively). Four weeks later, mGFP<sup>+</sup>/TdT<sup>+</sup> OB cells and mGFP<sup>+</sup> cells in Cx, Hp, St, and



**Figure 3. Clonal Relationships between Postnatal OB Interneurons and Cx, Hp, St, and Sp Neurons**

(A) Venn diagrams throughout this figure represent clonal relationships between OB and neurons found in the Cx, Hp, St, or Sp (number of clones indicated in the center of each circle). The intersection shows the number of clones that contain both OB and Cx, Hp, St, or Sp neurons. For the total number of amplified versus microdissected cells including all OB and all Cx, Hp, St, and Sp cells, see [Table S1](#).

(B–E) Three experimental groups were analyzed: P6 ( $n = 5$ , Venn diagrams in C), P28 ( $n = 6$ , in D), and mGFP<sup>only</sup> ( $n = 6$ , in E) progenitor groups. See also [Table S2](#).

Sp were analyzed in five and six hemispheres of the P6 and P28 groups, respectively. If OB neurons share their origin with embryonically born neurons and glia in Cx, Hp, St, and Sp, they should have the same barcode, i.e., belong to the same clone.

In the P6 group, we identified 49 mGFP<sup>+</sup>/TdT<sup>+</sup> clones in the OB and 694 mGFP<sup>+</sup> clones of neurons and glia in the rest of the forebrain (from 335 mGFP<sup>+</sup>/TdT<sup>+</sup> cells in the OB and 2,637 mGFP<sup>+</sup> cells in Cx, Hp, St, and Sp; retroviral library injected at E12.5

[ $n = 3$ ] and E15.5 [ $n = 2$ ] ([Figures 3A–3C](#); [Table S1](#)). Surprisingly, clonal relationships between P6 OB neurons and Cx, Hp, St, or Sp neurons were only found in E12.5 injected animals (4 out of 11 clones at E12.5 versus 0 out of 38 clones at E15.5). In mice injected at E15.5, approximately 1/4 of OB clones contained only glia in Cx, Hp, St, and Sp (astrocytes and/or oligodendrocytes) ([Table S2](#)). One third of these clones contained mGFP<sup>+</sup>/TdT<sup>+</sup> oligodendrocytes, indicating that they were also produced postnatally. This is consistent with previous studies showing that, in addition to neurons, the postnatal V-SVZ also generates oligodendrocytes ([Rowitch and Kriegstein, 2010](#)). In the P28-group, we identified 14 mGFP<sup>+</sup>/TdT<sup>+</sup> OB and 1,128 mGFP<sup>+</sup> Cx, Hp, St, and Sp clones (from 60 mGFP<sup>+</sup>/TdT<sup>+</sup> cells in OB and 3,683 mGFP<sup>+</sup> cells in Cx, Hp, St, and Sp; retroviral library injected at E11.5 [ $n = 1$ ], E12.5 [ $n = 1$ ], E13.5 [ $n = 2$ ], and E15.5 [ $n = 2$ ]). Out of the 14 double-labeled OB clones, two were clonally related to mGFP<sup>+</sup> Cx neurons from animals injected at E11.5 and E12.5, respectively ([Figure 3D](#); [Table S1](#)).

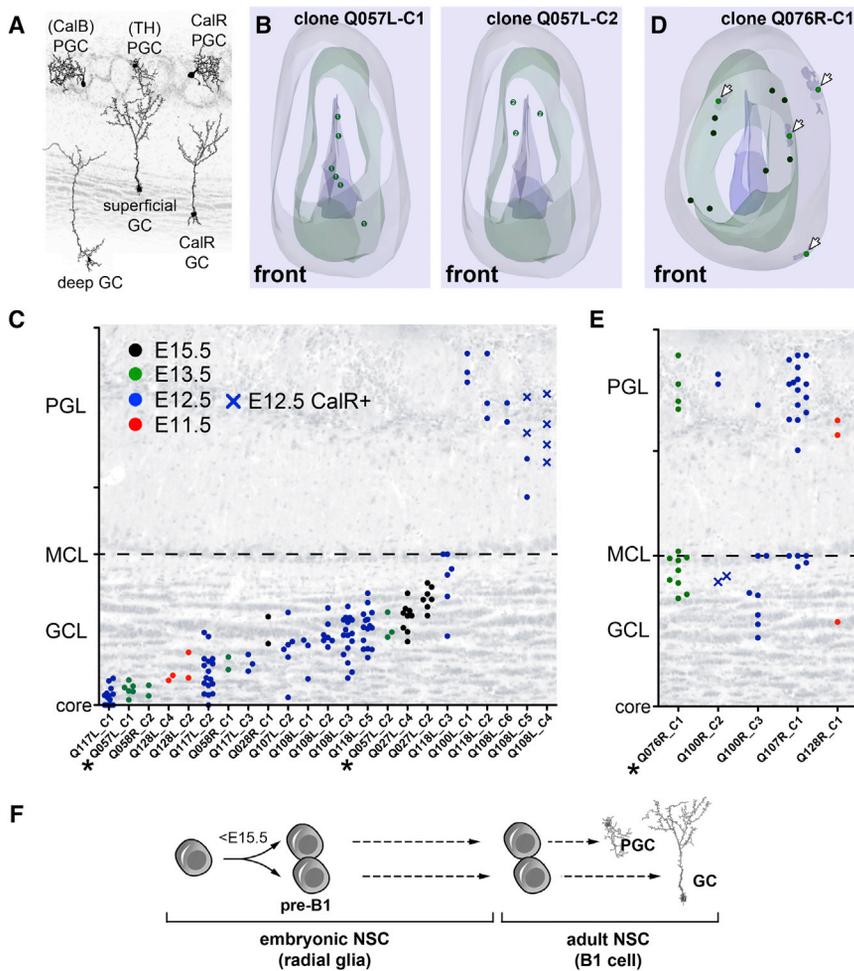
Next, we identified the barcodes of mGFP<sup>+</sup> but TdT-negative OB neurons (mGFP<sup>only</sup>) in the P28 group. This allowed us to expand our analysis to include OB neurons produced before P28, a period when many OB interneurons are born ([Bayer, 1983](#); [Batista-Brito et al., 2008](#)). Of 201 mGFP<sup>only</sup> OB clones (828 dissected cells) and 1,228 Cx, Hp, St, and Sp clones, 40 shared barcodes ([Figure 3E](#); [Table S1](#)) and were, therefore, clonally related. Of these, 33 clones had mGFP<sup>only</sup> OB cells as well as sibling neurons in Cx, Hp, St, and Sp. Interestingly, the percentage of clones containing both OB interneurons and Cx, Hp, St, Sp, neurons decreased significantly with embryonic age: from 33% at E11.5 to <2% in brains injected at E15.5 ( $p$  value < 0.0001, [Table S1](#)).

In addition to mGFP<sup>+</sup>/TdT<sup>+</sup> neurons in the OB of the P6 and P28 groups, we also found eight double-labeled cells in the subgranular zone (SGZ) of the Hp, a second germinal zone that continues receiving new neurons postnatally ([Zhao et al., 2008](#); [Ming and Song, 2011](#)). None of these postnatal SGZ clones carried the same barcode as mGFP<sup>+</sup>/TdT<sup>+</sup> OB neurons, suggesting that embryonic progenitors for postnatal neurogenesis in the OB and Hp are already separate by E11.5.

Taken together, these results demonstrate a direct clonal relationship between progenitor cells that generate OB interneurons postnatally and those that produce neurons for other regions of the telencephalon during embryonic development. The progenitor cells that produce these clones decrease rapidly between E11.5 and E15.5, suggesting that the common progenitor cells of pre-B1 cells and embryonic Cx, Hp, St, or Sp progenitors are present at early stages of development. Given that many neurons destined for Cx, Hp, St, or Sp are still produced at, and after, E15.5 ([Takahashi et al., 1999](#)), the above results suggest that pre-B1 cells become separate from forebrain RGs before this age.

### Postnatal NSCs Become Regionally Specified before E15.5

Depending on their location in the V-SVZ, B1 cells produce specific subtypes of OB neurons (see [Introduction](#)). Stereological 3D reconstructions showed that mGFP<sup>+</sup>/TdT<sup>+</sup> neurons derived from progenitor cells transduced at different stages (E11.5 to E15.5;



**Figure 4. Adult NSCs Become Specified before E15.5**

(A) Main OB interneuron subtypes and their relative layer-specific positions. PGCs are classified by the expression of CalR, TH, and CalB. GCs are classified as deep, intermediate, or superficial, plus CalR expressing.

(B) NeuroLucida 3D reconstruction of an OB (Q057L) with two P28-generated mGFP<sup>+</sup>/TdT<sup>+</sup> clones: one containing deep GCs (clone Q057L-C1 [circles in left panel]) and one containing intermediate GCs (clone Q057L-C2 [circles in right panel]). Note that clones are dispersed rostro-caudally but occupy specific positions in the GC layer (see [Movie S1](#)). The OB surface (gray), mitral cell layer (green) and the OB core (blue) are shown.

(C) The relative OB distances of individual mGFP<sup>+</sup>/TdT<sup>+</sup> cells within clones show that the majority of P28-generated clones contained either one subtype of GC or PGC (39 of 44 multi-cell [ $\geq 2$ ] clones; 23 are shown; remaining clones are plotted in [Figure 6](#)).

(D) NeuroLucida 3D reconstruction of one P28-generated clone containing both superficial GCs and PGCs (clone Q076L-C1). Arrows denote PGCs. (E) Relative OB distances of individual cells in mixed GC/PGC clones generated at P28 (5 of 44 multi-cell [ $\geq 2$ ] clones); these clones contained one GC subtype (superficial or intermediate) and PGCs. Clones containing deep and superficial GCs were not encountered in any of our embryonic injections.

(F) Pre-B1 cells are specified for the production of GC or PGC subtypes during early embryonic development before E15.5 (see text).

Closed circles and crosses denote CalR-negative and -positive cells, respectively. Asterisks mark NeuroLucida-reconstructed clones in (C) and (E). GCL: granule cell layer, MCL: mitral cell layer, PGL: periglomerular cell layer. See also [Figures S2H–S2J](#) and [Table S3](#).

P6 and P28 groups) were widely distributed throughout the rostro-caudal extent of the OB ([Movie S1](#)). However, individual clones resided in specific locations in the GC layer, from deep to superficial positions, or in the PGC layer ([Figures 4A–4C](#), [S2H](#), and [S2I](#)). The majority of mGFP<sup>+</sup>/TdT<sup>+</sup> multi-cell ( $\geq 2$  cells) clones (89%) were restricted to deep, intermediate, or superficial locations in the GC layer or were exclusively composed of PGC neurons. Seven clones contained CalR<sup>+</sup> cells, suggesting an origin in the medial wall.

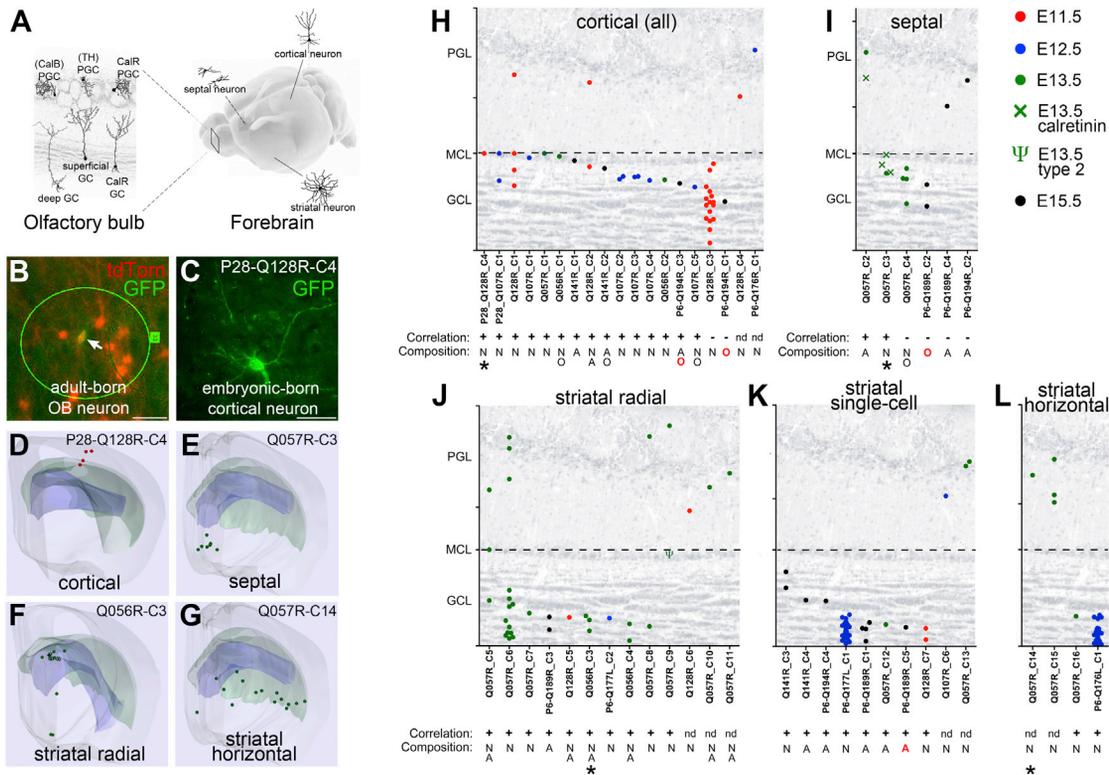
We also observed that 7 (11%) of 66 mGFP<sup>+</sup>/TdT<sup>+</sup> multi-cell clones contained a mixture of GCs and PGCs (from both the P6 and P28 groups) ([Figures 4D](#) and [S2J](#)). These mixed GC/PGC clones were rare, but more frequent among the brains that received the retroviral library at early embryonic stages ([Table S3](#)). To extend our analysis of mixed GC/PGC clones to a larger number of cells, we also studied all P28-mGFP<sup>only</sup> OB neurons (i.e., derived before P28). Of 116 mGFP<sup>only</sup> multi-cell clones, 23 were mixed GC/PGC clones (20%) ([Table S4](#)); these mixed GC/PGC clones were significantly more common at earlier injection times and became rare by E15.5 (E11.5: 36%; E12.5: 50%; E13.5: 24%; E15.5: 3.6% [p value < 0.0001]). Interestingly, these mixed GC/PGC clones showed evidence of GC specification, as

they never contained both deep and superficial GCs ([Figures 4E](#) and [S2J](#)). These observations suggest that the majority of juvenile and adult B1 cells became restricted for the production of specific OB interneuron subtypes very early in embryonic development ([Figure 4F](#)).

**Postnatal NSCs Retain the Positional Information of Their Embryonic Predecessors**

If postnatal B1 cells inherit their positional information from their embryonic progenitor cells, it should be reflected in the positions of clonally related Cx, St, and Sp cells. We mapped the locations of all cells within clones that contained both mGFP<sup>+</sup>/TdT<sup>+</sup> and mGFP<sup>only</sup> OB interneurons and mGFP<sup>+</sup> Cx, St, and Sp neurons and glia. We grouped each OB clone according to the location of clonally related cells in Cx, St, and Sp ([Figures 5A–5G](#) and [S3](#)), which likely reflects the position of the progenitor cell at the time of transduction. A remarkable relationship emerged: 15 of 17 clones in the cortex (which contained radially oriented pyramidal neurons and/or glia [[Figure 5H](#); [Movie S2](#)]) were clonally related to superficial GCs, with some containing PGCs.

In the striatum, we were able to distinguish radially and horizontally distributed clones ([Figures 5F](#), [5G](#), and [S3](#)) ([Halliday](#)

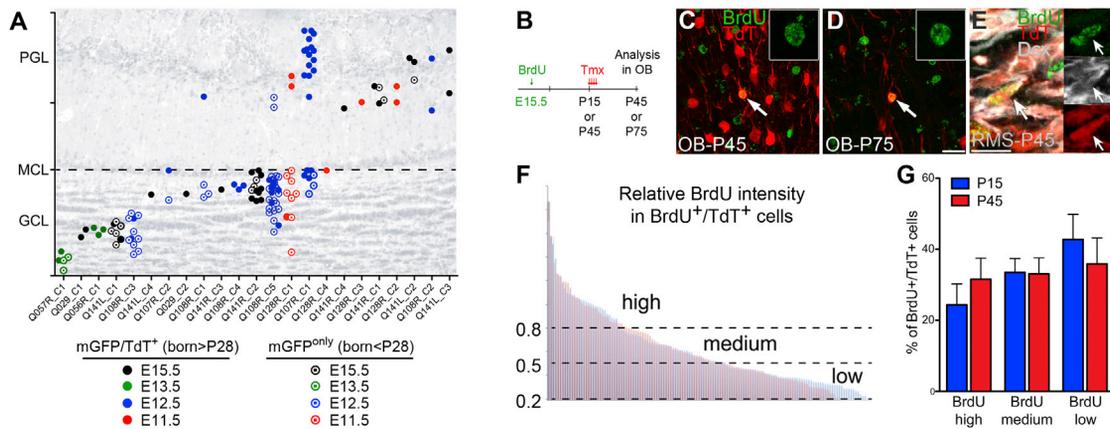


**Figure 5. Different Types of OB Interneurons Are Lineage Related to Specific Subtypes of Forebrain Neurons in a Region-Specific Manner**

(A) We investigated the location and types of cells in Cx, St, and Sp that were clonally related to the different subtypes of OB neurons. (B and C) Example of a clone (P28-Q128R-C4) containing a mGFP<sup>+</sup>/TdT<sup>+</sup> superficial GC (arrow in B) and a GFP<sup>+</sup> neuron found in cortex (C). (D–G) NeuroLucida 3D reconstructions of a radial clone in cortex (red symbols from E11.5 retroviral injection; P28-Q128R-C4 in D), septum (green symbols from E13.5 retroviral injection; Q057-C3 in E), and striatum (green symbols from E13.5 retroviral injection; F and G). Striatal clones can be further distinguished as radial (Q056R-C3 in F) and horizontal (Q057R-C14 in G). Neurons (•), astrocytes (\*), and ependymal cells (◆) are shown. The forebrain surface (gray), lateral ventricle (blue), and corpus callosum (green) are shown. (H–L) Relative OB positions of individual cells in the OB were grouped and classified according to their clonal relationships to cortical (H), septal (I), or radial (J) single-cell (K) or horizontal (L) striatal clones. Closed circles and crosses denote CalR-negative and -positive cells, respectively. Ψ denotes a type 2 cell in Q057R-C9 (in J). The expected correlation based on the previous mapping of the postnatal V-SVZ (Merkle et al., 2007, 2014) is indicated by a + or –. “n.d.” indicates that cell subtype was “not determined” due to technical limitations (see Experimental Procedures), but these cells were CalR negative as expected. Asterisks in (H), (I), (J), and (L) mark NeuroLucida-reconstructed clones in (D), (E), (F), and (G), respectively. For additional examples, see Figure S3. Scale bars represent 25 and 50 μm in (B) and (C), respectively. See also Figure S4.

and Cepko, 1992; Reid and Walsh, 2002). Horizontal striatal clones had cells that were widespread rostro-caudally (Movie S3). Half of these striatal horizontal clones contained deep GCs (Figure 5L); the others were related to PGCs in the OB. Radial striatal clones were confined rostro-caudally (Movie S4) and had a high proportion of astrocytes, which are allocated to radial domains (Tsai et al., 2012). OB cells that were clonally related to these radial striatal clones were found in deep and intermediate GC layers (9 of 10 GC-containing clones [Figure 5J]). The other striatal clone was related to a type 2 OB interneuron, a recently identified neuronal subtype that is derived from B1 cells in the lateral side of the very anterior-ventral V-SVZ (Q057R\_c9, Ψ in Figure 5J) (Merkle et al., 2014). Clones containing a single striatal cell, which could not be classified as radial or horizontal, also had siblings with deep or intermediate GCs (Figure 5K). Finally, CalR<sup>+</sup> OB interneurons were only found in clones containing septal cells (Movie S5) consistent with their production in the medial V-SVZ (Figure 5I) (Merkle et al., 2007).

The above results indicate a remarkable level of regional specification of B1 cells as early as E11.5. We were curious as to whether specification can occur even earlier; however, intraventricular injections of the retroviral library before E11.5 are technically challenging. In order to confirm the early regional specification of the NSC lineage, we selectively labeled E9.5 cortical and dorsal septal progenitor cells with a single dose of Tmx in *Emx1::CreER;Ai14* embryos. At P28, a cohort of dividing progenitor cells was labeled by a 7 day BrdU treatment, and the OB was analyzed 3 weeks later. In agreement with our clonal findings, the vast majority of TdT<sup>+</sup>/BrdU<sup>+</sup> cells were found in the superficial GC layer (Figure S4). In addition, a significant fraction of TdT<sup>+</sup>/BrdU<sup>+</sup> OB interneurons were TH<sup>+</sup> and CalR<sup>+</sup>, consistent with *Emx1* expression in cortical and septal progenitors, respectively (Kohwi et al., 2007, Young et al., 2007). P28 Tmx treatment to label adult *Emx1*<sup>+</sup> progenitors also resulted in the generation of superficial GCs and TH<sup>+</sup> PGCs, indicating that *Emx1*



**Figure 6. Pre-B1 Cells Remain Quiescent until Their Reactivation in the Postnatal Brain**

(A) Relative OB positions of mGFP<sup>+</sup>/TdT<sup>+</sup> (filled circles, born after P28) and mGFP<sup>only</sup> (open circles, born before P28) cells within clones (experiment in Figures 3D and 3E). Eleven out of 23 mGFP<sup>+</sup>/TdT<sup>+</sup> OB clones were clonally related to mGFP<sup>only</sup> OB neurons, and of these, 4 were mixed, suggesting an origin in the embryo (see also Table S4).

(B) E15.5 pregnant *Nestin::CreER;Ai14* females were BrdU injected; littermates received Tmx at either P15 or P45, and their OBs were analyzed 28 days later. (C–E) Independent of the time of Tmx administration, brightly labeled BrdU<sup>+</sup> nuclei were detected in TdT<sup>+</sup> OB neurons (C and D) and TdT<sup>+</sup>/DCX<sup>+</sup> neuroblasts in the RMS (E), indicating limited BrdU dilution.

(F) Distribution of the relative BrdU intensities in BrdU<sup>+</sup>/TdT<sup>+</sup> OB neurons from both age groups (blue: Tmx P15, n = 132 cells; red: Tmx P45, n = 126 cells). 7 mice/group, three independent experiments).

(G) Percentages of TdT<sup>+</sup> OB neurons with high, medium, or low BrdU intensities in P15 (blue) or P45 (red) Tmx-treated mice. Data are expressed as mean ± SEM, n = 7 mice/group, three independent experiments each (n.s.).

GCL: granule cell layer, MCL: mitral cell layer, PGL: periglomerular cell layer. See also Table S4.

expression is maintained from early embryonic stages to adulthood (Figure S4).

Taken together with the barcoding clonal analysis, these data demonstrate that positional information within individual embryonic progenitor cells, determining the types of neurons and glia produced for different forebrain regions throughout embryogenesis, is inherited by pre-B1 cells as early as E11.5 (and possibly earlier as suggested by the *Emx1* lineage experiments).

### B1 Cells Are Mostly Quiescent after Their Embryonic Generation

We next asked whether our clonal analyses of the P28 group from QmGFP-OL library injections could also provide insights about the postnatal self-renewing capabilities of B1 cells. If progenitor cells had divided repeatedly to generate different cohorts of OB neurons at different postnatal ages, we would expect that most OB clones containing mGFP<sup>+</sup>/TdT<sup>+</sup> cells should also contain mGFP<sup>only</sup> neurons produced before P28. More than half (12 of 23) of the mGFP<sup>+</sup>/TdT<sup>+</sup> OB clones did not have clonal relationship to other mGFP<sup>only</sup> OB neurons (262 mGFP<sup>only</sup> OB clones) (Table S4; Figure 6G). The remaining 11 clones (containing both mGFP<sup>+</sup>/TdT<sup>+</sup> and mGFP<sup>only</sup> neurons) may be derived from either self-renewing postnatal B1 cells (that divided before and after P28) or progenitor cells that divided symmetrically in the embryo to generate two pre-B1 sister cells: one dividing before and one dividing after P28. Of these 11 clones, 4 were mixed GC/PGC clones, suggesting labeling in the embryo of a more primitive progenitor cell that generated sister pre-B1 cells—one producing GCs and the other PGCs—which are produced before E15.5 (Figure 4 and Reid et al., 1999). In addition,

we also investigated whether OB neurons were clonally related to B1 cells in the V-SVZ. Clones containing B1 cells and OB neurons were extremely rare: of all double-labeled mGFP<sup>+</sup>/TdT<sup>+</sup> OB neurons analyzed from all of our embryonic injections, we only found one clone containing a labeled B1 cell (P6 group: 1 of 49; P28 group: 0 of 14) (Table S2). Even when taking into account all clones containing mGFP<sup>only</sup> OB neurons, only 7 (3.5%) had B1 cells in the walls of the lateral ventricles (Table S2). As indicated above, we do not know whether these correspond to sister pre-B1 cells that diverged in the embryo or to self-renewing B1 cells.

The above experiments suggest that B1 cells, after their embryonic production, do not divide repeatedly to produce multiple generations of OB neurons in postnatal life; otherwise they should dilute their BrdU labeling (Figures 1A–1C). In order to more directly test this possibility, we injected *Nestin::CreER;Ai14* timed-pregnant females with BrdU at E15.5 to label dividing pre-B1 cells. Littermates received Tmx at P15 or P45, and BrdU intensity was quantified in BrdU<sup>+</sup>/TdT<sup>+</sup> OB neurons 4 weeks later (Figures 6A–6D). If B1 cells undergo self-renewing divisions to produce multiple cohorts of neurons at different postnatal ages, neurons produced after P45 should contain lower BrdU levels compared to those produced after P15. However, we observed that the BrdU intensity in BrdU<sup>+</sup>/TdT<sup>+</sup> neurons in both groups was indistinguishable, indicating that separate sets of B1 cells became activated at different ages (Figures 6E and 6F). Some DCX<sup>+</sup> neuroblasts in the RMS were also brightly labeled with BrdU, indicating that young neurons were produced from another cohort of brightly labeled B1 cells shortly before sacrifice (P45 and P75). These experiments further suggest

that the majority of B1 cells are not dividing repeatedly during postnatal life to produce different generations of young neurons.

The above experiments strongly suggest that after their production in the embryo, pre-B1 cells remain largely quiescent for different periods of time until they become reactivated to generate separate cohorts of neurons postnatally. These observations argue against systematic *in vivo* self-renewal during postnatal life to generate neurons at different ages.

## DISCUSSION

In this study, we uncovered the lineage relationships of embryonic and postnatal NSCs. Using BrdU and *in utero* delivery of GFP-expressing retroviruses, we found that the majority of the pre-B1 cells were generated from embryonic cells that divided between E13.5 and E15.5. Our results suggest that B1 cells remain relatively quiescent until postnatal life, when they are reactivated to produce OB neurons. Clonal lineage analysis of forebrain progenitor cells demonstrates the presence of a common progenitor for neurons in Cx, Sp, or St and pre-B1 cells that generate OB neurons during postnatal life. Interestingly, the number of clones showing this relationship progressively decreased between E11.5 and E15.5. This suggests that, during this period, the embryonic and postnatal NSC lineages diverge. Clones with both OB cells and embryonically generated forebrain neurons revealed that the positional information of embryonic progenitor cells is maintained in postnatal NSCs, preserving region-specific production of different OB neuronal subtypes throughout life.

### Lineage Tracing with Retroviral Libraries

How adult stem cells are related to embryonic stem cells remains unanswered for many tissues, in part because it is technically challenging to follow the progeny of individual progenitor cells from the embryo into postnatal life. In the V-SVZ, understanding the embryonic origin of postnatal NSCs is further complicated by the long migration that separates the final progeny (interneurons in the OB) from progenitor cells within the walls of the lateral ventricles. Here we used a retroviral library carrying at least  $10^5$  tags (barcodes) (Golden et al., 1995) to infer lineage relationships. Other lineage-tracing methods, based on the use of fluorescent reporters of different colors (Livet et al., 2007; Schepers et al., 2012; García-Marqués and López-Mascaraque, 2013), do not provide the complexity in tag diversity to convincingly demonstrate that two cells in very different regions are derived from a single progenitor cell.

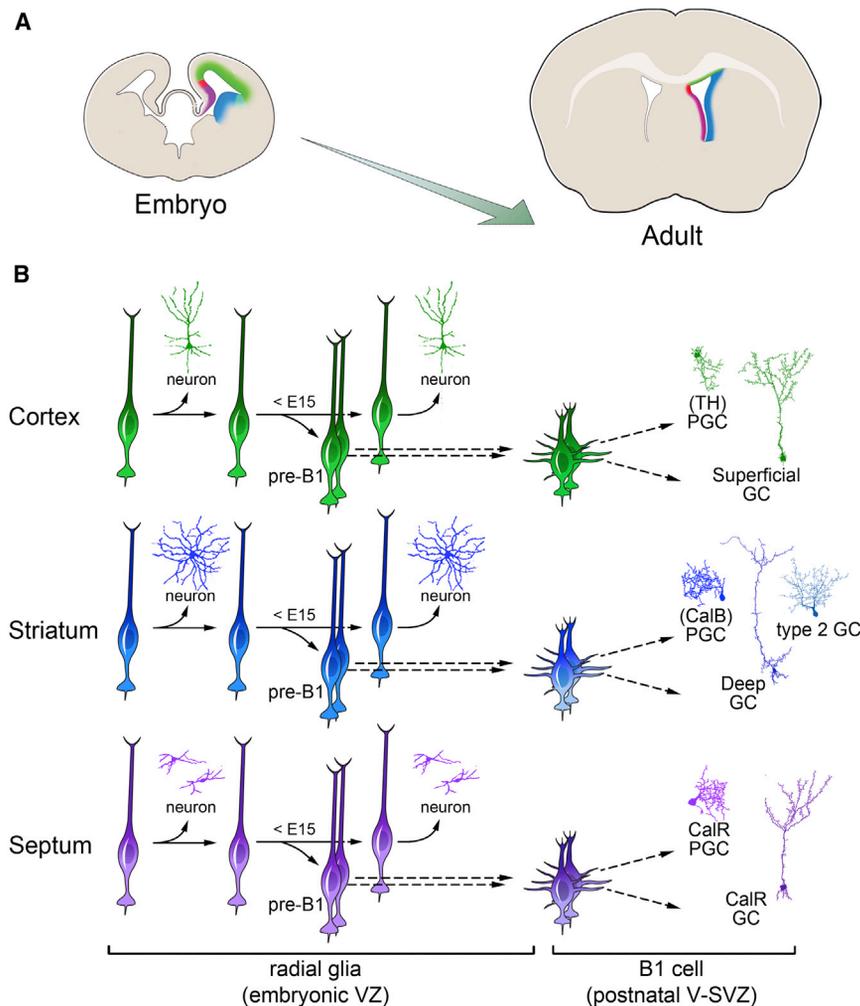
The retroviral barcode method also has limitations that could affect the interpretation of results. Viral inactivation (Walsh and Cepko, 1992; Golden et al., 1995) may result in an underestimation of clone size and/or complexity. In addition, mGFP that allows identification of transduced cells may not reach detection threshold. Because the rate of viral inactivation may vary among cell types and injection times, we assayed for silent genomes by randomly capturing pieces of tissue containing mGFP-negative cells. In the OB, where the high concentration of PGCs and GCs would make it more likely to capture silenced cells, we did find mGFP-negative microdissected cells carrying retroviral tags. This indicates that a fraction of the cells derived from

progenitor cells transduced in the embryo inactivate the reporter gene; the frequency of false negative was low (4 of 252 mGFP-negative microdissected OB tissues yielded a barcode [Table S4]). As indicated above, we could recover the barcode sequence from 51.4% of microdissected mGFP<sup>+</sup> cells, which also artificially reduces our clonal sizes. We therefore do not make inferences about clone size and limit our conclusions to the clonal relationships among cells that were positively identified.

It is also possible that two cells could be derived from independent transductions with viruses carrying the same barcode. Theoretically, given the *in vitro* tested complexity of the library ( $10^5$ ), the probability of false positives is small (Golden et al., 1995). When we compared the frequency of barcodes in six brains with a total of 1,446 different tags, we found 1 tag that was repeated in 3 brains and 48 tags that were repeated in 2 brains (Table S5). This suggests that some barcodes are overrepresented in the library, increasing the probability of “lumping” errors, i.e., the assumption that cells with the same barcode are siblings when they are actually derived from more than one progenitor cell. From the observed number of repeated tags and the number of clones per brain, less than 2.4% of the barcode tags had a probability to produce “lumping” errors (Table S5). Because our conclusions are based on the analysis of 2,186 different tags, we conclude that the vast majority of the cells that share the same barcode were derived from a single progenitor. Furthermore, the close correlation observed between OB subtypes and the location of clonally related cells in the forebrain strongly supports our conclusion that cells presumed to belong to the same clone were indeed derived from single progenitor cells in the embryo.

### Temporal Progression of Region-Specific NSC Lineages

During development, NSCs change in potential and generate different types of neurons and glial cells at different times (Livezey and Cepko, 2001; Kohwi and Doe, 2013). This process, which has been best studied in *Drosophila* and is referred to as temporal identity specification, depends on transient changes in transcription factor expression that are related to the birth order. In the mammalian brain, progenitors change over time, gradually becoming restricted in potential (McConnell, 1995). For example, a period of neurogenesis in which different types of neurons are sequentially produced is followed by a period of gliogenesis (Rowitch and Kriegstein, 2010). This raises the question of the origin of adult NSCs, a subset of postnatal progenitor cells that maintain the capacity to generate neurons throughout life. Where do these cells come from? Here we show that subpopulations of embryonic progenitor cells that produce different types of neurons and glial cells in the Cx, St, and Sp also generate pre-B1 cells. Although the embryonic progenitor cells that generate both embryonic neuronal progeny and B1 cells are clearly multipotent, they do not generate all types of forebrain cells. Instead, our findings suggest that, as early as E11.5, the neurogenic potential of embryonic progenitor cells is highly restricted by their location. This regional identity is maintained from embryonic to adult stages (Figure 7). Remarkably, this spatial identity encodes for the production of very different



**Figure 7. Embryonic Origin of Postnatal NSCs**

(A) Left: Coronal section of the embryonic (E12.5) mouse brain illustrating regional organization of cortical (green), striatal/LGE (blue), and septal (purple) VZ progenitor cells. Right: These progenitor domains are retained in the postnatal brain (dorsal [green], ventral [blue], and medial [purple]) and contain NSCs that generate specific subtypes of interneurons that migrate to the OB.

(B) Individual lineages illustrating the progression from embryonic to adult NSCs. Postnatal NSCs arise from region-specific pre-B1 cells that progressively diverge from other embryonic progenitors that produce specific sets of neurons in cortex (green), striatum (blue), and septum (purple). Later in life, these same regions harbor dorsal (green), ventral (blue), and medial (purple) V-SVZ B1 cells that produce superficial GCs, intermediate/deep GCs, and CalR<sup>+</sup> GCs and PGCs, respectively. Pre-B1 cells become restricted for the production of specific subtypes of GCs or PGCs as early as E11.5 and before E15.5.

types of neurons in the embryo and in the adult. For example, progenitor cells in the cortex, which in the embryo produce pyramidal cells (glutamatergic), but not local interneurons, also generated B1 cells that during postnatal life give rise to very specific subpopulations of superficial GC and PGC interneurons (GABAergic) in the OB.

Temporal identity specification may be orchestrated by a number of cell-intrinsic and -extrinsic factors (Livsey and Cepko, 2001; Kohwi and Doe, 2013). A recent study using time-lapse clonal analysis *in vitro* showed that individual E11.5 cortical progenitors could generate Tbr1<sup>+</sup> projection neurons followed by GAD67<sup>+</sup>/SP8<sup>+</sup> OB interneurons (Cai et al., 2013). These observations suggest that this identity switch may be controlled by intrinsic factors in cortical progenitor cells, as it has been proposed to take place during deep-to-superficial projection neuron and projection neuron-to-glia fate changes (Qian et al., 2000; Shen et al., 2006). Consistent with the concept that spatial specification can have very different outcomes depending on the timing, a recent study shows that conditional ectopic expression of the dorsal LGE transcription factor *Gsx2* results in different

embryonic progenitor cells that contribute to different areas of the forebrain.

#### Pre-B1 Cells Become Quiescent until Their Reactivation in the Postnatal Brain

Our results indicate that the majority of pre-B1 cells divide around E13.5–E15.5. Several lines of evidence suggest that B1 cells then remain relatively quiescent, until being reactivated at different ages in the postnatal brain: (1) P21 B1 cells showed little evidence of BrdU dilution from pre-B1 cells. (2) B1 cell progenies generated at different times had indistinguishable intensities of BrdU when given at E15.5. (3) VCAM1, a marker of quiescent B1 cells (Kokovay et al., 2012; Codega et al., 2014), was expressed in progenitor cells between E13.5 and P0. Finally, (4) a significant fraction (52%) of P28 clones did not contain OB cells born before P28. In these experiments, we used the *Nestin::CreER;Ai14* mouse line, which has been shown to label the majority of primary progenitors in the V-SVZ with no evidence of nonspecific labeling (Lagace et al., 2007). Clones containing both mGFP<sup>+</sup>/TdT<sup>+</sup> and mGFP<sup>only</sup> OB interneurons could result from (1) self-renewing B1 cells that generated progeny before



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