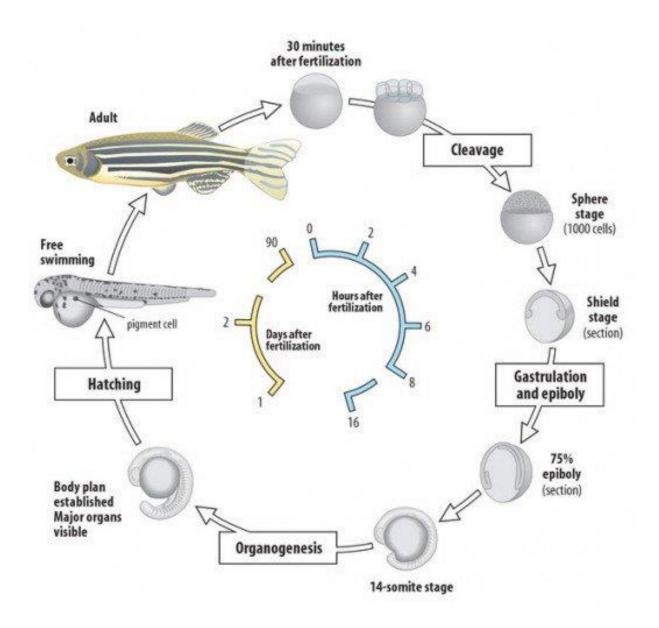
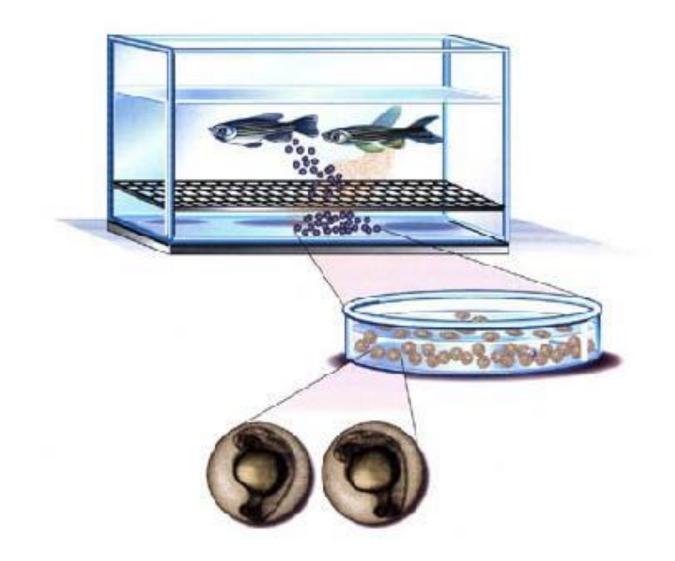
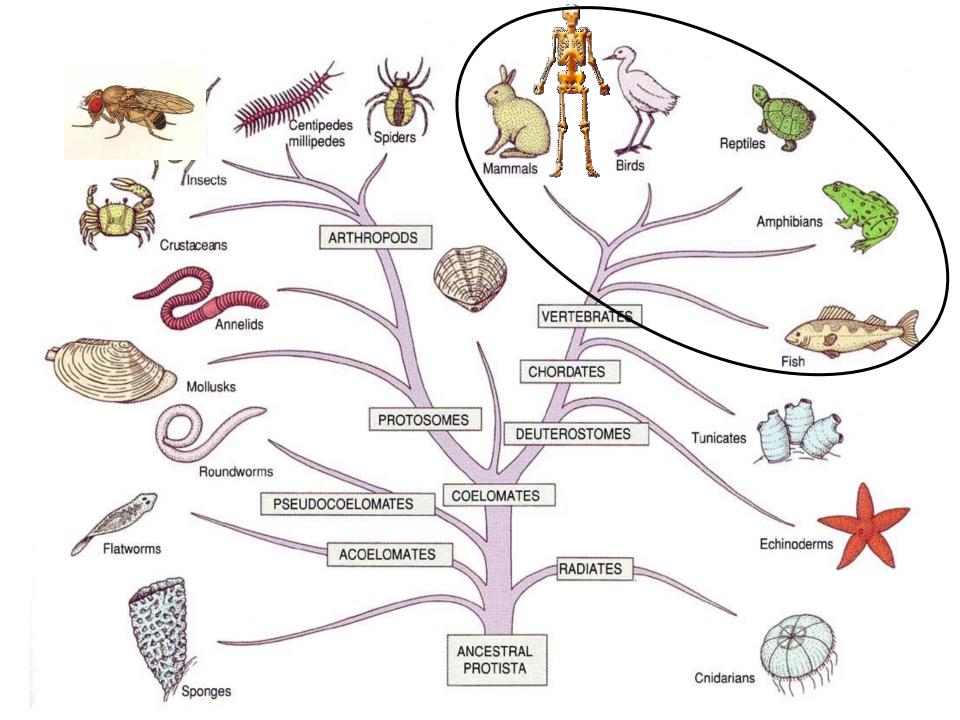
### Zebrafish, a model of choice for biomedical research

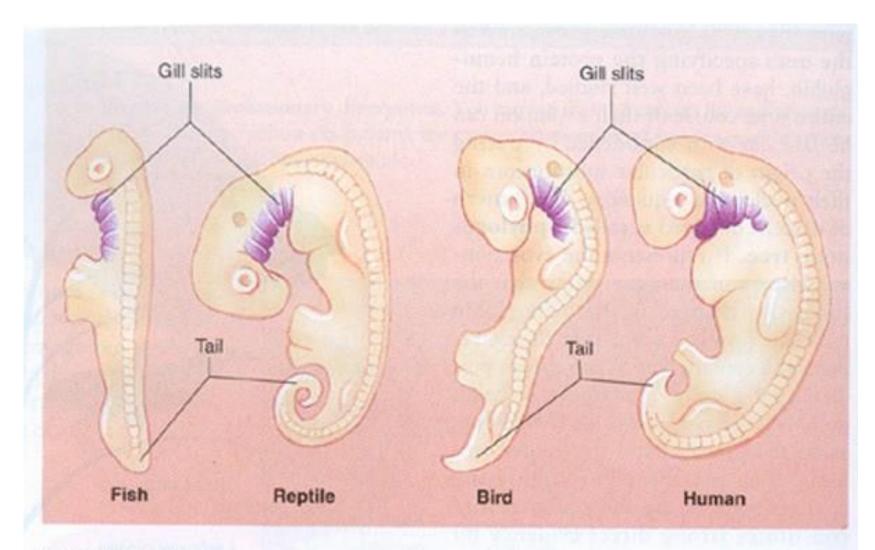
Yoav Gothilf Dept. Neurobiology, Tel Aviv University yoavgothilf@gmail.com





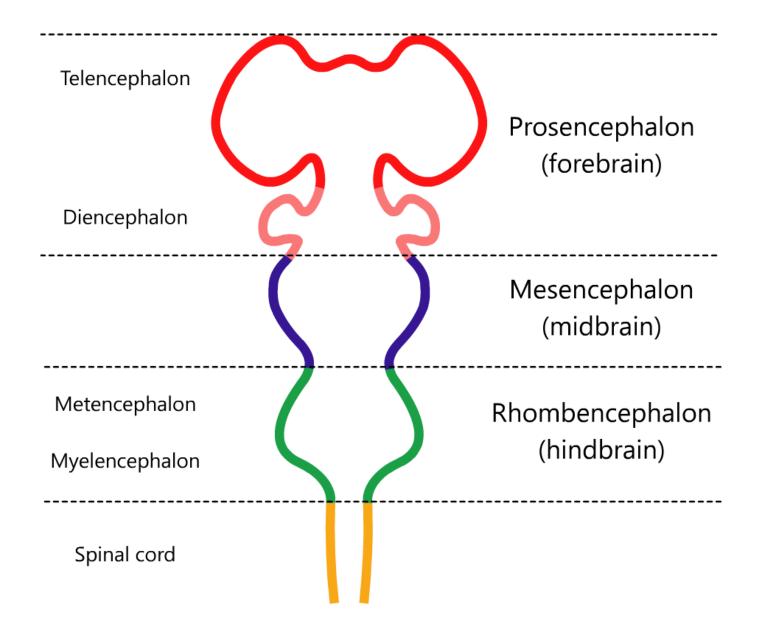
https://youtu.be/4c-Kw4timVA





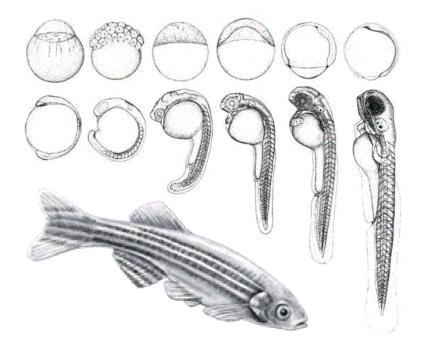
#### **FIGURE 20.18**

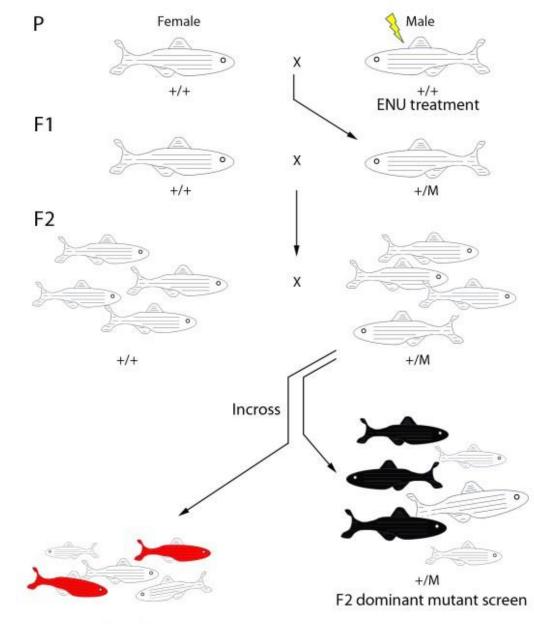
Our embryos show our evolutionary history. The embryos of various groups of vertebrate animals show the features they all share early in development, such as gill slits (*in purple*) and a tail.



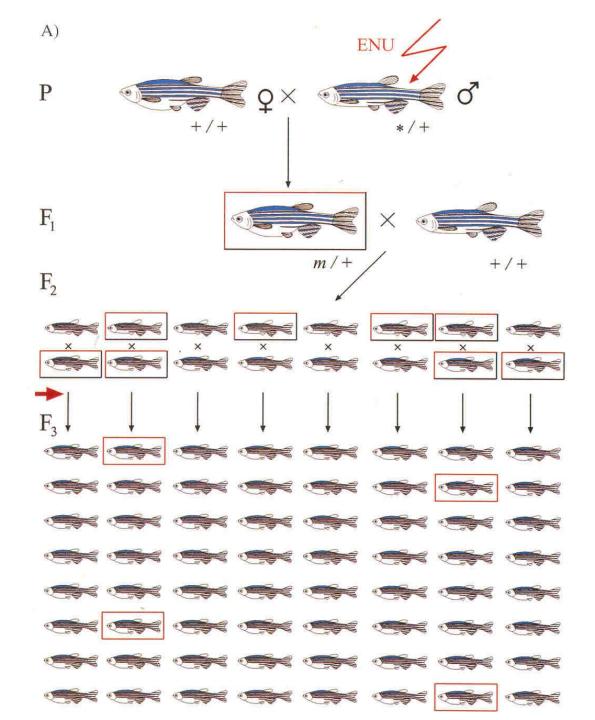
# Zebrafish

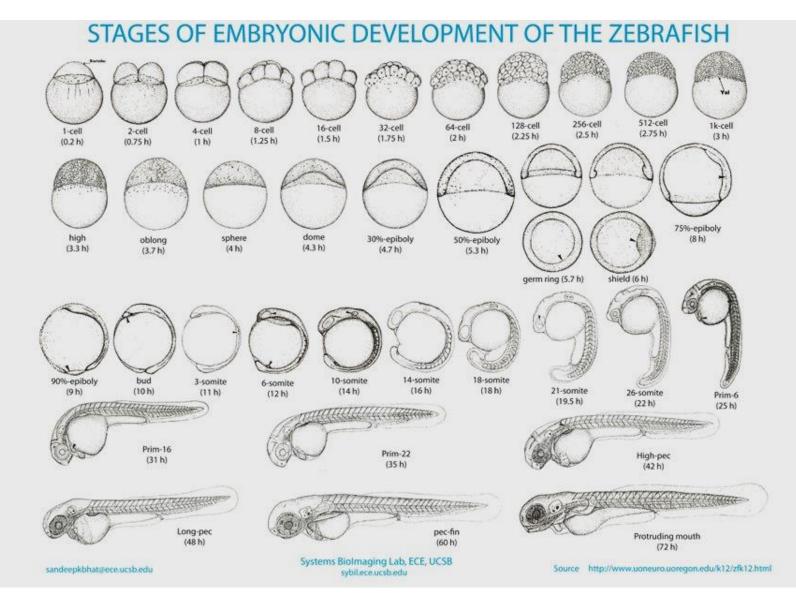
- Vertebrate
- External fertilization
- Many embryos
- Transparent embryos
- Fast development
- Short generation time
- Easy to maintain

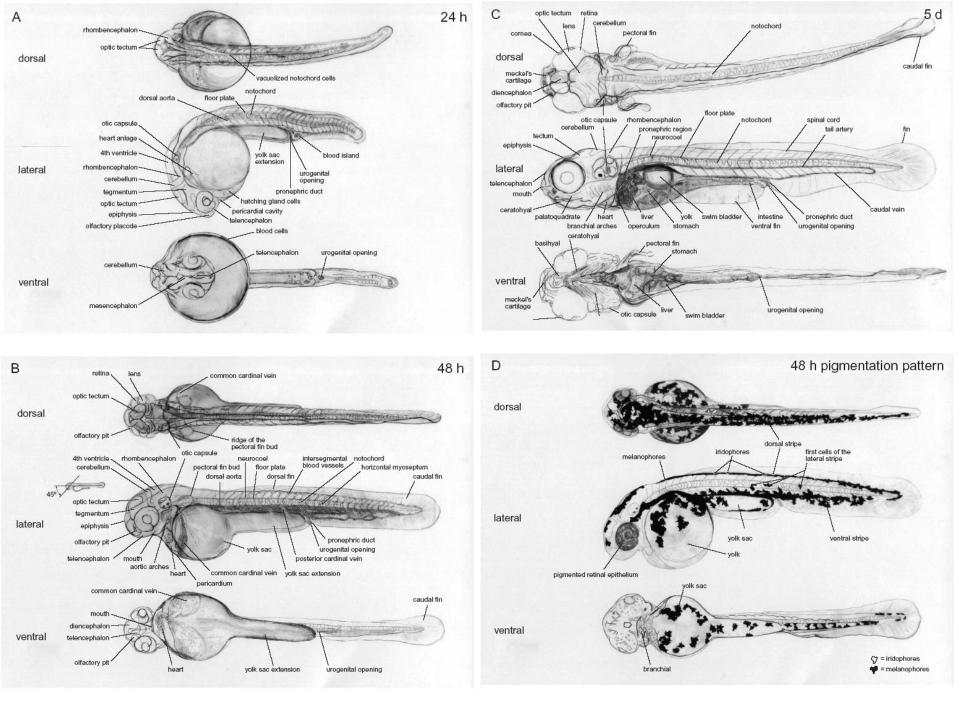


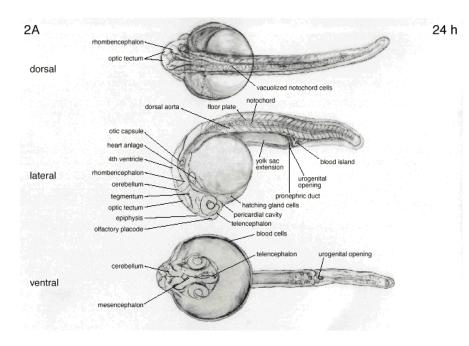


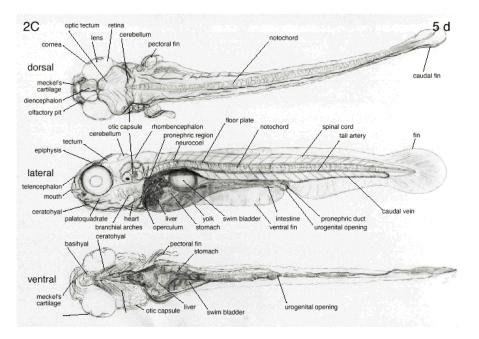
+/M x +/M F3 recessive mutant screen

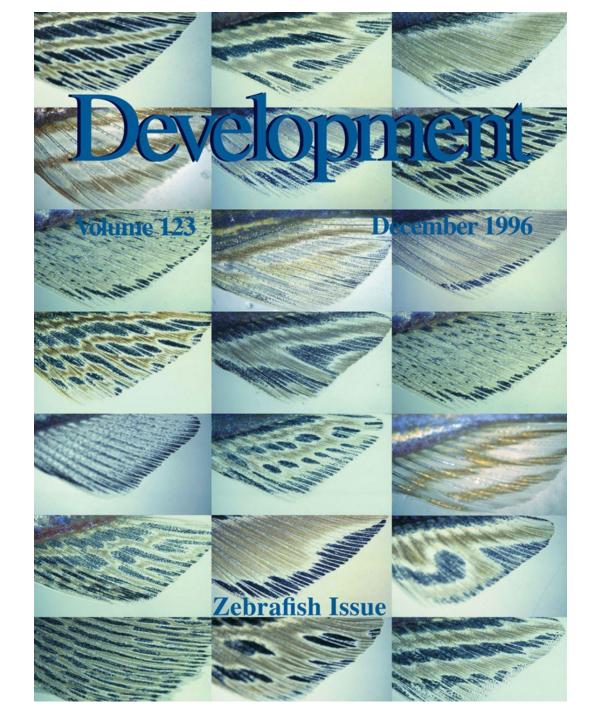






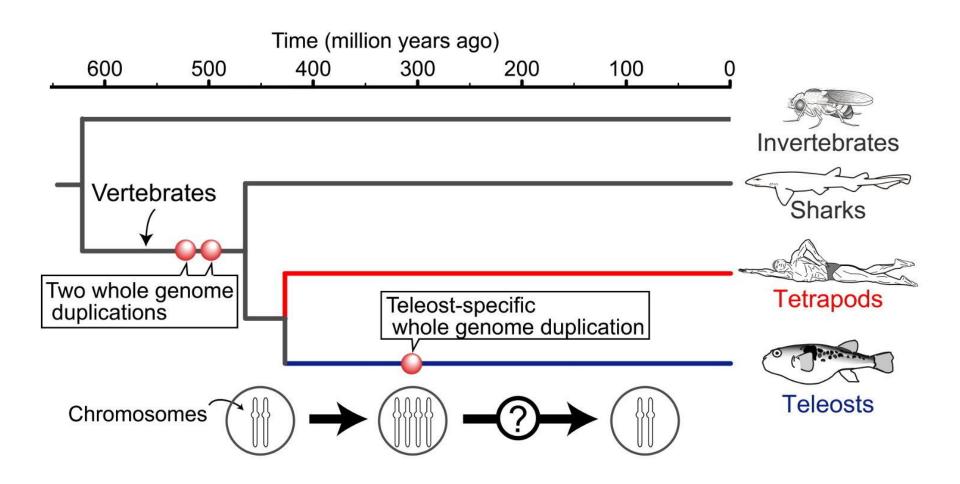




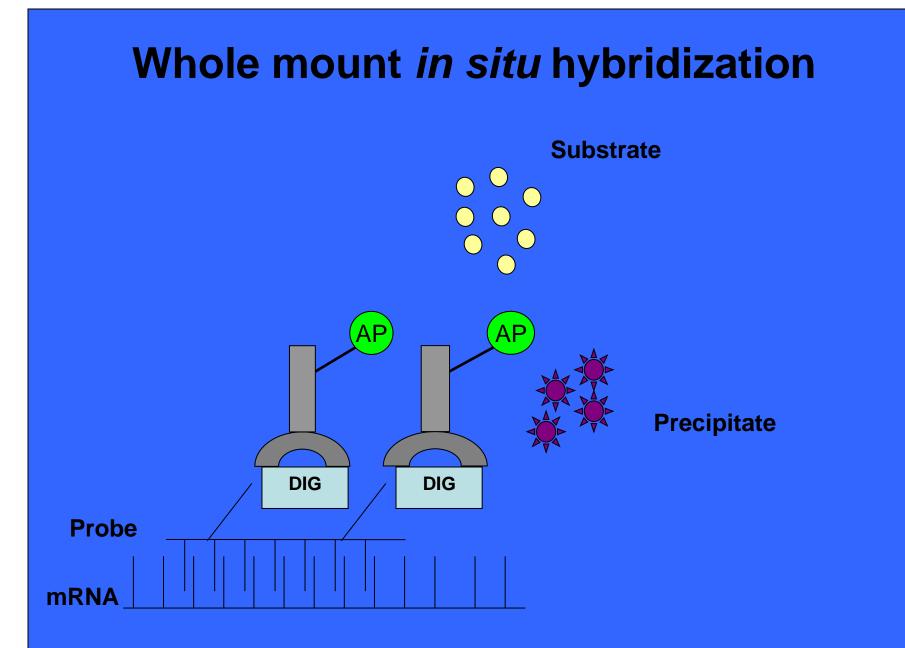


# Zebrafish, a model of Choice

- Vertebrate
- Large number of accessible transparent embryos
- Fast development and short generation time
- Developmental mutants
- Gene expression analysis
- Transgenesis
- Gene knockdown, knockout and knockin
- Behavioral tests
- Imaging capabilities



Whole genome duplication is a rare evolutionary event that has played a dramatic role in diversification



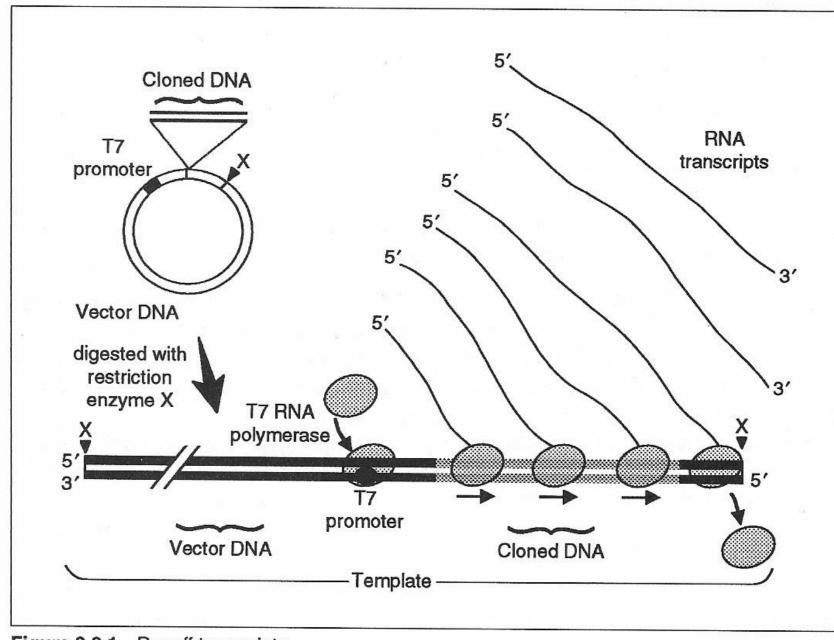
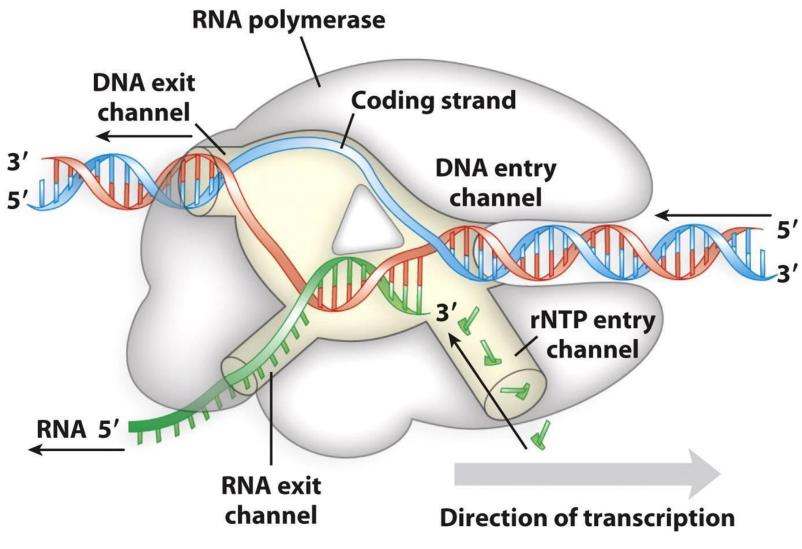
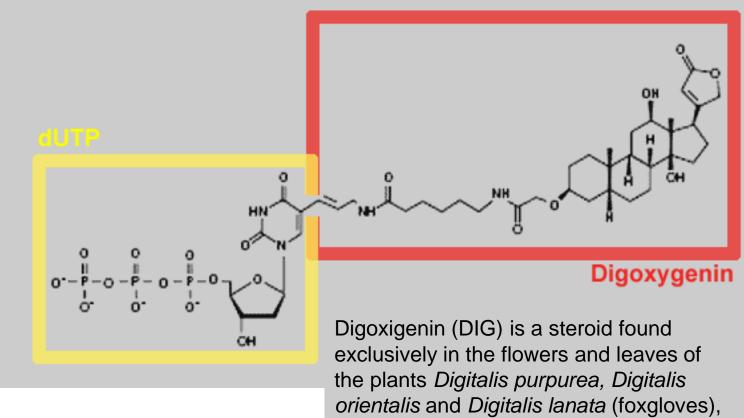


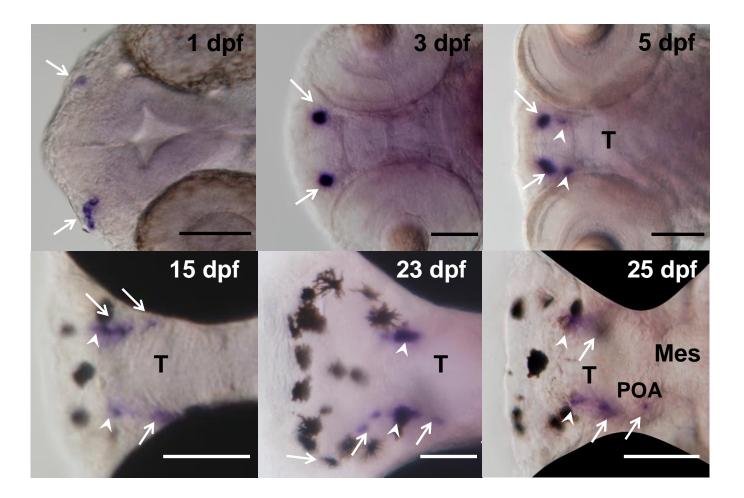
Figure 3.8.1 Runoff transcripts.





where it is attached to sugars

#### **GnRH3 mRNA localization during zebrafish development**

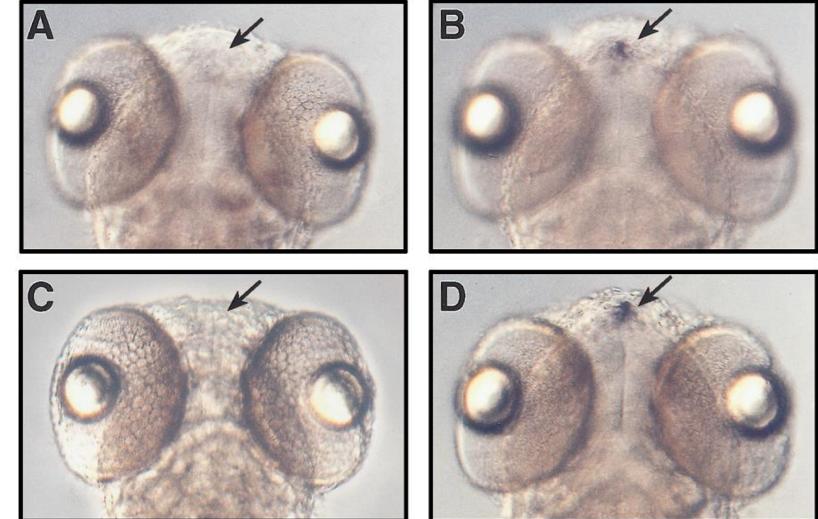


Ori Palevitch 2005

#### AANAT2 mRNA expression in zebrafish larvae

ZT6

**ZT18** 

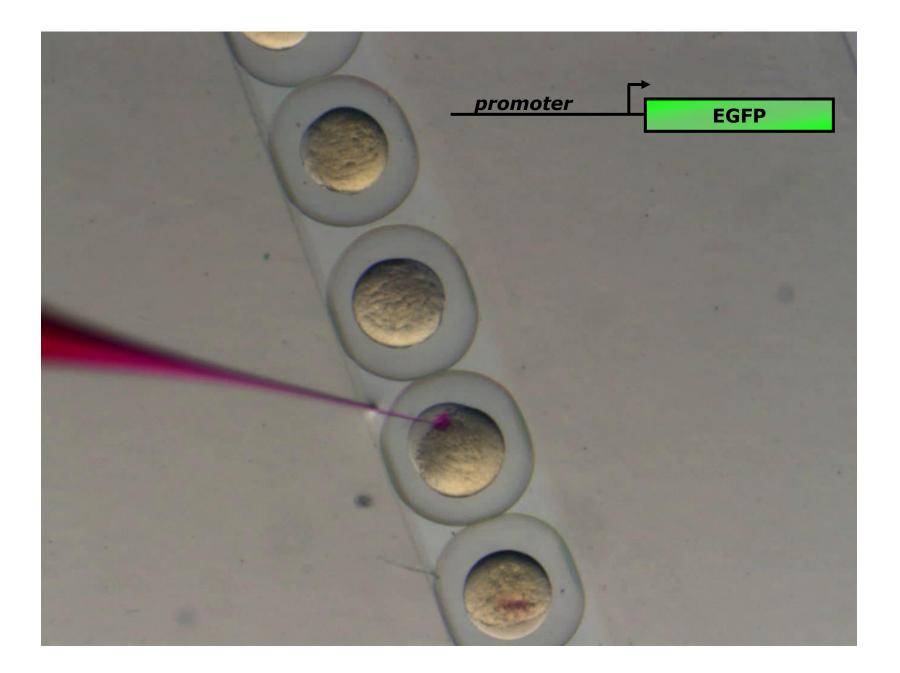


LD

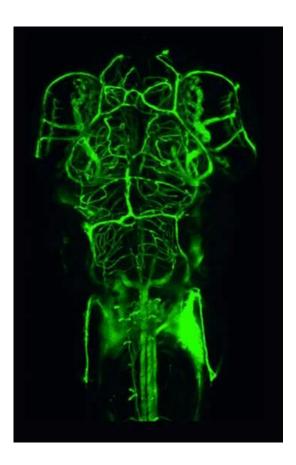
DD

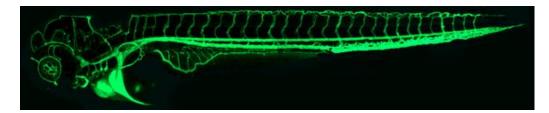
Gothilf et al., 1999

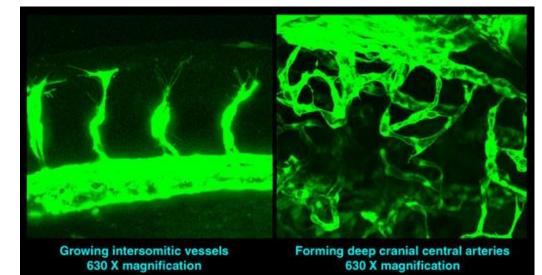
Transgenesis: Method development and utilization for visualizing and manipulating neuros



### Transgenic fish in which GFP is driven by vascular-specific promoters

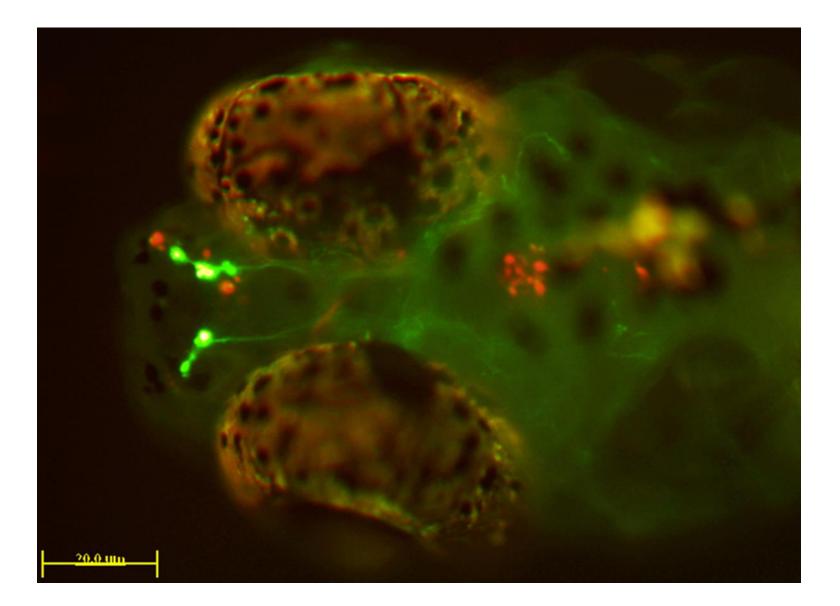


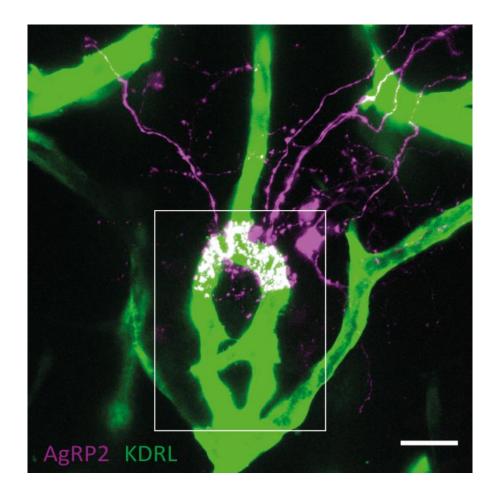




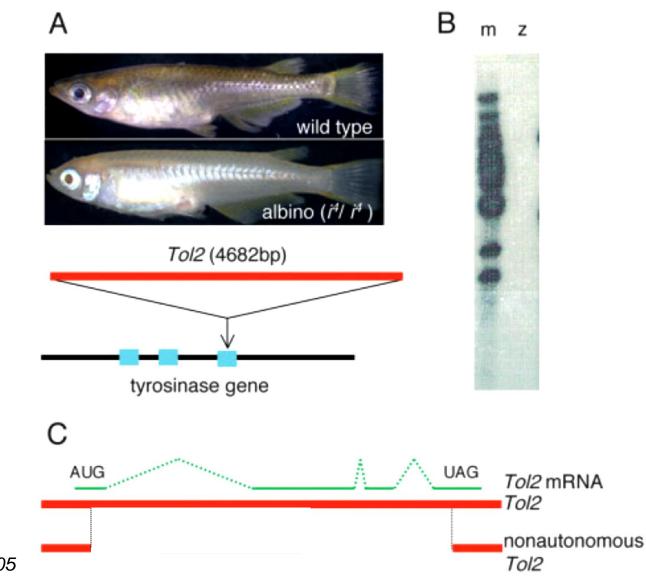
https://youtu.be/yk7TWOtrphM



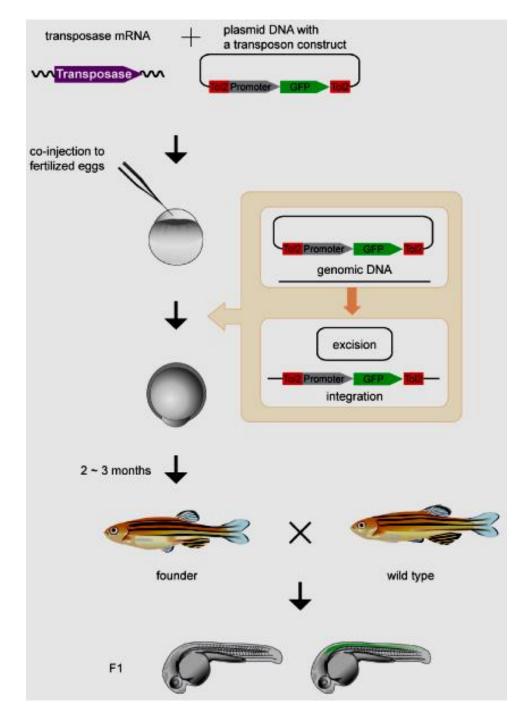




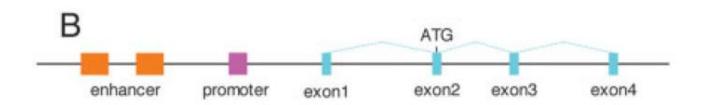
# Discovering the Tol2 transposase system



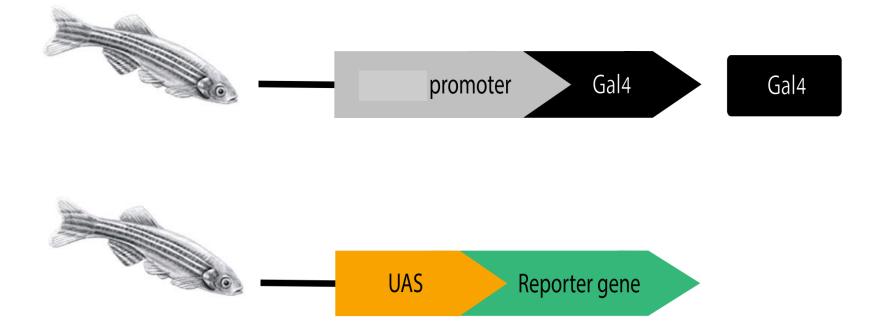
Kawakami 2005



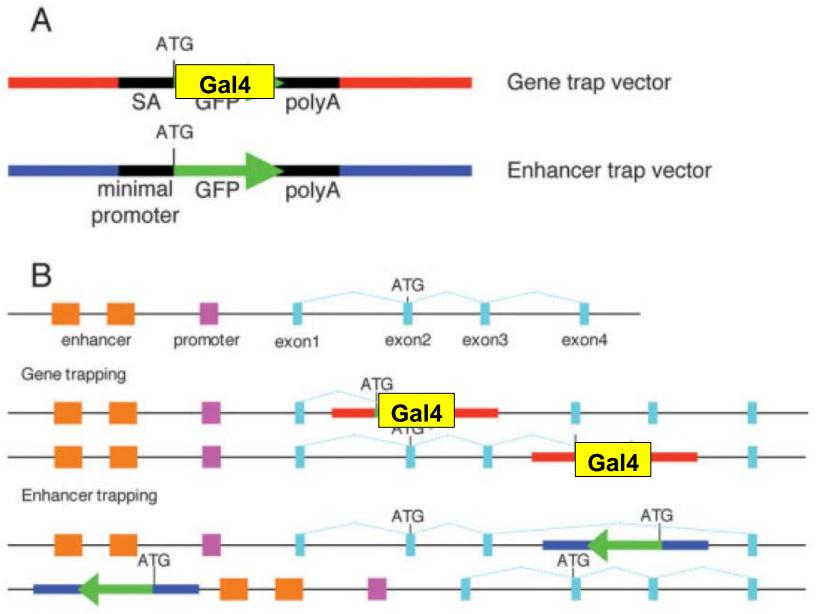
# Gene and enhancer trap using the Tol2 system



### The GAL4-UAS system

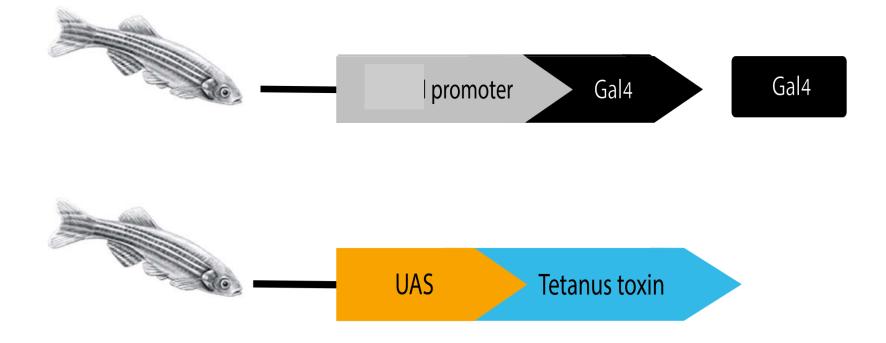


# Gene and enhancer trap using the Tol2 system

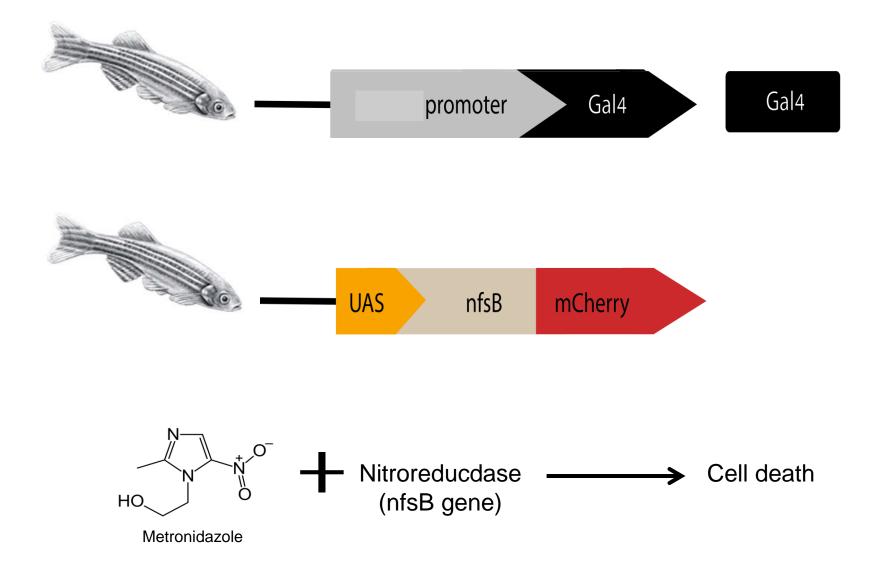


Kawakami 2005

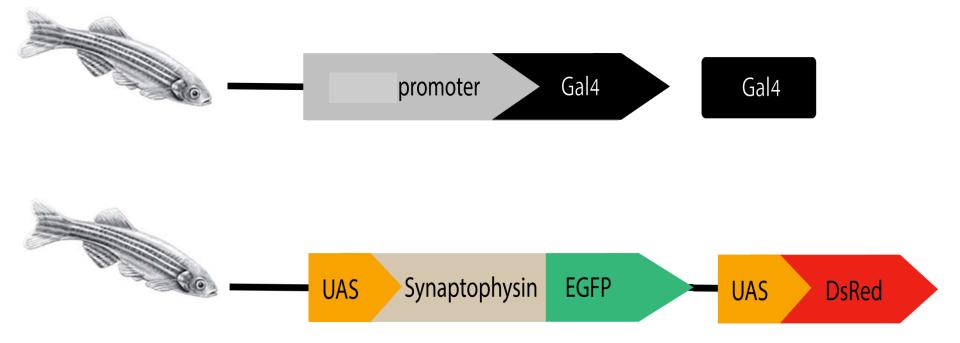
## Specific neuronal inhibition

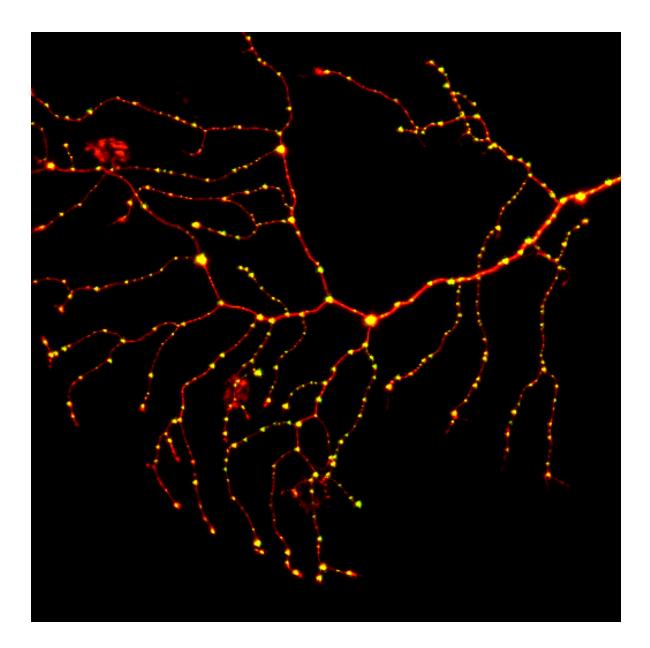


### **Neuronal ablation**

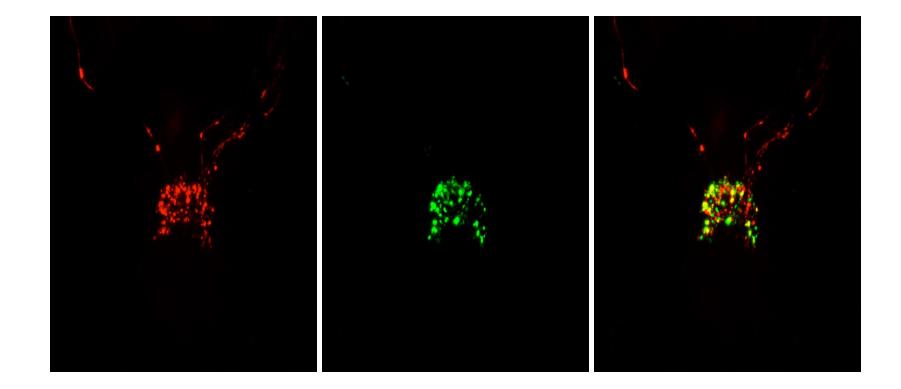


## Synaptic plasticity



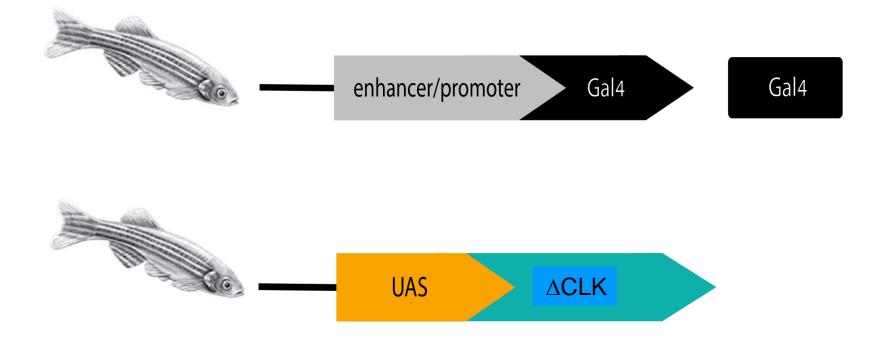


Lior Appelbaum, BIU

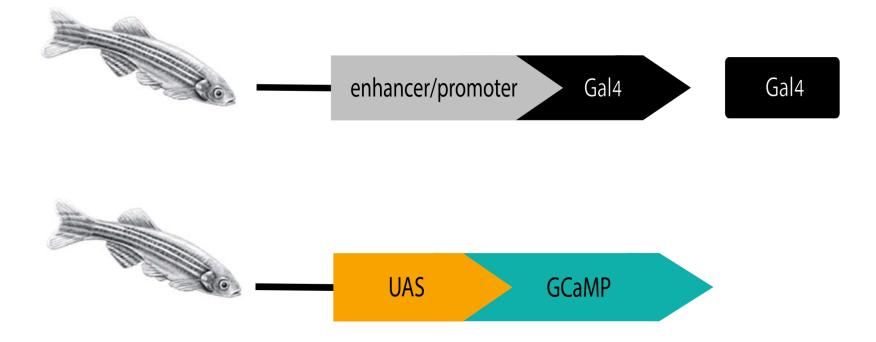


Shainer et al., 2017

#### Blocking the clock

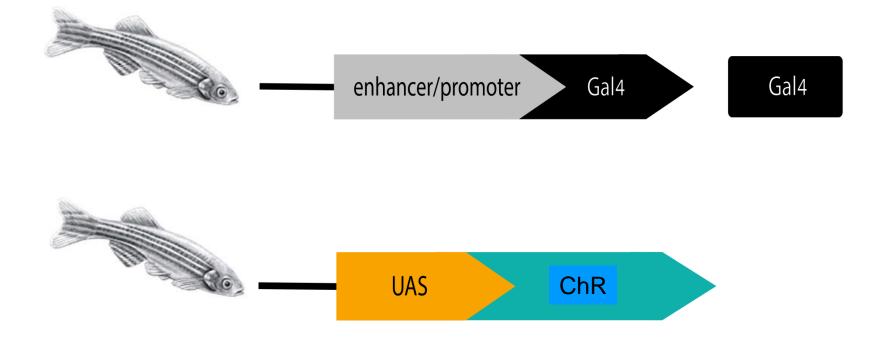


#### Seeing neurons in action, using Ca<sup>++</sup> sensor

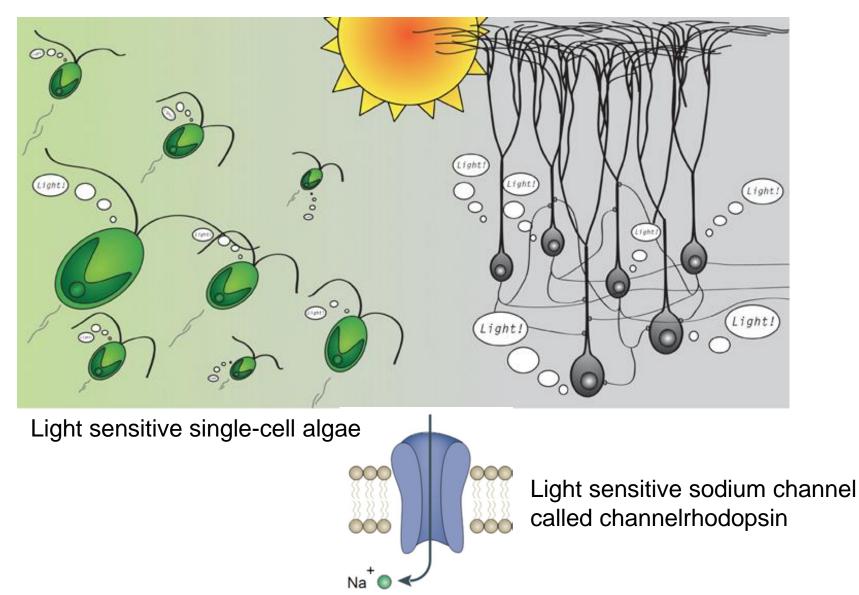


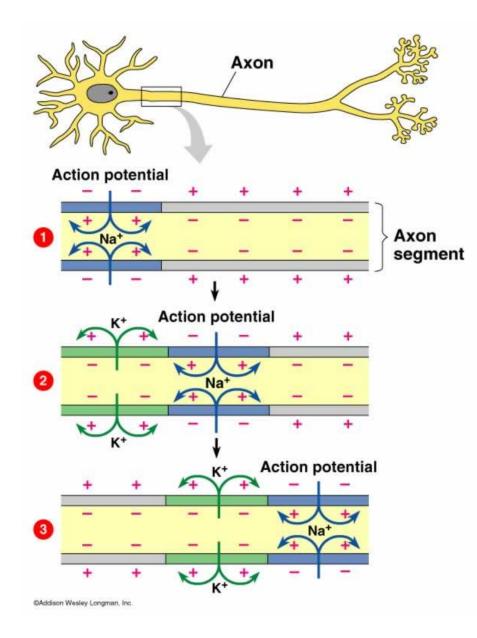
#### https://www.youtube.com/watch?v=1Q-g1uCvYOA

#### Activating neurons

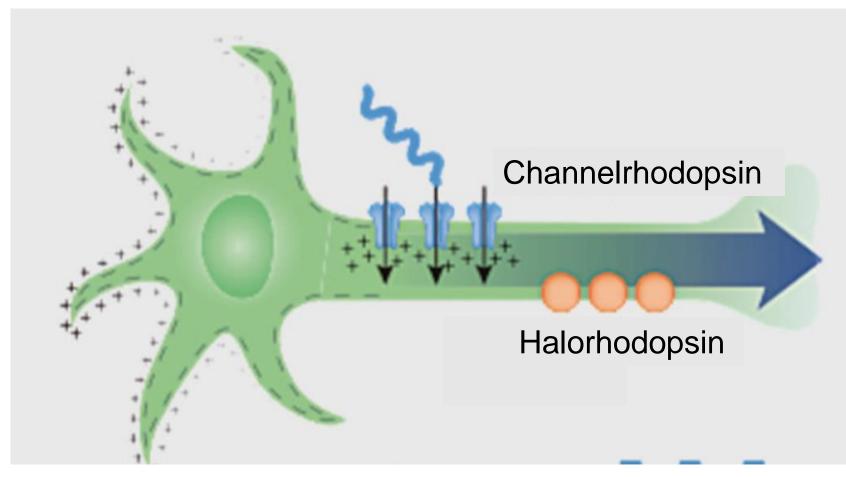


## **Optogenetics**

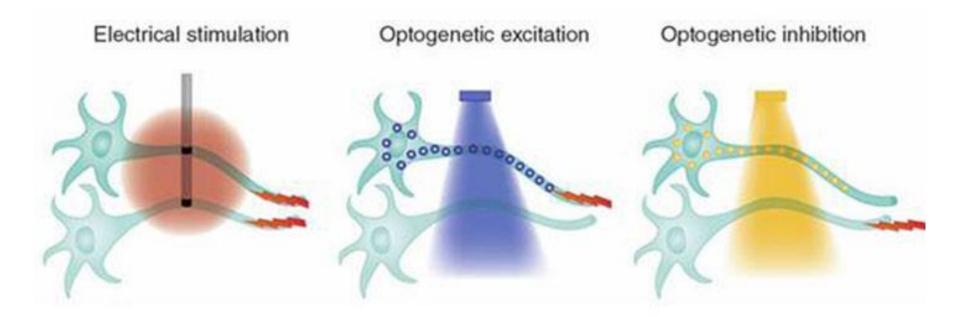


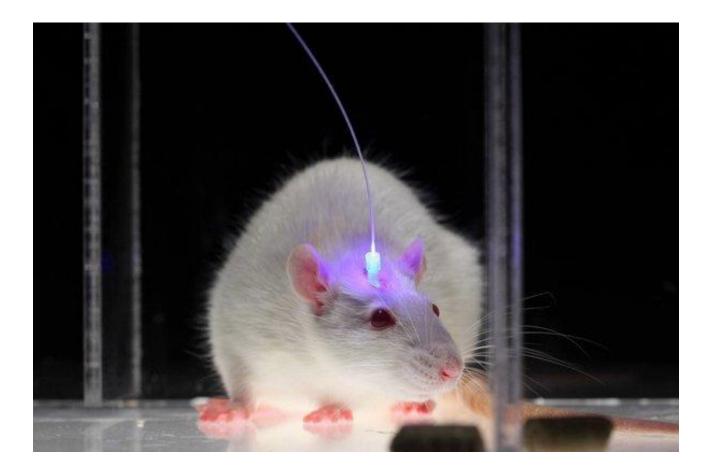


#### Generation of action potential by light pulse

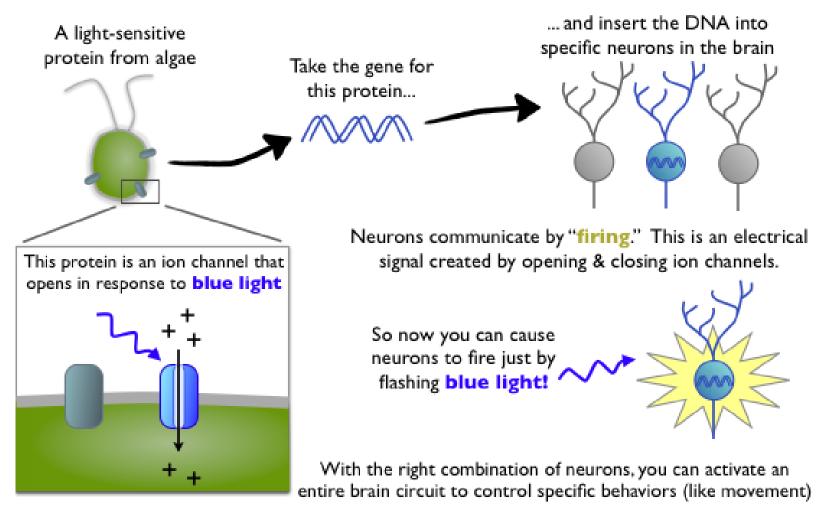


The blue-light sensitive Channelrhodopsin and the yellow light-activated chloride pump halorhodopsin together enable activation and silencing of neural activity





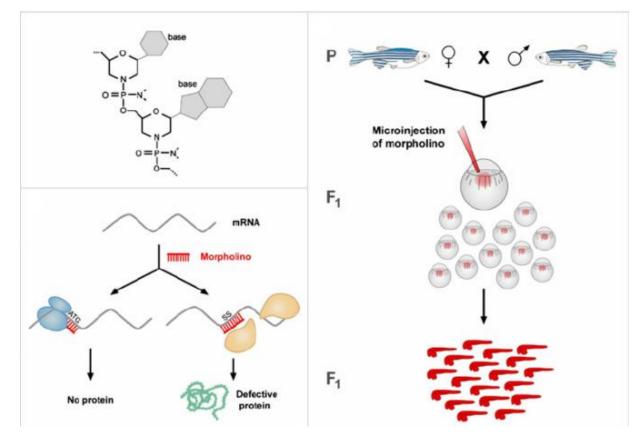
## **How optogenetics works**



https://www.youtube.com/watch?v=I64X7vHSHOE https://www.youtube.com/watch?v=rfEKc\_0iaJo https://www.youtube.com/watch?v=IW4j8\_k8pmE

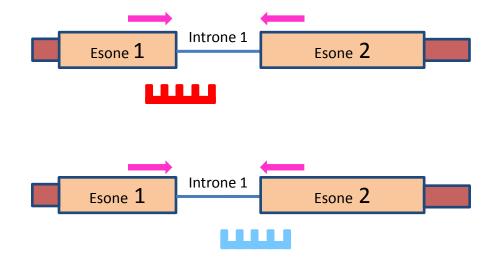
## Gene <u>knockdown</u> for studying gene function

Using morpholino-modified oligonucleotides (MO)

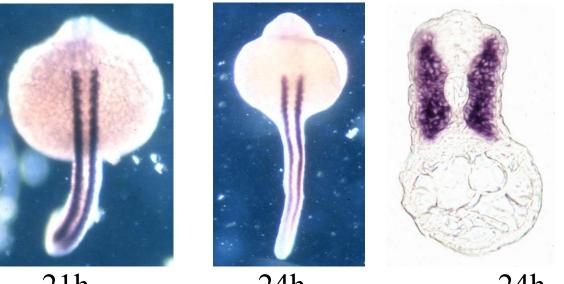


## Gene <u>knockdown</u> for studying gene function

Using morpholino-modified oligonucleotides (MO)



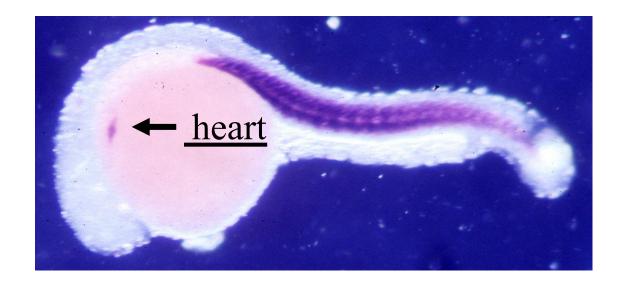
## Muscle-specific expression of Smyd1

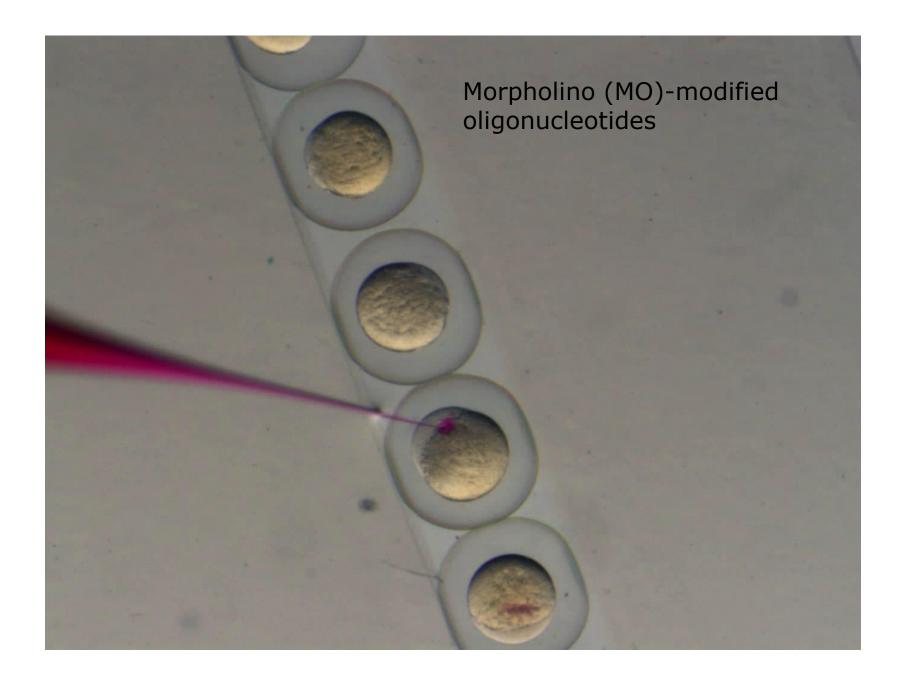


21h

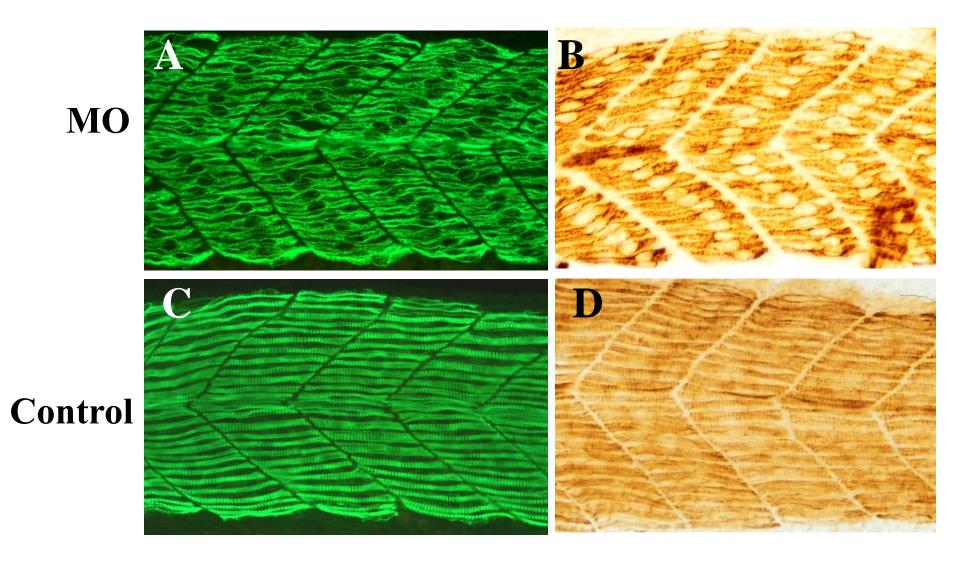
24h

24h



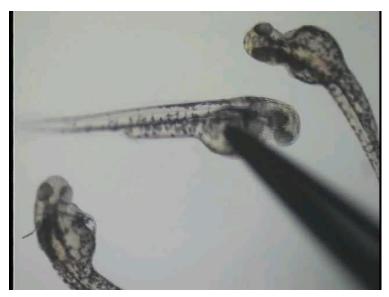


#### Knockdown of Smyd1 expression disrupts muscle cell differentiation

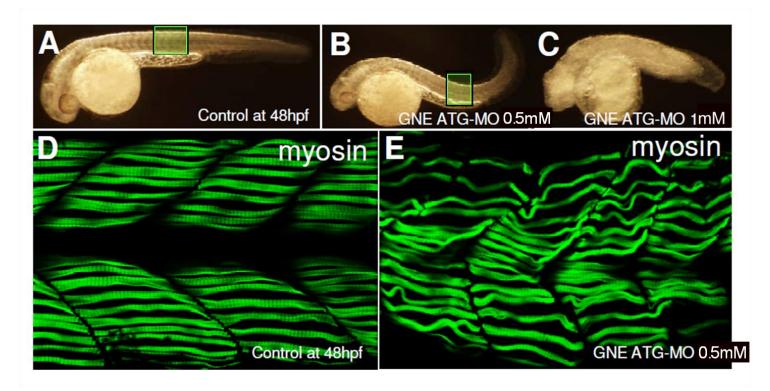


Knock down of Smyd1: Larvae are paralyzed and heart doesn't beat





## **GNE-mediated Hereditary Inclusion Body Myopathy**



GNE (UDP N-Acetylglucosamine 2-epimerase/N-Acetylmannosamine kinase)

Daya et al., 2014

# GNE MO knockdown phenotypes

## <u>1mM Splice Morpholino - 3dpf</u>



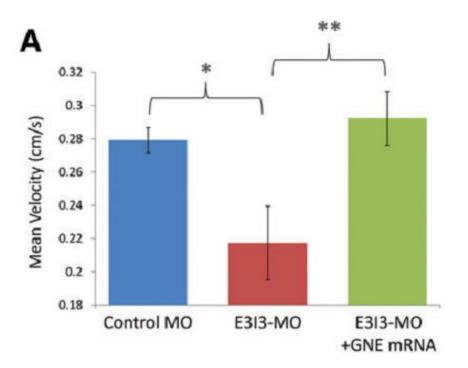
Control

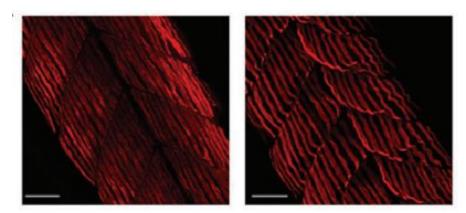
Mild

Intermediate

Severe

#### Larvae locomotor activity



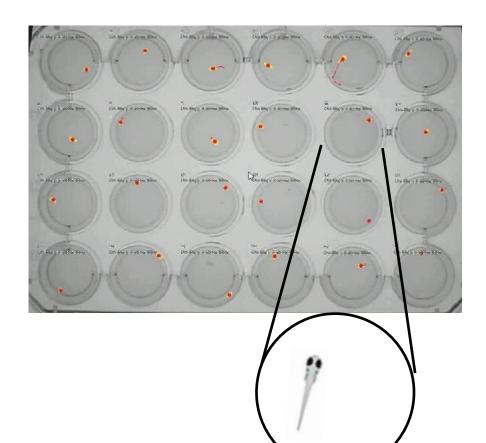


Immunostaining of 7 dpf larvae with slow muscle myosin antibody

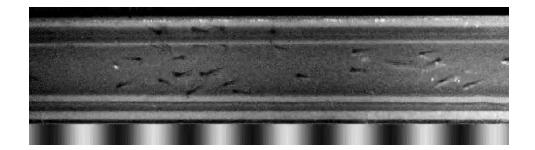
### Monitoring locomotor activity of zebrafish larvae



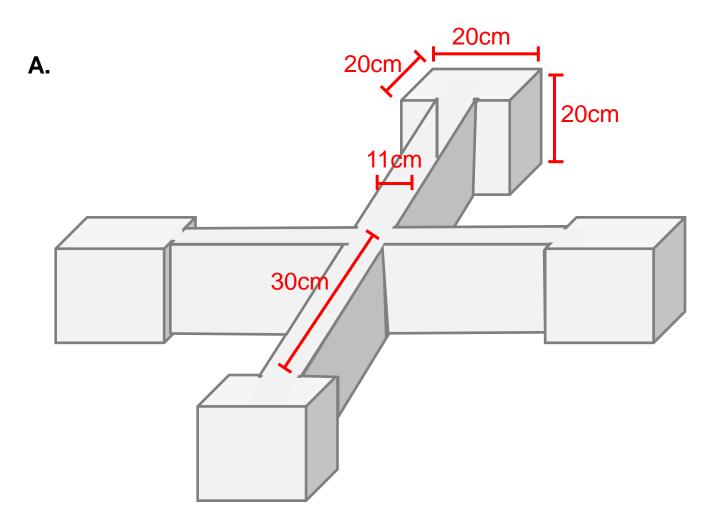
Danio Vision, Noldus



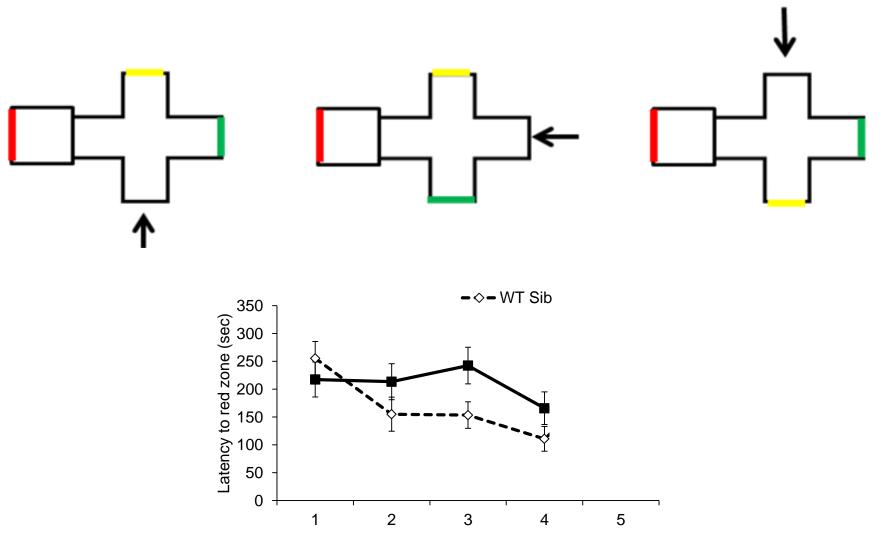
#### Vision test



# IQ test for fish

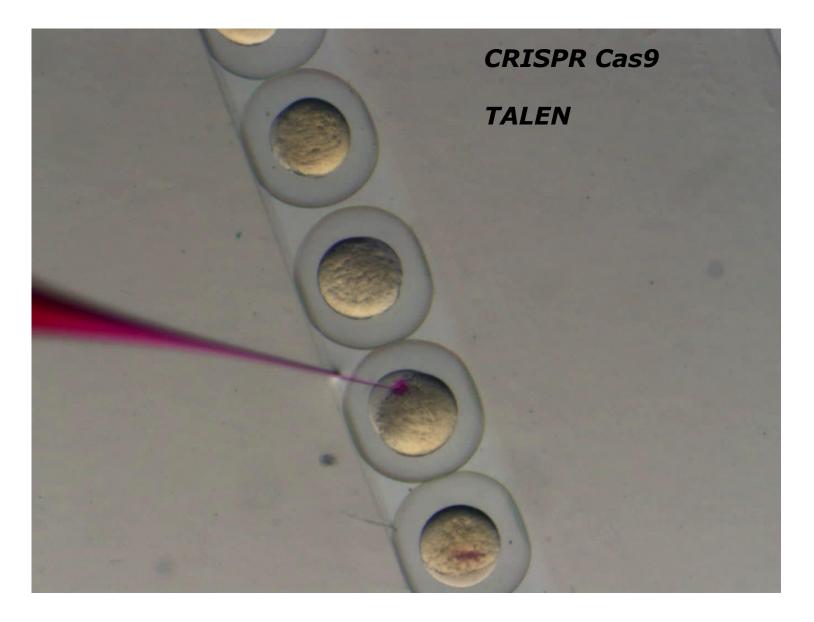


# IQ test for fish

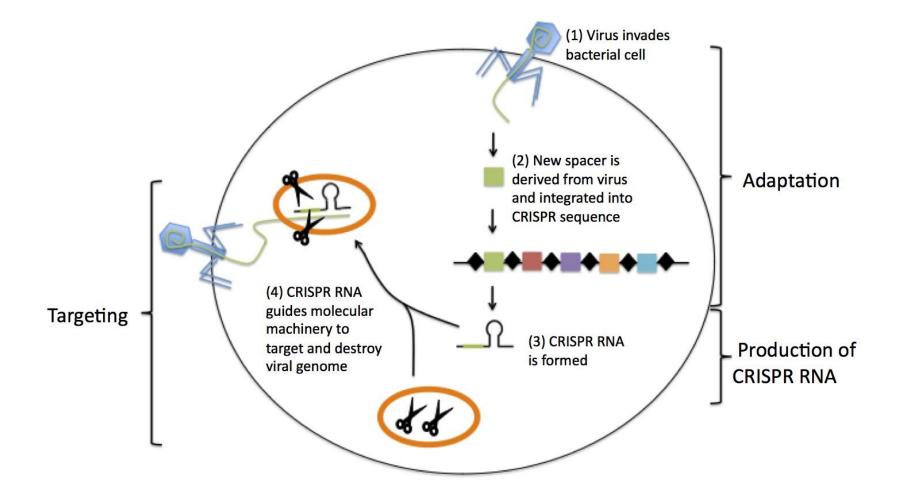


Trail

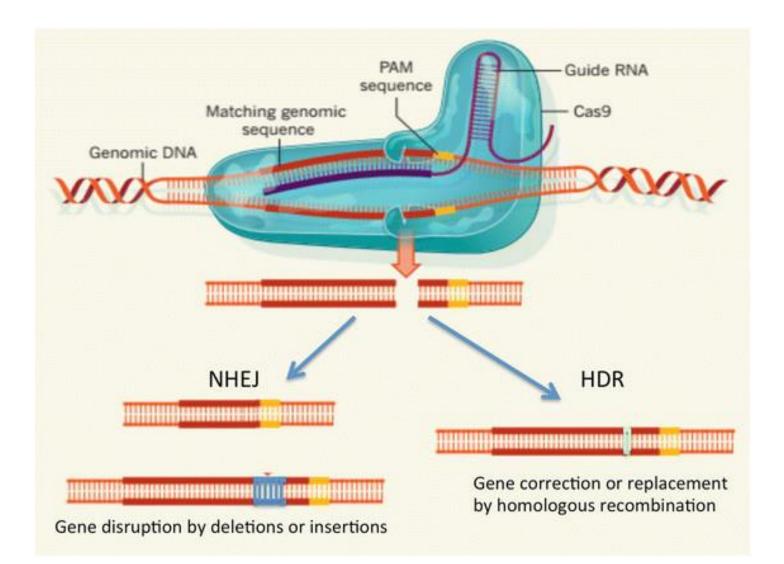
## Gene knockout for studying gene function

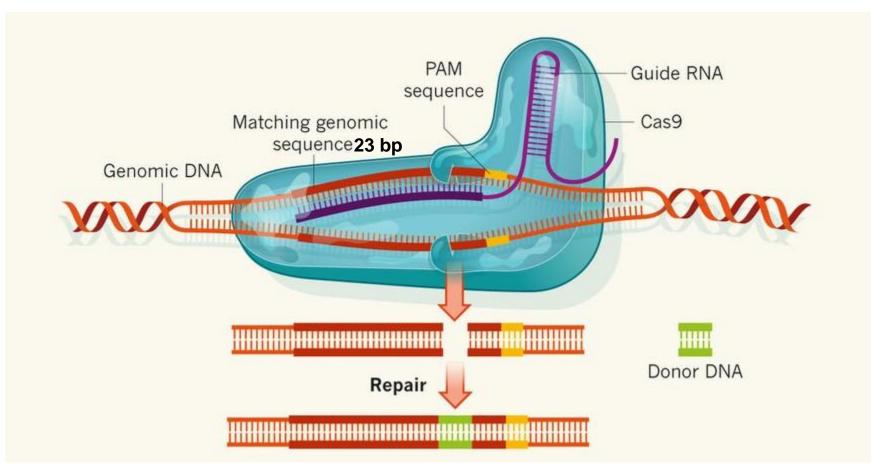


## clustered regularly interspaced short palindromic repeats (CRISPR)

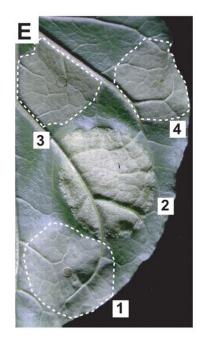


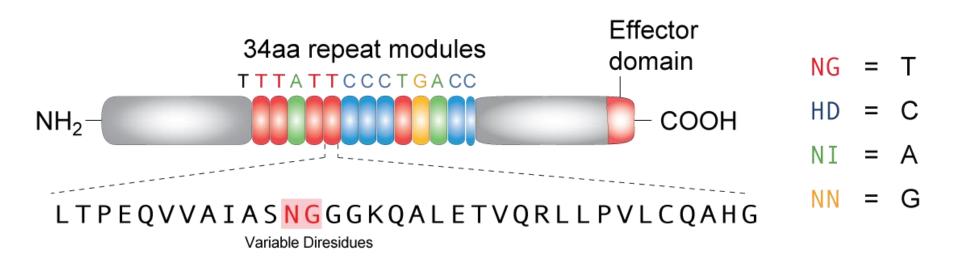
https://en.wikipedia.org/wiki/CRISPR



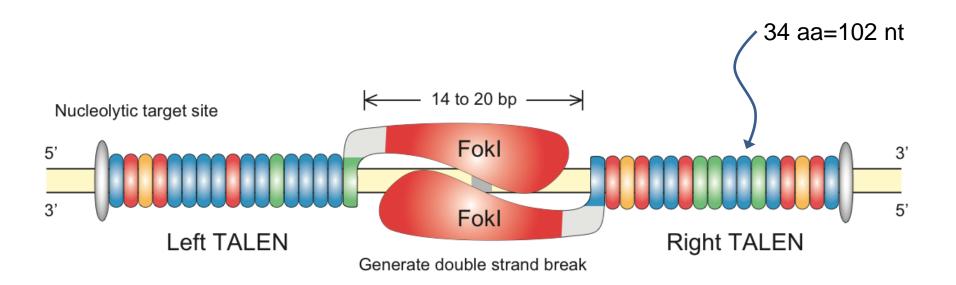


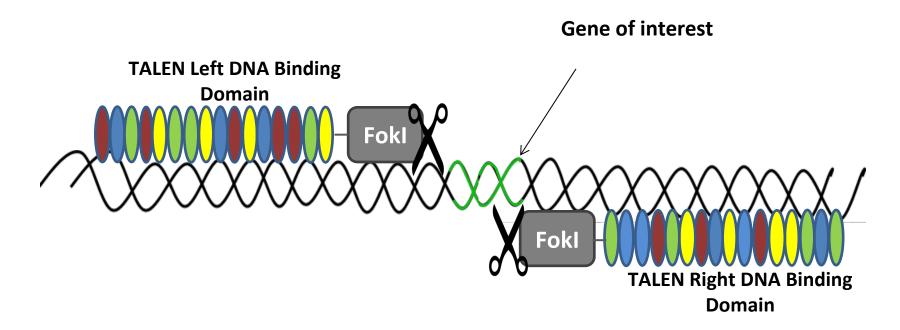
- Cell size modifications
- Transcription Activator-Like Effector - TALE



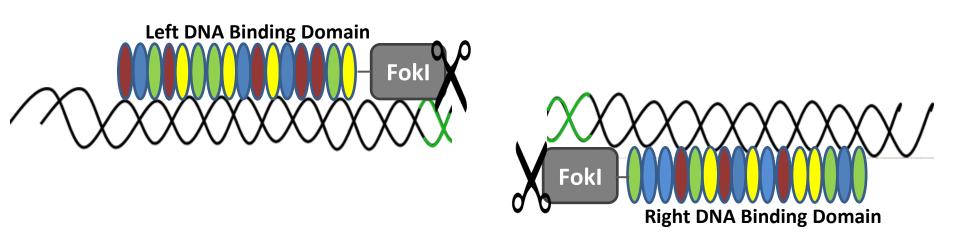


- TALE DNA-targeting domains contain 13-28 repeats of 34 aa each.
- Each repeat displays a different pair of residues at positions 12 & 13 (RVD) that associates preferentially with different nucleotides.
- H-histidine; D-aspartate; N-asparagine; G-glycine; I-Isoleucine

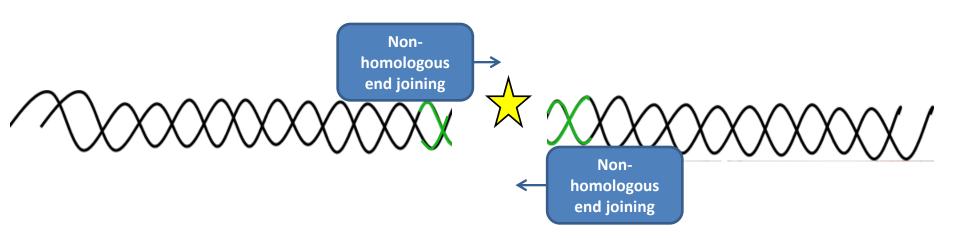




#### TALEN domains bind to a gene of interest and FOKI nucleases induce a double-stranded break.



#### TALEN domains bind to a gene of interest and FOKI nucleases induce a double-stranded break.

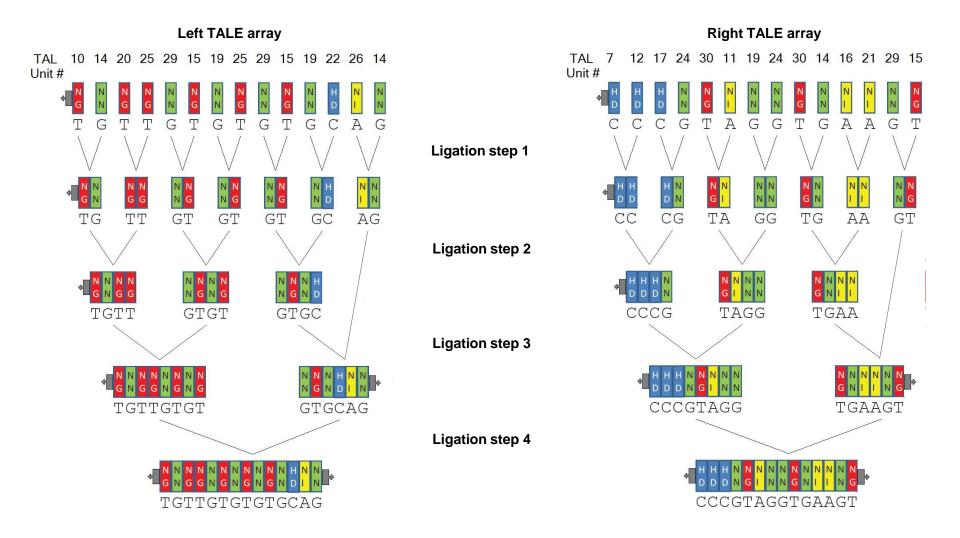


#### Non-homologous end joining (NHEJ) repair system introduces indels (insertions and/or deletion) into the gene sequence.

Mutated (inherited) gene sequence

#### Non-homologous end joining (NHEJ) repair system introduces indels (insertions and/or deletion) into the gene sequence.

## **Construction of TALE arrays**



- What are the advantages and disadvantages of zebrafish as a model for human genetic diseases?
- Whole genome duplication: What could be the consequence and why is this a consideration?

- What are the advantage of CRISPR over TALEN?
- What are the advantage of TALEN over CRISPR?
- Which of the two systems is mostly used in zebrafish research?