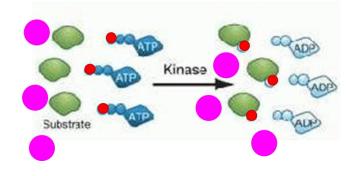
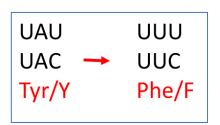


...the lecture will start soon...

Summary of the previous lesson

- kinase assay
- Phosphomymetic
- point mutation
- planning of an experiment to investigate the role the phosphorylation of a specific tyrosine in the interaction between two proteins





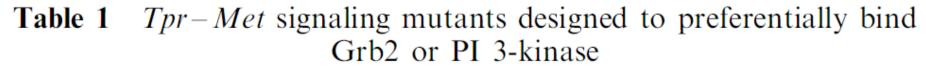


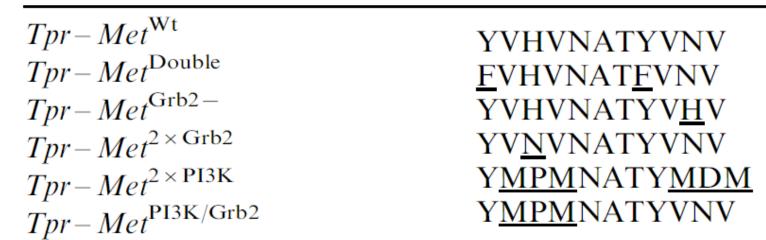
phospho-thyrosine

thyrosine

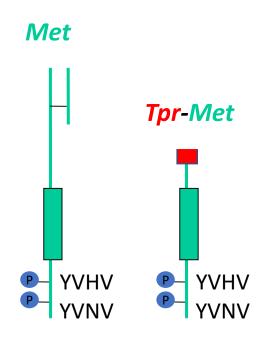
- identification of pathways involved in transformation and metastasis

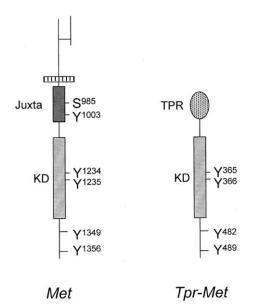
Docking site mutants



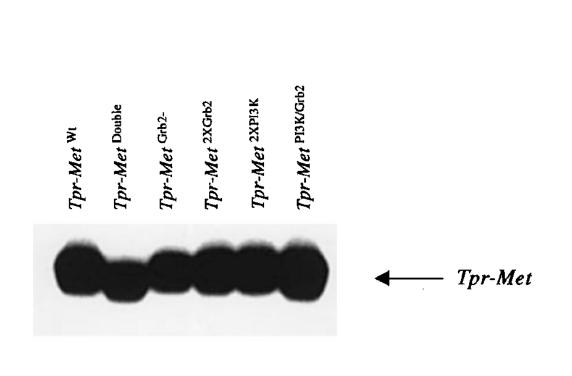


Signalling mutants of the oncogenic form of the Met receptor (Tpr-Met) were generated by site-directed mutagenesis. The consensus sequences for the SH2 domains of Grb2 and p85 (the regulatory subunit of PI 3-kinase) were designed according to Songyang et al. (1993). Mutagenized residues are underlined.



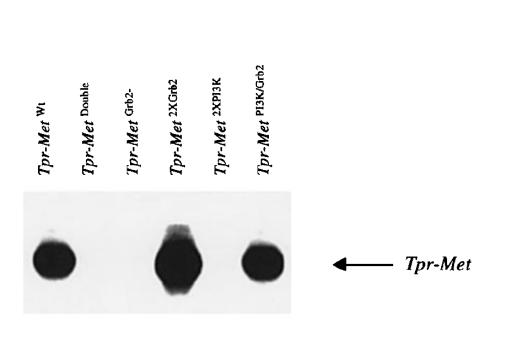


Kinase activity of Tpr-Met mutants



- transient trasfection of COS cells with different Tpr-Met constructs
- protein extraction
- IP anti Met
- kinase assay in vitro (with γ -32P ATP)
- SDS-PAGE
- autoradiography
- the bands are radioactive, because they are phosphorylated with radioactively labeled γ -32P ATP
- wild type and mutant Tpr Met proteins were immunoprecipitated from COS-1 cells transfected with the corresponding constructs, using antibodies specific for human *Met*
- immunoprecipitated proteins were subjected to in vitro kinase assay with $[\gamma^{-32}P]ATP$
- labeled proteins were separated on 8% SDS-PAGE

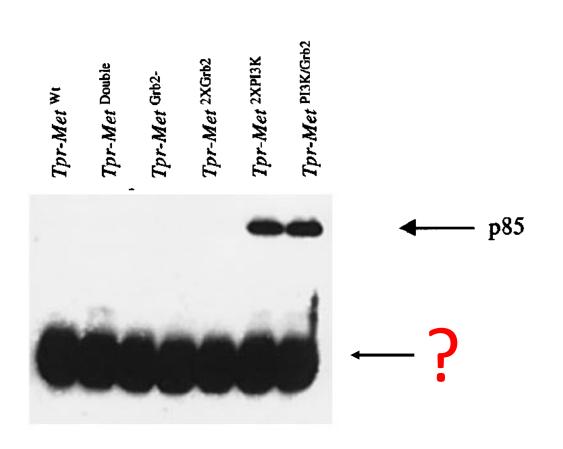
Association of Tpr-Met mutants with Grb2



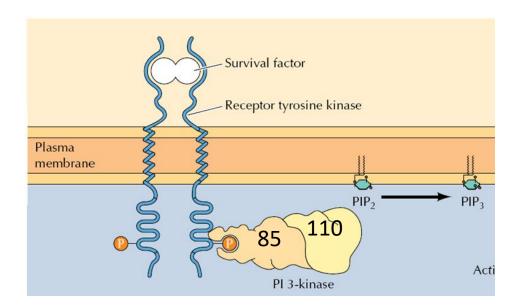
- fused protein: **Grb2-glutathione-transferase**, immobilized on **sepharose-glutathione beads**
- transient transfection of COS cells with various Tpr-Met constructs
- protein extraction
- "pull down": extracted proteins incubated together with the beads: some proteins bind to Grb2 and precipitate together with the beads
- kinase assay in vitro on precipitated proteins (with γ -32P ATP)
- SDS-PAGE
- X-ray film exposure
- bands are proteins precipitated together with Grb2 and are radioactive because they are phosphorylated with radioactively labeled ATP

Grb2 fusion protein (approximately 500 ng/point) was immobilized on Glutathione-Sepharose beads and incubated with lysates of COS-1 cells containing comparable amounts of Tpr-Met mutants. Complexes were washed and the amount of Tpr-Met bound to Grb2 was visualized by in vitro kinase assay with [[γ -32P]ATP. Labeled proteins were separated on 8% SDS± PAGE.

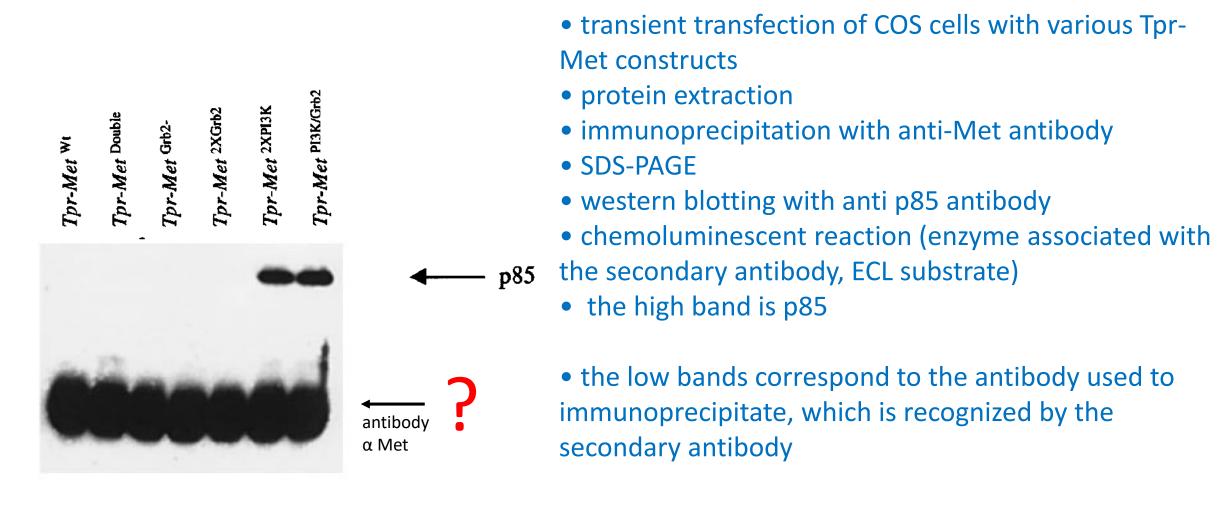
Association of Tpr-Met signaling mutants with PI 3-kinase



- transient transfection of COS cells with various Tpr-Met constructs
- protein extraction
- immunoprecipitation with anti-Met antibody
- SDS-PAGE
- western blotting with anti p85 antibody
- chemoluminescent reaction (enzyme associated with the secondary antibody, ECL substrate)
- the high band is p85

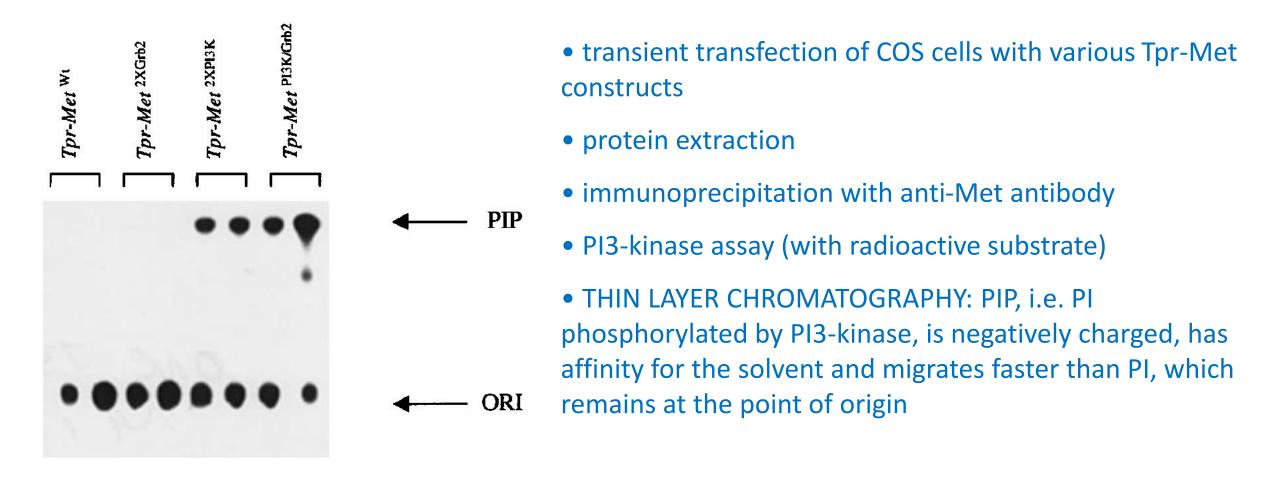


Association of Tpr-Met signaling mutants with PI 3-kinase



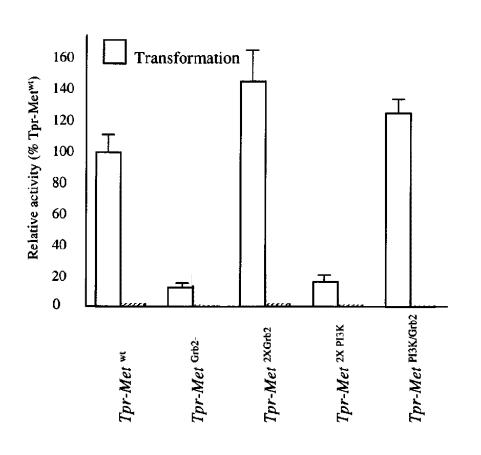
Tpr-Met mutants were immunoprecipitated with anti-Met antibodies from lysates of transfected COS-1 cells. Associated p85 was visualized by Western blotting with anti-p85 antibodies. Exposure was optimized to show the amount of p85 associated to the Tpr-Met ^{2xPI3K} and Tpr- Met ^{PI3K/Grb2} mutants. Upon longer exposure a faint p85 band can also be detected in the Tpr-Met Wt lane, as previously shown.

Association of Tpr-Met signaling mutants with Grb2 and PI3K



Tpr-Met mutants were immunoprecipitated with anti-Met antibodies from lysates of transfected COS-1 cells and the amount of Tpr-Met associated PI 3-kinase activity was determined by PI 3-kinase assay. The position of the phosphatidylinositol-3-phosphate (PIP) product of the PI 3-kinase reaction is indicated

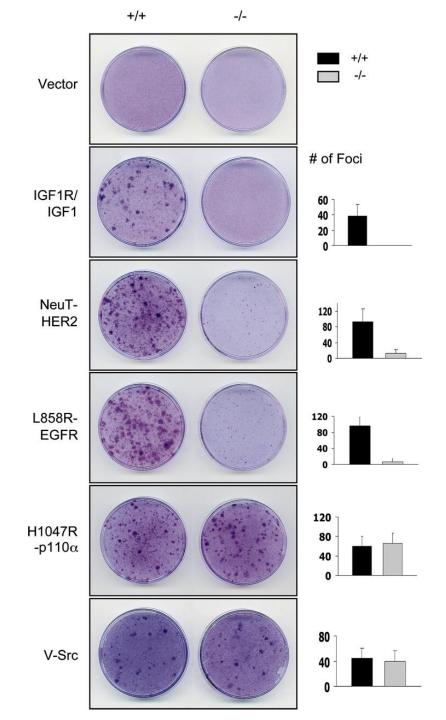
Grb2 coupling is necessary for transformation



FOCUS FORMING ASSAY

- transfection of Fisher rat fibroblast cells with
 10 µg of various Tpr-Met constructs
- count of the foci that are formed for each construct (focus forming assay)
- transformed cells forming foci were recovered for metastasis assay

The transforming potential of the indicated Tpr-Met mutants was evaluated by focus forming assay using Fisher rat fibroblasts. The values reported are expressed as percentage of the Tpr-Met^{Wt} transforming activity (150+12 foci/10 μ g of DNA) and represent the average of three independent experiments.

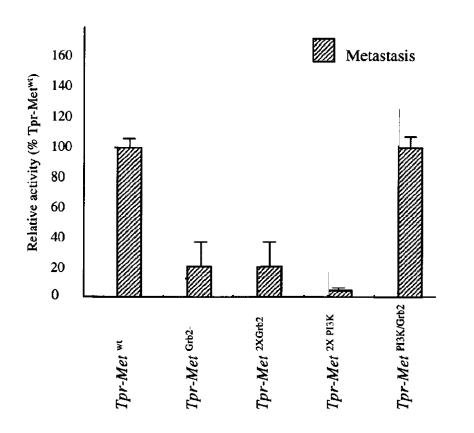


Example of focus forming assay

The p110 α isoform of PI3K is essential for proper growth factor signaling and oncogenic transformation

The ablation of p110 α impairs transformation induced by various oncogenic signals. Immortalized p110 α (+/+) and (-/-) MEFs were infected with a control empty-vector virus, a virus overexpressing wild-type IGF1R, or viruses carrying various oncogenes: mutant alleles of EGFR, L858R-EGFR, NeuT-HER2, or the tumor mutant allele of p110α, H1047R, as indicated. Cells were transfected with pSG5 expressing v-src. Cells were then cultured in the medium containing 5% FBS or containing 2% FBS but with addition of insulin (30 µg/ml) or IGF1 (50 ng/ml) in the case of cells infected with IGF1R. Foci were scored when wild-type MEFs were cultured for 3 weeks, p110αknockout (-/-) cells for 4.5 weeks.

The metastatic potential of Tpr-Met needs coupling to both Grb2 & PI3-kinase

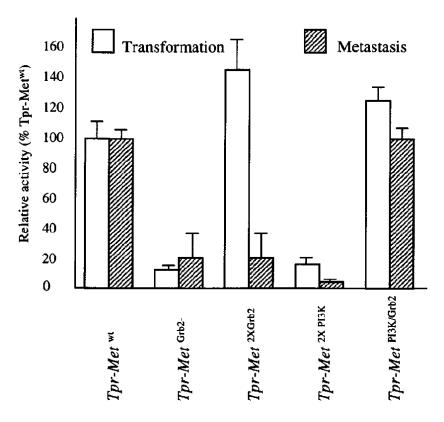


METASTASIS ASSAY

- recovered the transformed cells that form the foci
- injection of transformed cells into the tail vein
- after two weeks, evaluation of the number of mice that died from lung metastases

The metastatic ability of Tpr-Met signaling mutants was tested in an experimental metastasis assay. Bars indicate the percentage of mice dead because of lung metastases 2 weeks after injection into the tail vein of cells transformed by the various Tpr-Met signaling mutants. The values reported are expressed as percentage of the lethality caused by Tpr-Met^{Wt} and represent the average of three independent experiments (18 mice/group)

The metastatic potential of Tpr-Met needs coupling to both Grb2 & PI3-kinase

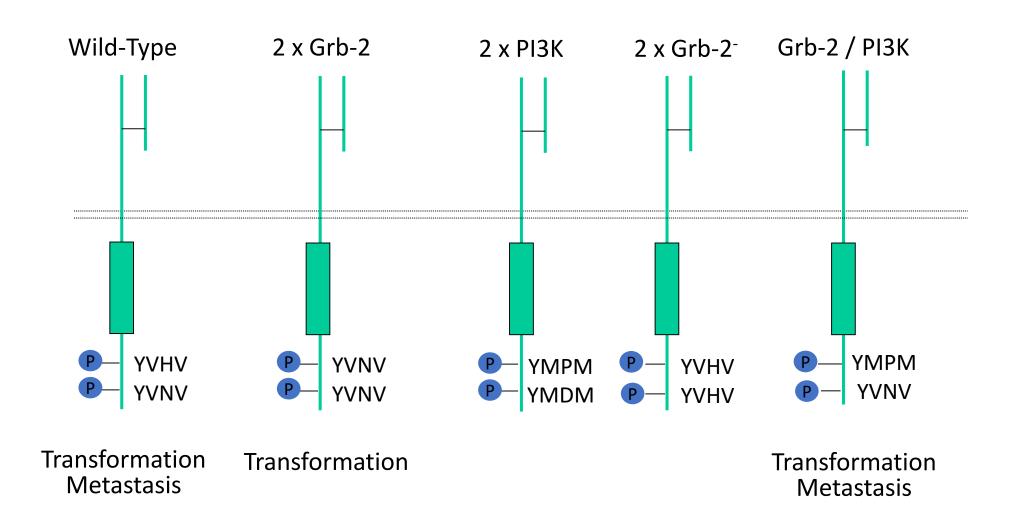


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Dissection of MET signalling pathways



Two examples of correct answers from the students:

4 plates are required

1-2: WT (IGF-1 -/+)

3-4: RNAi IRS1/IRS2 (IGF-1 -/+)

In total cell lysate: WB: p85, IGF1-R, IRS1/IRS2

In IP anti IGF1-R: WB: p85, IGF1-R

The more complete, containing all necessary information; it considers that all proteins are endogenously expressed in the cell. I added the WB for IGF1-R in the IP samples. A final comment on the possible results could be added.

- 4 plates: 2 transfected with p85 of PI3K and IGF-1R, -/+ IGF-1 treatment

2 transfected with p85 of PI3K and IGF-1R and IRS1/IRS2, -/+ IGF-1 treatment

- protein extraction
- IP against IGF-R1
- SDS-PAGE on total cell lysates and IP
- western blot for p85 PI3K and IGF-1R to verify if p85 PI3K interacts with IGF-1R and to investigate if they interact when IRS1/IRS2 is present.
- if they do not interact when IRS1/IRS2 is not present, it suggests that the presence of IRS1/IRS2 is necessary for the interaction between p85 PI3K and IGF-1R.

Another complete version, that considers that not all proteins are endogenously expressed in the cell. Nevertheless, it should be verified that IRS1/IRS2 are <u>really</u> not endogenously expressed. It contains a useful comment to the possible results.